

Supplementary information

Roadmap for alleviating the manifestations of ageing in the cardiovascular system

**In the format provided by
the authors and unedited**

Roadmap for alleviating manifestations of ageing in the cardiovascular system

Luca Liberale^{1,2,88}, Simon Tual-Chalot^{3,88†}, Simon Sedej^{4,5,6}, Stefano Ministrini⁷, Georgios Georgopoulos^{8,9}, Myriam Grunewald¹⁰, Magnus Bäck^{11,12}, Marie-Luce Bocheton-Piallat¹³, Reinier A. Boon¹⁴⁻¹⁶, Gustavo Campos Ramos¹⁷, Menno P.J. de Winther¹⁸, Konstantinos Drosatos¹⁹, Paul C. Evans²⁰, Jane F. Ferguson^{21,22}, Sofia K. Forslund-Startceva²³⁻²⁷, Claudia Goettsch²⁸, Mauro Giacca^{29,30}, Judith Haendeler³¹, Marinos Kallikourdis^{32,33}, Daniel F.J. Ketelhuth³⁴, Rory R. Koenen³⁵, Patrick Lacolley³⁶, Esther Lutgens³⁷, Pasquale Maffia³⁸⁻⁴⁰, Satomi Miwa³, Claudia Monaco⁴¹, Fabrizio Montecucco^{1,2}, Giuseppe Danilo Norata⁴², Elena Osto^{43,44}, Gavin D. Richardson³, Niels P. Riksen⁴⁵, Oliver Soehnlein⁴⁶, Ioakim Spyridopoulos⁴⁷, Sophie Van Linthout^{26,48}, Gemma Vilahur^{49,50}, Jolanda J. Wentzel⁵¹, Vicente Andrés⁵², Lina Badimon⁵³, Athanase Benetos⁵⁴, Christoph J. Binder⁵⁵, Ralf P. Brandes⁵⁶, Filippo Crea⁵⁷, David Furman⁵⁸⁻⁶⁰, Vera Gorbunova⁶¹, Tomasz J. Guzik⁶², Joseph A. Hill^{63,64}, Thomas F. Lüscher^{65,66}, María Mittelbrunn^{67,68}, Alessio Nencioni^{2,69}, Mihai G. Netea^{70,71}, João F. Passos⁷², Kimon Stamatelopoulos⁹, Nektarios Tavernarakis⁷³, Zoltan Ungvari^{74,75}, Joseph C. Wu⁷⁶, James L. Kirkland⁷⁷, Giovanni G. Camici^{7,78}, Stefanie Dimmeler^{15,16,79}, Guido Kroemer⁸⁰⁻⁸², Mahmoud Abdellatif^{4,80,81,83} & Konstantinos Stellos^{3,9,84-87†}

Authors affiliations

¹ First Clinic of Internal Medicine, Department of Internal Medicine, University of Genoa, Genoa, Italy.

² IRCCS Ospedale Policlinico San Martino Genoa – Italian Cardiovascular Network, Genoa, Italy.

³ Biosciences Institute, Vascular Biology and Medicine Theme, Faculty of Medical Sciences, Newcastle University, Newcastle Upon Tyne, UK.

⁴ Department of Cardiology, Medical University of Graz, Graz, Austria.

⁵ BioTechMed Graz, Graz 8010, Austria.

⁶ Institute of Physiology, Faculty of Medicine, University of Maribor, 2000 Maribor, Slovenia.

⁷ Center for Molecular Cardiology, University of Zurich, Schlieren, Switzerland.

⁸ Department of Physiology, School of Medicine, University of Patras, Patras, Greece.

⁹ Department of Clinical Therapeutics, School of Medicine, National and Kapodistrian University of Athens, Athens, Greece.

¹⁰ Faculty of Medicine, The Hebrew University of Jerusalem, Jerusalem, Israel.

¹¹ Translational Cardiology, Centre for Molecular Medicine, Department of Medicine Solna, and Department of Cardiology, Heart and Vascular Centre, Karolinska Institutet, Stockholm, Sweden.

¹² INSERM, University of Lorraine and Nancy University Hospital, Nancy, France.

¹³ Department of Pathology and Immunology, Faculty of Medicine, University of Geneva, Switzerland.

¹⁴ Department of Physiology, Amsterdam Cardiovascular Sciences, Amsterdam UMC location VUmc, Amsterdam, The Netherlands.

¹⁵ Institute for Cardiovascular Regeneration, Goethe University Frankfurt, Frankfurt, Germany.

¹⁶ German Centre for Cardiovascular Research (DZHK), Partner Site Rhine-Main, Frankfurt am Main, Germany.

¹⁷ Department of Internal Medicine I / Comprehensive Heart Failure Centre, University Hospital Würzburg, Würzburg, Germany.

¹⁸ Department of Medical Biochemistry, Amsterdam Cardiovascular Sciences: Atherosclerosis & Ischemic Syndromes; Amsterdam Infection and Immunity: Inflammatory diseases; Amsterdam UMC location AMC, Amsterdam, The Netherlands.

¹⁹ Metabolic Biology Laboratory, Cardiovascular Center, Department of Pharmacology, Physiology, and Neurobiology, University of Cincinnati College of Medicine, Cincinnati, OH, USA.

²⁰ William Harvey Research Institute, Barts and The London Faculty of Medicine and Dentistry, Queen Mary University of London, London, UK.

²¹ Division of Cardiovascular Medicine, Vanderbilt University Medical Center, Nashville, TN, USA.

²² Vanderbilt Microbiome Innovation Center, Nashville, TN, USA.

²³ Experimental and Clinical Research Center, Max Delbrück Center for Molecular Medicine and Charité-Universitätsmedizin Berlin, Berlin, Germany.

²⁴ Charité-Universitätsmedizin Berlin, Freie Universität Berlin and Humboldt-Universität zu Berlin, Berlin, Germany.

²⁵ Max Delbrück Center for Molecular Medicine in the Helmholtz Association (MDC), Berlin, Germany.

²⁶ German Center for Cardiovascular Research (DZHK), partner site Berlin, Berlin, Germany.

²⁷ Structural and Computational Biology Unit, EMBL, Heidelberg, Germany.

²⁸ Department of Internal Medicine I, Division of Cardiology, Medical Faculty, RWTH Aachen University, Aachen, Germany.

²⁹ King's College London, British Heart Foundation Centre of Research Excellence, London, UK.

³⁰ Department of Medical Sciences, University of Trieste, Italy.

³¹ Cardiovascular Degeneration, Medical Faculty, University Hospital and Heinrich-Heine University Düsseldorf, Germany.

³² Adaptive Immunity Lab, IRCCS Humanitas Research Hospital, Rozzano (Milan), Italy.

³³ Humanitas University, Pieve Emanuele (Milan), Italy.

³⁴ Cardiovascular and Renal Research Unit, Department of Molecular Medicine, University of Southern Denmark, Odense, Denmark.

³⁵ CARIM–School for Cardiovascular Diseases, Department of Biochemistry, Maastricht University, Maastricht. The Netherlands.

³⁶ Université de Lorraine, Inserm, DCAC, Nancy, France.

³⁷ Department of Cardiovascular Medicine & Immunology, Mayo Clinic, Rochester, MN, USA.

³⁸ School of Infection & Immunity, College of Medical, Veterinary and Life Sciences, University of Glasgow, Glasgow, UK.

³⁹ Department of Pharmacy, School of Medicine and Surgery, University of Naples Federico II, Naples, Italy.

⁴⁰ Africa-Europe CoRE in Non-Communicable Diseases & Multimorbidity, African Research Universities Alliance (ARUA) & The Guild of European Research-intensive Universities, Glasgow, United Kingdom.

⁴¹ Kennedy Institute, NDORMS, University of Oxford, Oxford, UK.

⁴² Department of Pharmacological and Biomolecular Sciences, Università degli Studi di Milano, Milan, Italy.

⁴³ Division of Physiology and Pathophysiology, Otto Loewi Research Center for Vascular Biology, Immunology and Inflammation, Medical University of Graz, Graz, Austria.

⁴⁴ Vetsuisse Faculty, University of Zurich Zurich, Switzerland.

⁴⁵ Radboud University Medical Center, Department of Internal Medicine, Nijmegen, The Netherlands.

⁴⁶ Institute of Experimental Pathology, University of Münster, Münster, Germany.

⁴⁷ Translational and Clinical Research Institute, Vascular Biology and Medicine Theme, Faculty of Medical Sciences, Newcastle University, Newcastle Upon Tyne, UK.

⁴⁸ Berlin Institute of Health (BIH) at Charité - Universitätmedizin Berlin, BIH Center for Regenerative Therapies (BCRT), Berlin, Germany.

⁴⁹ Research Institute, Hospital de la Santa Creu I Sant Pau, IIB-Sant Pau, Barcelona, Spain.

⁵⁰ CIBERCV, Barcelona, Spain.

⁵¹ Cardiology, Biomedical Engineering, Erasmus MC, Rotterdam, The Netherlands.

⁵² Centro Nacional de Investigaciones Cardiovasculares (CNIC), CIBERCV, Madrid, Spain.

⁵³ Cardiovascular Health and Innovation Research Foundation (FICSI) and Cardiovascular Health and Network Medicine Department, University of Vic (UVIC-UCC), Barcelona, Spain.

⁵⁴ Department of Geriatrics University Hospital of Nancy and Inserm DCAC, Université de Lorraine, Nancy, France.

⁵⁵ Department of Laboratory Medicine, Medical University of Vienna, Vienna, Austria.

⁵⁶ Institute for Cardiovascular Physiology, Goethe University, Frankfurt am Main, Germany.

⁵⁷ Centre of Excellence of Cardiovascular Sciences, Ospedale Isola Tiberina - Gemelli Isola, Roma, Italy.

⁵⁸ Buck Institute for Research on Aging, Novato, CA, USA.

⁵⁹ Stanford 1000 Immunomes Project, Stanford School of Medicine, Stanford, CA 94305 USA.

⁶⁰ IIMT, Universidad Austral, Consejo Nacional de Investigaciones Científicas y Técnicas, Pilar 1629, Argentina.

⁶¹ Departments of Biology and Medicine, University of Rochester, Rochester, NY, USA.

⁶² Centre for Cardiovascular Sciences, University of Edinburgh, Edinburgh, UK.

⁶³ University of Texas Southwestern Medical Center, Dallas, TX, USA.

⁶⁴ Moss Heart Center, University of Texas Southwestern Medical Center, Dallas, TX, USA.

⁶⁵ Heart Division, Royal Brompton and Harefield Hospital and National Heart and Lung Institute, Imperial College, London, UK.

⁶⁶ Cardiovascular Academic Group, King's College, London, UK.

⁶⁷ Consejo Superior de Investigaciones Científicas (CSIC), Centro de Biología Molecular Severo Ochoa (CSIC-UAM), Universidad Autónoma de Madrid (UAM), Madrid, Spain.

⁶⁸ Columbia Center for Translational Immunology. Department of Medicine, Columbia University, New York, NY, USA.

⁶⁹ Dipartimento di Medicina Interna e Specialità Mediche-DIMI, Università degli Studi di Genova, Genova, Italy.

⁷⁰ Department of Internal Medicine and Radboud Center for Infectious Diseases, Radboud University Medical Center, Nijmegen, The Netherlands.

⁷¹ Department of Immunology and Metabolism, Life and Medical Sciences Institute, University of Bonn, Bonn, Germany.

⁷² Department of Physiology and Biomedical Engineering, Robert and Arlene Kogod Center on Aging, Mayo Clinic, Rochester, MN, USA.

⁷³ Medical School, University of Crete, and Institute of Molecular Biology and Biotechnology, Foundation for Research and Technology-Hellas, Heraklion, Greece.

⁷⁴ Department of Neurosurgery, University of Oklahoma Health Sciences Center, Oklahoma City, OK, USA.

⁷⁵ International Training Program in Geroscience, Doctoral School of Basic and Translational Medicine/Department of Public Health, Semmelweis University, Budapest, Hungary.

⁷⁶ Stanford Cardiovascular Institute, Stanford University School of Medicine, Stanford, CA, USA.

⁷⁷ Center for Advanced Gerotherapeutics, Division of Endocrinology, Cedars-Sinai Medical Center, Los Angeles, CA, USA.

⁷⁸ Department of Research and Education, University Hospital Zurich, Zurich, Switzerland.

⁷⁹ Cardiopulmonary Institute Frankfurt, Frankfurt am Main, Germany.

⁸⁰ Centre de Recherche des Cordeliers, Equipe labellisée par la Ligue contre le cancer, Université Paris Cité, Sorbonne Université, Inserm U1138, Institut Universitaire de France, Paris, France.

⁸¹ Metabolomics and Cell Biology Platforms, Institut Gustave Roussy, Villejuif, France.

⁸² Institut du Cancer Paris CARPEM, Department of Biology, Hôpital Européen Georges Pompidou, AP-HP, Paris, France.

⁸³ BioTechMed-Graz, Graz, Austria.

⁸⁴ Department of Cardiovascular Research, Medical Faculty Mannheim, Heidelberg University, Mannheim, Germany.

⁸⁵ German Centre for Cardiovascular Research (DZHK), Partner Site Heidelberg/Mannheim, Mannheim, Germany.

⁸⁶ Helmholtz Institute for Translational AngioCardioScience (HI-TAC), Mannheim, Germany.

⁸⁷ Department of Medicine, University Medical Centre Mannheim, Heidelberg University, Mannheim, Germany.

⁸⁸ These authors contributed equally: Luca Liberale; Simon Tual-Chalot

[†] e-mail: simon.tual-chalot@newcastle.ac.uk; konstantinos.stellos@medma.uni-heidelberg.de

Funding sources

L.L. was supported by the Italian Ministry of Health (Ricerca Corrente 2022-2024). This work was supported by the Italian Ministry of Health – 5 x 1000. This article is based upon work supported also by the COST Action EU-METAHEART (CA22169) supported by COST (European Cooperation in Science and Technology). S.T-C is supported by the British Heart Foundation (PG/23/11093) and the Royal Society (RG/R1/241197). S.S. acknowledges funding received from BioTechMed-Graz (Flagship Project INTERACD⁺) and the Medical University of Graz (Flagship Project VASC-HEALTH). M.Gr. is supported by the Israel Science Foundation grant (1632/22). M.B. is supported by the Swedish Research Council (grant number 2023-02652), Hjärt-Lungfonden (grant number 20210560), OmegaPerMed (funded by Agence Nationale de la Recherche and VINNOVA under the frame of ERA PerMed), and CARE-IN-HEALTH (funded by the European Union within HORIZON EUROPE Health Framework Programme under grant agreement 101095413). M-L.B-P. is supported by the Swiss National Science Foundation (10.000.969). R.A.B. is supported by the European Union (ERC, project number 101002599), the Dutch Heart Foundation (ReGenLnc), the European Union (Horizon 2020, Grant No. 825670), and the Deutsche Forschungsgemeinschaft (TRR267). G.C.R. is supported by the Deutsche Forschungsgemeinschaft (DFG grant numbers 517001338 and 453989101). M.P.J.dW. is supported by the Netherlands Heart Foundation (CVON GENIUS II: 2017-20); Amsterdam UMC; Amsterdam Cardiovascular Sciences; ZonMW (Open competition 09120011910025), the European Union (Marie-Curie DN: MIRACLE) and the Epi-Guide-Edit project (KIC1.ST01.20.045) of the research programme KIC Key Technologies 2020 which is partly financed by the Dutch Research Council (NWO). K.D. is supported by the National Heart, Lung, and Blood Institute (HL151924). S.K.F-S is supported by the Deutsches Zentrum für Herz-Kreislauf-Forschung (DZHK, "XCVD" Grant No. 81Z0100113), the European Union (Horizon 2020, "IMMEDIATE" Grant No. 101095540), and the Deutsche Forschungsgemeinschaft (DFG, "HFpEF" SFB1470). C.G. is supported by the Deutsche Forschungsgemeinschaft (German Research Foundation, Transregional Collaborative Research Centre TRR 219; project ID 322900939, project C02). M.Gi. is supported by the European Research Council (ERC) Advanced Grant 787971 "CuRE"; British Heart Foundation (BHF) Programme Grant RG/19/11/34633; grants 825670 "CardioReGenix" and 874764 "REANIMA" from the European Commission Horizon 2020 programme; grant 20CVD04 from Fondation Leducq. J.Ha. is supported by DFG funding - CRC1116 and HA2868/14-1. M.K. is supported by AIRC (IG_24988 and 5x1000_22757) and the Italian Ministry of Universities and Research (PRIN 2020L45ZW4 and 2022SLL3YZ). D.F.J.K. research is supported by grants from the Novo Nordisk Foundation (0064142; 0075258); CARE-IN-HEALTH (funded by the European Union within HORIZON EUROPE Health Framework Programme under grant agreement 101095413); Independent Research Fund Denmark (2034-00136B); Simon Fougner Hartmanns Familiefond (2023-0066); and the University of Southern Denmark. R.R.K. is supported by the collaborative grant "MegaCardiocyte" from the Dutch and British Heart Foundations and the (DZHK) Deutsches Zentrum für Herz-Kreislauf-

Forschung and a project grant from the Netherlands Thrombosis Foundation (2021_01). P.L. is supported by the Investments for the Future program under grant agreement No ANR-15-RHU-0004, the Agence Nationale de la Recherche (ANR-13-BSV1-0026), and the “Fédération Française de Cardiologie”. E.L. is supported by the LeDucq Checkpoint Athero international network of excellence. P.M. is supported by British Heart Foundation grants (PG/19/84/34771, FS/19/56/34893A, PG/21/10541, PG/21/10557, PG/21/10634), FRA 2020 - Linea A, University of Naples Federico II/Compagnia di San Paolo, and the Italian Ministry of University and Research (MUR) PRIN 2022 (2022T45AXH). S.Miw. is supported by the European Commission Twinning Project to Universities of Coimbra (Portugal), Diabetes European Foundation for the Study of Diabetes grant, UK SPINE Translational Grant. C.M. is supported by EU H2020 H2020-SC1-2016-2017-TAXINOMISIS, EU PROJECT 797788 STRIKING STREAKS-Marie Skłodowska-Curie Individual European Fellowship (C.M.), EU FP7-HEALTH-F2-2013-602222-Athero-Flux, EU FP7-HEALTH-F2-2013-602114-Athero-B-Cell, the Novo Nordisk Foundation (NNF15CC0018346 and NNF0064142), the Kennedy Trust for Rheumatology Research (KTRR; KENN161701, KENN202101 and KENN192004), the British Heart Foundation (FS/18/63/34184B and RG/F/23/110105), Leducq Foundation for Cardiovascular research (Research Grant n° 22CVD02), and the Oxford NIHR Biomedical Research Centre. F.M. work is supported by #NEXTGENERATIONEU (NGEU) and funded by the Ministry of University and Research (MUR), National Recovery and Resilience Plan (NRRP), project MNESYS (PE00000006) - (DN. 1553 11.10.2022). G.D.N. is supported by Progetti di Rilevante Interesse Nazionale [PRIN 2022 7KTSAT], Ricerca Finalizzata, Ministry of Health [RF-2019-12370896], PNRR Missione 4, [Progetto CN3-National Center for Gene Therapy and Drugs based on RNA Technology], PNRR Missione 4 [Progetto MUSA-Multilayered Urban Sustainability Action], PNRR Missione 6 [PNRR-MAD-2022-12375913], European Commission [EUROPEAID/173691/DD/ACT/XK Nanokos]. E.O. is supported by the Swiss National Science Foundation, grant number PRIMA: PR00P3-179861/1, the Heubergstiftung, The Philhuman Stiftung and the Swiss Heart Foundation, Switzerland. N.P.R. is supported by a CVON grant from the Dutch Heart Foundation and Dutch Cardiovascular Alliance (CVON2018-27), and a Matching grant from the Dutch Heart Foundation (01-003-2021-03462021. NPR was further supported by a Project Program Grant of the NHLBI (Project 15-0893 and NIH/NHLBI P01HL131478). O.S. is supported by DFG TRR332 A2 & Z1, CRC1123 A6, CRC1009 B13, KFO342 P1, IZKF and IMF of the Medical Faculty Münster, Novo Nordisk, the Leducq Foundation, EU PRAETORIAN. S.V.L. is supported by the Deutsche Forschungsgemeinschaft (DFG, German Research Foundation) – SFB-1470 (SFB-1470-A07) and Project 536819681 – and by the Deutsche Krebshilfe (Project 70115119). G.V. is supported by Grant PID2021-128891OB-I00 and PLEC2021-007664- NextGenerationEU funded by MCIN/AEI/10.13039/501100011033 and Fondo Europeo de Desarrollo Regional (FEDER) A way of making Europe; and the SEC/FEC-INV-TRL 20/015 funded by the Spanish Society of Cardiology, and CERCA programme/Generalitat de Cataluña. V.A. is supported by the Ministerio de Ciencia, Innovación y Universidades (MICIU) and Agencia

Estatal de Investigación (AEI) (grant PID2022-141211OB-I00 funded by MICIU/AEI /10.13039/501100011033 and ERDF/EU), and the European Joint Program Rare Diseases Joint Transnational Call (EJPRD22-049) and Instituto de Salud Carlos III (ISCIII) (AC22/00020). The CNIC is supported by the MICIU, the ISCIII, and the Pro-CNIC Foundation and is a Severo Ochoa Center of Excellence (grant CEX2020-001041-S funded by MICIU/AEI/10.13039/501100011033).38. LB is supported by Spanish Ministry of Science and Innovation and Agencia Estatal de Investigacion (AEI) MICIN/AEI/ 10.13039/501100011033-[PID2019-107160RB-I00] with co-funding from the European Social Fund; by the National Institute of Health Carlos III (ISCIII) grant PMP22/00108 with Next Generation EU funds from the Recovery and Resilience Mechanism (RRM) Program, Red RICORS TERAV- RD21/0017/0013 and CIBERCV - CB16/11/00411, cofounded by FEDER "Una Manera de Hacer Europa"; SGR2021 grant from Generalitat de Catalunya and ERA-CVD Joint Transnational Call 2020 and H2020-JTI-IMI2-2017IMI-CARDIATEAM – 821508. A.B. is supported by FHU CARTAGE-PROFILES. C.B. is supported by Leducq Foundation (Transatlantic Network of Excellence; TNE-20CVD03) and the EU Horizon Europe (TillT; 101080897). R.P.B. is supported by DFG SFB1531 (456687919). V.G. is supported by NIA, MWRF, Michael Antonov Foundation, Impetus Grants. J.Hi. work was supported by grants from the NIH: HL-128215, HL-147933, HL-155765, HL-164586, and S10RR023729. T.F.L. is supported by Swiss National Science Foundation, Swiss Heart Foundation, Swiss Heart Foundation and Foundation for Cardiovascular Research – Zurich Heart House, Zurich and Bern Switherland. M.M. is supported by Grant PID2022-141169OB-I00 funded by MICIU/AEI/ 10.13039/501100011033 and, by “ERDF A way of making Europe”, by Grant ERC-2021-CoG 715322-LetTBe from European Research Council, and the Y2020/BIO-6350 NutriSION-CM Synergy Grant from Comunidad de Madrid. M.G.N is supported by an ERC Advanced Grant (833247) and Spinoza Grant oft the Netherlands Organization for Scientific Research. J.F.P. is funded by NIH grants R01AG068048; UH3CA268103, R01AG082708, P01 AG062413 and The Glenn Foundation For Medical Research. N.T. is supported by a grant from European Union Horizon Europe program (Excellence Hubs - HORIZON-WIDERA-2022-ACCESS-04-01) “CHAngeing – Connected Hubs in Ageing: Healthy Living to Protect Cerebrovascular Function” funded under the grant agreement No. 101087071, and the Hellenic Foundation for Research and Innovation (grant No. HFRI—FM17C3-0869, NeuroMitophagy). Z.U. is supported by National Institute on Aging (RF1AG072295, R01AG055395, R01AG068295), TKP2021-NKTA-47, implemented with the support provided by the Ministry of Innovation and Technology of Hungary from the National Research, Development and Innovation Fund, the National Cardiovascular Laboratory Program (RRF-2.3.1-21-2022-00003) provided by the Ministry of Innovation and Technology of Hungary from the National Research, Development and Innovation Fund. J.C.W. is supported by National Institutes of Health grants R01 HL113006, R01 HL130020, and R01 HL171102. J.L.K. is supported by National Institutes of Health grants R33AG061456 and R37AG013925, the Connor Fund, Robert J. and Theresa W. Ryan, and the Noaber Foundation. G.G.C. is supported by Swiss National Science Foundation, Swiss Heart

Foundation, Swiss Heart Foundation and Foundation for Cardiovascular Research – Zurich Heart House, Zurich and Bern Switzerland. S.D. is supported by EU ERC-2021-ADG, GAP - 101053352, Neuroheart and the FOR 5643 HERZBLUT DI 600/12-1 G.K. is supported by the Ligue contre le Cancer (équipe labellisée); Agence National de la Recherche (ANR-22-CE14-0066 VIVORUSH, ANR-23-CE44-0030 COPPERMAC, ANR-23-R4HC-0006 Ener-LIGHT); Association pour la recherche sur le cancer (ARC); Cancéropôle Ile-de-France; Fondation pour la Recherche Médicale (FRM); a donation by Elior; European Joint Programme on Rare Diseases (EJPRD) Wilsonmed; European Research Council Advanced Investigator Award (ERC-2021-ADG, Grant No. 101052444; project acronym: ICD-Cancer, project title: Immunogenic cell death (ICD) in the cancer-immune dialogue); The ERA4 Health Cardinoff Grant Ener-LIGHT; European Union Horizon 2020 research and innovation programmes Oncobiome (grant agreement number: 825410, Project Acronym: ONCOBIOME, Project title: Gut OncoMicrobiome Signatures [GOMS] associated with cancer incidence, prognosis and prediction of treatment response, Prevalung (grant agreement number 101095604, Project Acronym: PREVALUNG EU, project title: Biomarkers affecting the transition from cardiovascular disease to lung cancer: towards stratified interception), Neutrocure (grant agreement number 861878 : Project Acronym: Neutrocure ; project title: Development of “smart” amplifiers of reactive oxygen species specific to aberrant polymorphonuclear neutrophils for treatment of inflammatory and autoimmune diseases, cancer and myeloablation); National support managed by the Agence Nationale de la Recherche under the France 2030 programme (reference number 21-ESRE-0028, ESR/Equipex+ Onco-Pheno-Screen); Hevolution Network on Senescence in Aging (reference HF-E Einstein Network); Institut National du Cancer (INCa); Institut Universitaire de France; LabEx Immuno-Oncology ANR-18-IDEX-0001; a Cancer Research ASPIRE Award from the Mark Foundation; PAIR-Obésité INCa_1873, the RHUs Immunolife and LUCA-pi (ANR-21-RHUS-0017 and ANR-23-RHUS-0010, both dedicated to France Relance 2030); Seerave Foundation; SIRIC Cancer Research and Personalized Medicine (CARPEM, SIRIC CARPEM INCa-DGOS-Inserm-ITMO Cancer_18006 supported by Institut National du Cancer, Ministère des Solidarités et de la Santé and INSERM). This study contributes to the IdEx Université de Paris Cité ANR-18-IDEX-0001. M.A. acknowledges support from the European Commission (H2020-MSCA-IF), Austrian Society of Cardiology (Präsidentenstipendium-ÖKG), Medical University of Graz (Start Fund), BioTechMed-Graz (Young Researcher Group) and the Austrian Science Fund (FWF; P34926 & I6931). MA and GK received funding from FWF and ANR (Ener-LIGHT consortium) under the umbrella of the Partnership Fostering a European Research Area for Health (ERA4Health) (GA N° 101095426 of the EU Horizon Europe Research and Innovation Programme. K.Ste. is supported by grants from the European Research Council (ERC) under the European Union's Horizon 2020 research and innovation programme (MODVASC, grant agreement No 759248), the German Research Foundation DFG (CRC1366 C07, project number 394046768), the Health+Life Science Alliance Heidelberg Mannheim GmbH and the Helmholtz Institute for Translational AngioCardioScience (HI-TAC).

Supplementary Table 1: Ageing hallmarks in cells of the circulatory system

	Genomic Instability	Telomere attrition	Epigenetic alteration	Loss of proteostasis	Disabled macroautophagy	Altered intercellular communication	Chronic inflammation	Dysbiosis	Deregulated nutrient-sensing	Mitochondrial dysfunction	Cellular senescence	Stem cell exhaustion
Vascular cells												
Endothelial cells	1,2	3	4,5,6,7,8	9	9,10,11	12	13,14,15,16,1 7,18	19	20	21,22	23,24,25	15
Smooth muscle cells	1,2,26	1,27	4,28	29	30	31	32	33	34,35,36	21	37,38	39
Pericytes			40				40				40	
Cardiac cells												
Cardiomyocytes	41	41	41	42	43,44	41,45	46,47	48	49	41,50,51	41	
Cardiac fibroblasts	52		53			53	53		54,55	56	57	58
Circulatory cells												
Hematopoietic stem cells	59	60,61	60,62,63	64	65,66	67,68	17,69	69	70	71,72	60,73	74,75
Lymphoid lineage												
B cells		76,77	78	79	80	81	81	82,83,84	85	86	87	88
T cells	3	77,89	78,90	91-93	94,95	96,97	98,99,100		101,102	86,95,103, 104	105,106	88
Natural killer cells		107				108,109,110	111,112,113, 114	115	116-120		121,122, 123	
Myeloid lineage												

Monocytes	124	125	126	127	128	128,129,130	131,132		133,134	133,134	125,131	74
Macrophages	135	136,137	138	139	136,140	141,142	138,143	144,145, 146	147,148	86	149,150, 151	
Granulocytes	152	153	152		152,154	155,156,157	156	158	134,159	86,160	161	74
Platelets		162	163	164	165	166,167,168	169,170		171	169,172	163	173
Erythrocytes		174				175			176,177		178	
Dendritic cells	179,180			181		129	182,183	184	185	186	187	188
<hr/>												
Level of evidence	I,Human & other organisms	II,Human & other organisms	I,Only Human	II,Only Human	I,Only preclinical	II,Only preclinical	No effect	Unknown				
Preclinical evidence: aged model or accelerated model of ageing. Human evidence: aged model of cell or human tissue. Evidence are (1) hallmark should manifest or happen during normal ageing, (2) experimental acceleration or increase of the hallmark accelerates normal ageing, and (3) experimental reduction of the hallmark slows normal ageing. Evidence I has at least 2 of the 3 above-mentioned point, while evidence II has only 1 of the above-mentioned point. The numbers in the boxes refer to the corresponding citations in the bibliography.												

Supplementary Table 2: Geropromoting mechanisms in human disease

	Genomic Instability	Telomere attrition	Epigenetic alteration	Loss of proteostasis	Disabled macroautophagy	Altered intercellular communication	Chronic inflammation	Dysbiosis	Deregulated nutrient-sensing	Mitochondrial dysfunction	Cellular senescence	Stem cell exhaustion
Cardiovascular system												
Hypertension	189,190, 191	190	192	193	193,194,195,19 6,197,198	199,200,201, 202	203,204	205,206	206,207,208, 209	195,196,210, 211	2,190	
Atherosclerosis	191,212	213,212	192	193	30,193, 214	199,215,216	204,217,218	219,220	207,208	214,221	190	222
Aneurysm/Arterial Dissection	223		224	225	226	199,215,227	228	229	228	230	231	231
ATTR Amyloidosis				223								
Atrial Fibrillation	232,233		232	234,235	234	236,237,238, 239,240	237,241,242	243	244	245,246,247	248	249
Non-AF arrhythmias	250		251,252	253	254	255,256	241,255,256	257	258	247,259	231	231,249
Age-related Heart Failure with preserved Ejection Fraction	260,261, 262,191	263,262, 264	265,262, 266	267,268, 269	267,270,271,19 5,272,273	274	103,204		207,275	103,195,261, 276	41,261, 277	
Cardiac Valve Degeneration	191	278	279	191		279	103,280		207	103,281	282	283
Brain												
Vascular Dementia	284,285	286,287	288,289, 290	291,292, 293	291,292,294	295,296,297, 298,299	103,203,300, 301	302,303	207,275,304, 305	103,306,307	308,309, 310	311
Alzheimer's Disease	312,313	314,315	289,290, 316	317,318, 319	317	320,321,322, 323	324,325	302	326,327,328	329	330	331
Parkinson's Disease	332	315,333, 334	335	336	337	338,339	340,341	342	343	344	345	346
Stroke	347,348, 349	350	347,351	352	353	354,355	356,357	358,359	360	361	347	

Kidney												
Chronic Kidney Disease	260	362,363	288	364,253, 365	195,271,366	215,367,368	103,204	369	370	103,306,371	231,372	231
Lung												
COPD/Emphysema		373,374, 375	376	377	378	49,379,380	379,381	382	383,384,385	386,378	379,387	388,389
Idiopathic Pulmonary Fibrosis		362,390, 391,374, 375,392	393	394,395	267,396	397	398,399	400	383	401,402	398,403, 404	405
Ear, nose, and throat diseases												
Presbycusis	406,407, 408		408,409, 410	411	407	412	407		413,414	415,416,417, 418,419	406,407	
Macular Degeneration/ Retinal atrophy	260,285, 420	421	288,422	423,424, 425	426,423	427	426,428,429, 430	431	426,432,433	434	426,429	429,433
Cataracts	420,435, 212,213	212,213	288	436,437, 438	439,440	441	442,443		443	221,371	444	422
Liver												
Metabolic dysfunction-associated steatotic liver disease		445,446			447	447,448,449, 450	204	451,452	370	453,454,455, 456	457	458
Liver Fibrosis		362,446, 459,460	458	458	267	67,448,449,461	103,204	451	370	103,306,454, 455	462	458
Bone Marrow												
Myelodysplastic syndromes and Hematologic Neoplastic Disorders	420	463,464, 465,466	467,468	469	271	470,471	472	473	474,475,476	221,371,477		
Anemia	285	362,478, 479	480	481		482	103	483		103,484	485,285	485,285
Pancreas												
Diabetes	212,408	287,459, 212,213, 362	459,486	193	198,447	201,367,487, 488	204	452	207,208,489, 490	491,492	459	231

Systemic												
Osteoarthritis	212	213,362	288	493,494		495	103,495	369	207	103	444,496, 497,498	231
Osteoporosis	260,435, 408,420	287,213, 362,435	288,499	493,494	271	201,448,449, 500,501	103,502	369	207,369	103	444,501	458,501
Sarcopenia	260,435, 408,420	287,362, 435	288,499	493,494	267,271	201,448,449, 500	103,502	369	369	103	231,444	231,458
Infertility	435,212, 408,503	463,504, 212,505	499	493,494, 365	440	506,507	204	508	207	306	444	
Solid tumors	260,212, 285,420	463,212, 213,505	480,509, 510	511	271	449,512,513	217	514	207	371,455,515	459	
Graying/Loss of hair	212,408, 420,516	517,212, 213,505	499	364	267	215,449,518	103,502	369	207	103,306,371, 491	231	231,458
Skin alterations (tight skin, thin skin, hyperkeratosis)	212,408, 420,516	287,212, 213,505, 517	499	364,493, 494,253	267	67,215	103,502		207	103,371	231,498	231,458, 519
Light green: experimental evidence (genetic models, treatments) Dark green: clinical evidence (randomized clinical trials with drugs acting on ageing hallmarks, genetic syndromes with accelerated ageing (references are highlighted in red)). The numbers in the boxes refer to the corresponding citations in the bibliography.												

Supplementary Table 3: Lifestyle interventions as potential geroprotective and gerotherapeutic strategies for the cardiovascular system

Experimental approaches	Study design	Ageing hallmark affected	Effects on the circulatory system	Refs.
Caloric restriction and dietary interventions				
8-week treatment program	RCT, 43 healthy adult males; 50-72 years	Epigenetic	↓Triglycerides ↑5-methyltetrahydrofolate	⁵²⁰
For Phase 2 of the CALERIE™ study participants were randomized 2:1 to either a 25% CR diet or an ad libitum diet for 2 years.	199 healthy young-to-middle-aged individuals	Senescence	↓Circulating biomarkers of ageing ↑Marker of metabolic health	⁵²¹
2-year moderate CR (13%)	Multicentre, RCT, 238 healthy non-obese young and middle-aged; 21-50 years; men (72) and women (166)	↓ Oxidative stress ↓ Low-grade systemic inflammation ↑ Nitric oxide bioavailability	↓LDL-cholesterol, ↓Total cholesterol to HDL-cholesterol ratio ↓Blood pressure ↓CRP ↑Insulin sensitivity index ↓Metabolic syndrome severity score	⁵²²
6 months of 25% CR alone or in combination with exercise	RCT, 36 healthy non-obese men and women	↓ Inflammation	↓Body weight ↓Triacylglycerols CR+ exercise: ↓LDL-cholesterol ↓Diastolic blood pressure ↓ CRP in the controls versus CR+ exercise ↓ Estimated 10-year CVD risk by 29% in CR and 38% in the CR+ exercise (↔ control group)	⁵²³
8-week low-energy diet (810 kcal/daily)	Multicentre RCT, 2224 overweight individuals; 1504 women, 720 men with pre-diabetes	↓Inflammation	Men and Women: ↑Insulin sensitivity (35% of participants reverted to normoglycaemia) Men vs. Women: ↓Body weight ↓Metabolic syndrome Z-score ↓CRP ↓Fat mass ↓Heart rate Women vs. Men: ↓HDL cholesterol ↓Free-fat mass ↓Waist circumference ↓Pulse pressure	⁵²⁴
CR alone or in combination with exercise	RCT, 100 obese men and women with heart failure with preserved	↓ Inflammation	CR and exercise: ↑Peak oxygen consumption ↓LV hypertrophy ↓Diastolic dysfunction	⁵²⁵

	ejection fraction≥ 60 years		CR: ↓CRP ↓Total cholesterol ↓LDL-cholesterol	
12-months of CR (20%) or exercise	RCT, 48 normal weight and overweight middle-aged 29 women and 17men; 57 ± 3 years	↓Inflammation	↓Body (fat) mass ↓LDL-cholesterol ↓Total cholesterol/HDL ratio ↓HOMA-IR index ↓CRP	526
Fasting-mimicking diet (FMD): 3 cycles for 5 consecutive days per month for 3 months	RCT, 100 generally healthy participants (37 men; 42.2 ± 12.5 years and 63 women; 43.3 ± 11.7 years)	↑Nutrient signaling	↓Body mass index ↓Blood pressure ↓Fasting glucose ↓IGF-1 ↓Triglycerides ↓Total and low-density lipoprotein cholesterol ↓CRP	527
Long-term CR (11%) coupled with high intensity physical activity	Observational study; 54 healthy community-dwelling participants (29 men; 74.5 ± 0.7 years) and 25 women; 74.7 ± 0.6 years	↑Nutrient signaling	↓Coronary heart disease ↓Body mass index ↑Plasma dehydroepiandrosterone ↓Mortality from age-associated diseases	528
Long-term CR (for an average of 6 years)	Observational study; 15 men and 3 women (range 35–82 years).	↓Inflammation	↓Body weight ↓Fat mass ↓Blood pressure ↓Total cholesterol ↓LDL-cholesterol ↓Total cholesterol to HDL-cholesterol ratio ↓Triglycerides ↑Fasting glucose ↓Fasting insulin ↓CRP ↓Carotid artery intima media thickness	529
Exercise training				
Lifelong exercise	Cross-sectional/observational study; 102 healthy seniors (>64 years of age) stratified into 4 groups: sedentary, casual, regular, competitive	↑Myocardial energetics ↑Expression of genes involved in fatty acid oxidation ↓Low-grade systemic inflammation	Regular exercisers vs. sedentary subjects: ↑Peak oxygen uptake ↑LV hypertrophy ↑LV distensibility ↓LV stiffness ↑LV compliance	530
Treadmill exercise	Observational study, 122,007 patients (53.4 ± 12.6 years;	Unknown	Risk-adjusted all-cause mortality was inversely proportional to cardiorespiratory fitness and was lowest in elite performers.	531

	72 173 [59.2%] male)		Extremely high aerobic fitness was associated with benefit in older patients and those with hypertension.	
Cardiorespiratory fitness and mortality risk for different races, sex, and age	Observational study involving 750,302 U.S. veterans aged 30 to 95 years (mean age 61.3 ± 9.8 years) during a standardized exercise treadmill test	Unknown	↓Mortality by >50% across all age groups, races, and sex, independent of comorbidities upon moderate intensity physical exercise >150 minutes weekly.	⁵³²
Prolonged, sustained endurance training	Observational study, 12 healthy sedentary seniors (69.8 ± 3 years; 6 women, 6 men) and 12 Masters athletes (67.8 ± 3 years; 6 women, 6 men)	Unknown	↑Stroke volume ↔Contractility	⁵³³
4-year CR and physical activity	Multi-center, RCT, 5,145 volunteers (age 45-76 years), with established DM2 and overweight or obesity. Study follow-up: 11.5 years	Unknown	↔Risk of cardiovascular morbidity (heart attacks and stroke) or cardiovascular-related death	^{534,535}

Abbreviations: CR: caloric restriction; CRP: C-reactive protein; HOMA-IR: homeostatic model assessment for insulin resistance; LDL: low-density lipoproteins; LV: left ventricle; HDL: high-density lipoproteins; RCT: randomized controlled trial

Supplementary Table 4: Potential geroprotective and gerotherapeutic strategies on the cardiovascular system				
Experimental approaches	Study design	Ageing Hallmark affected	Effects on the circulatory system	Refs.
Current geroprotective and gerotherapeutic strategies used in human				
Senolytics				
Fisetin	RCT, fisetin two administrations of 2 mg/kg for three days – 2 weeks interval in between	Cell senescence	Endpoints: carotid femoral and brachial artery PWV. <u>Ongoing</u>	NCT06133634
Telomerase activators				
TA-65®	RCT, patients with MI >65 years old. TA-65® 16 mg/day, 12 months	Inflammation	↓ hsCRP (-62%), ↓ adverse events (-30%)	⁵³⁶
Anti-diabetic drugs				
Metformin	RCT, impaired glucose tolerance, metformin 850 mg bid, 3 years	Nutrient-sensing; gut microbiota	↓ risk of diabetes = risk of MACE	⁵³⁷
GLP-1RA	RCT, non-diabetic obese or overweight, semaglutide 2.4 mg/week, 3 years	Nutrient-sensing	↓ body weight ↓ risk of MACE	^{538,539}
mTOR inhibitor				
Everolimus	RCT, patients with STEMI undergoing PCI, everolimus 7.5 mg/day (days 1-3) + 5.0 mg/day (days 4-5)	Nutrient-sensing, autophagy, inflammation	No difference in MI size or microvascular obstruction 30 days after the event	⁵⁴⁰
Polyamines				
Bifidobacterium animalis + Arginine	RCT, healthy adults, dietary supplementation for 12 weeks,	Gut microbiota, autophagy (↑ production of spermidine)	↑ brachial artery FMD	⁵⁴¹
Sirtuins activators				
Nicotinamide mononucleotide (NMN)	RCT, healthy adults, nicotinamide mononucleotide 300, 600, or 900 mg/day vs placebo, 60 days	Epigenetic regulation; oxidative stress	↑ performance 6MWT = insulin-sensitivity	⁵⁴²
Resveratrol	Meta-analysis of RCTs, adults with metabolic syndrome, resveratrol 150-3000 mg/day, 1 week to 1 year	Epigenetic regulation; oxidative stress	↑ FMD	⁵⁴³
Anti-inflammatory				
Colchicine	RCT, patients with chronic coronary artery disease, 0.5 mg/day, 2.5 years	Inflammation	↓ risk of MACE	⁵⁴⁴
Canakinumab	RCT, patients with very high cardiovascular risk and hsCRP >2 mg/L, canakinumab 50, 100,	Inflammation	↓ risk of MACE	²¹⁸

	150 mg s.c. every 3 months, 3.7 yrs			
Future perspectives for geroprotective and gerotherapeutic interventions.				
Caloric restriction mimetics				
Spermidine supplementation	Spermidine supplementation to drinking water starting from the age of 4 months (life-long) or 18 months (late-in-life) of C57BL/6J female mice	Autophagy, Mitochondrial dysfunction, Epigenetics, Senescence, Inflammation	10-15% extension in median lifespan, depending on age at when treatment is initiated ↑ endothelial function, vascular stiffness	¹⁹⁵
NAD+ Precursors (nicotinamide, nicotinamide riboside, and nicotinamide mononucleotide)	Supplementation to 4 months to 28 months old C57BL/6J mice, Dahl salt-sensitive rats, ZSF1 obese rats, or mouse model of dilated cardiomyopathy	Mitochondrial dysfunction, Epigenetics, DNA stability, inflammation, senescence	0-5% extension in median lifespan depending on age when treatment is initiated ↓ cardiac hypertrophy and diastolic dysfunction, protecting from HFpEF and dilated cardiomyopathy ↓ vascular remodelling and endothelial dysfunction	^{545,211,546,547,548}
Genetic therapies				
systemic VEGFA overexpression in the circulation (1.5-2 fold)	VEGF transgenic mice	Mitochondrial dysfunction, Senescence, Inflammation	up to 48% in male mice and 39% in female mice extended lifespan ↑ perfusion and oxygenation of tissues ↓ loss of capillaries	⁴⁴⁹
ACBP neutralization	i.p. injection to 8 week old C57BL/6J female mice	Autophagy, Inflammation, Senescence	Reduced chemotherapy-induced cardiac senescence and dysfunction	⁵⁴⁹
Mitochondria				
Mitochondria-targeted antioxidants	i.p. injection to 24-month-old C57BL/6 mice	Mitochondrial dysfunction, Inflammation	↑ neurovascular coupling, cerebral microvascular and cognitive functions	³⁰⁷
mTOR inhibitor				
Rapamycin	Microencapsulated rapamycin incorporated in the chow diet starting at 270 days or 600 days in mice	Nutrient-sensing, autophagy, inflammation	14% for females and 9% for males extended lifespan	⁵⁵⁰
Parabiosis				
heterochronic parabiosis	Anastomosis surgery up to 3 months in 3- to 22-month old mice	Epigenetics, Proteostasis, Senescence, Inflammation	6-week extension in median lifespan of old mice after detachment Global multi-omic improvement ↑ endothelial function and different tissue functions	^{461,551,500}
Senolytic				
Dasatinib and quercetin	bi-weekly administration starting at 24-27 months of age	Senescence	36% higher lifespan and 64.9% lower mortality hazard	⁵⁵²

			↑ maximal walking speed, hanging endurance, grip strength, treadmill endurance, and daily activity	
Small molecules compounds				
Taurine supplementation	Daily oral administration to 14 months old female mice for 10 to 12 months.	All hallmarks	12% in female and 10% in male mice lifespan extension ↑ energy expenditure, bone mass, and muscle functions ↓ depression, anxiety-like behaviours, and reduced insulin resistance.	⁵⁵³
anti-IL-11	75-week-old mice for 25 weeks	Inflammation	25% in female and 22.5% in male mice median lifespan extension ↑ metabolism and muscle functions	⁵⁵⁴

Abbreviations:
 6MWT: 6-minute walk test; FMD: flow-mediated dilation; HFpEF: heart failure with preserved ejection fraction; hsCRP: high-sensitivity C-reactive protein; i.p.: intraperitoneal; MACE: major adverse cardiovascular event; MI: myocardial infarction; PCI: percutaneous coronary intervention; PWV: pulse wave velocity; RCT: randomized controlled trial; STEMI: ST-elevation myocardial infarction.

Supplementary Table 5. Clinical applicability of biological clocks and surrogate ageing markers							
Biological clocks	Approach/ Strategy	Predictive value for lifespan	Evidence level I, II, III	Predictive value for health span	Evidence level I, II, III	Therapeutic response	Evidence level I, II, III
Deep learning-based haematological ageing clock* ^{,555}	Several deep learning-based predictors of biological age trained upon population-specific 20 blood biochemistry biomarkers and haematological cell count datasets	Associated with all-cause mortality	I	N/A		N/A	
DNAmAge ⁵⁵⁶	At the molecular level, DNAmAge is a proximal readout of a collection of innate ageing processes that conspire with other, independent root causes of ageing to the detriment of tissue function. DNAmAge is defined as estimated ("predicted") age.	Associated with increased risk for all-cause mortality	I	Associated with risk for incident CV disease, stroke, different types of cancer, Parkinson's disease and dementia	II	Diet and lifestyle treatment leads to a decrease in DNAmAge. Vitamin D may slow down epigenetic ageing.	II
GlycanAge± ^{556,557}	Biological age test which determines biological age by measuring chronic inflammation through blood test. A panel of molecular measures based on glycans attached to immunoglobulin G antibodies associated with chronological age.	N/A		Associated with multiple diseases, among others CV disease and diabetes.	I	N/A	
PhenoAge and GrimAge ⁵⁵⁶	Both are epigenetic clocks that measure changes in DNA methylation levels at specific CpG sites that are highly correlated with calendar age.	Associated with all-cause mortality	I	Associated with risk of cancer, Alzheimer's disease, CHD. Predictive ability for age-at-menopause. Associated with multiple age-related clinical phenotypes (walking speed, frailty, and cognitive functions).	I	Significant reduction of PhenoAge by CR for 2 years. Reduction of GrimAge by plant-food-rich diet and exercise. Treatment with human umbilical cord plasma reduced GrimAge.	II
DunedinPoAm and DunedinPACE ⁵⁵⁶	Rate measure based on comparison of longitudinal change over time in 18 biomarkers of organ-system integrity among individuals who are all at the same chronological age.	Associated with all-cause mortality	I	Associated with incidence of multiple chronic diseases, including dementia and disability.	I	Significant reduction of DunedinPACE by CR for 2 years	II

Multiomic biological age estimation based on KDM⁵⁵⁶	Derived from modelling associations of biomarkers with chronological age in a reference sample and then applying parameters derived from these models in the target dataset to compute participants' biological age values.	N/A		May be a sign of healthy ageing.	I	N/A	
Ageing.AI, Deep Transcriptomic and Proteomic Clocks⁵⁵⁶	AI-based blood clocks, based on haematological parameters, transcriptomic and proteomic data.	Associated with all-cause mortality.	I	N/A		N/A	
Proteomic clocks^{558,559}	Proteomic clocks include protein-based biomarkers, they consist of an intermediate phenotype that is most proximal to age-related diseases, and thus may provide more accurate information on ageing and age-related pathologies.	May predict CV death	I	Screening of prostate cancer, prediction of CV events in CCS patients. Associated with body and liver fat, body mass, alcohol consumption, physical activity, conversion from pre-diabetes to DM	I	N/A	
Metabolomic clock⁵⁵⁸	Metabolomic clocks measure the structural and functional building blocks of an organism (through measurement of all metabolites and low-molecular-weight molecules in biological specimens) as a powerful link between genotype and phenotype in aging and age-related diseases.	Associated with all-cause, CV, cancer- and infection-related mortality	I	Associated with the risk of CV disease and functionality in individuals with advanced age.	I	N/A	
Inflammatory ageing clock⁵⁶⁰	A metric for age-related chronic inflammation based on blood immune biomarkers to aid in prediction of important ageing phenotypes and provide insights into the mechanisms leading to vascular ageing.	Associated with exceptional longevity in centenarians	II	Associated with multimorbidit y, immunosenescence, frailty and CV ageing.	II	N/A	
Immune aging (IMM-AGE) score⁵⁶¹	A high-dimensional trajectory of immune ageing that describes a person's immune status better than chronological age,	Better performance in predicting mortality in older	II	N/A		N/A	

		adults than the epigenetic clock.					
Surrogate ageing markers							
CMR radiomics ^{562,563}	CMR radiomics may be used for detailed cardiovascular phenotyping, by providing multiple quantifiers of ventricular shape and myocardial texture.	N/A		Associated with cardiac ageing	I	N/A	
Echocardiographic heart ageing patterns ⁵⁶⁴	A standard Doppler echocardiogram able to detect distinct heart ageing patterns (associations of age with LV mass, geometry, diastolic function, LA volume, and aortic root size) which reflect different biological susceptibilities to age-dependent diseases and provide a new tool for personalising timeliness and intensity of prevention.	Associated with all-cause and CV mortality	I	Predictive value for biological age, CV and non-CV events	I	N/A	
ECG-based heart age ⁵⁶⁵	ECG can reflect physiological status of the heart better than chronological age through specific ECG patterns.	Associated with all-cause and CV mortality	I	Associated with HF, stroke, CAD, AMI, AF and DM	I	N/A	
Retinal age gap ^{566,567}	The retinal age gap is the difference between the biological age of the retina -assessed by deep learning- and a person's chronological age. A positive value indicates an 'older' appearing retina.	Associated with all-cause mortality and mortality attributable to non-CV and non-cancer disease.	I	Associated with risk of stroke. Predictive value for biological age	I	N/A	
CXR-age ⁵⁶⁸	Estimation of a patient's age from CXR based on radiological findings.	Associated with all-cause and CV mortality	I	N/A		N/A	
Visible age-related signs** ⁵⁶⁹	Assumes that presence of visible age-related signs is a marker of the actual biological age of an individual.	Associated with all-cause mortality in older people	I	Increased risk of IHD and AMI. Correlated with innate immunity at	I	N/A	

				transcriptome level			
PWV	Marker of arterial stiffness, the velocity at which the pressure waves, generated by the systolic contraction of the heart, propagates along the arterial tree, usually between the common carotid and the common femoral artery.	Increased PWV values associated with all-cause and CV mortality ^{57 0-574}	I	Associated with diabetes, hypertension, stroke, CAD, hypertrophic cardiomyopathy, myocardial fibrosis, early mild diastolic HF, sleep-disordered breathing in post-stroke patients. ⁵⁷⁴⁻⁵⁸² Determinant of myocardial ischemic threshold ⁵⁸³		Only 2 studies ^{584,585} have showed in an indirect way that an improvement in outcomes is mediated through improvement in PWV. Large geroscience studies should confirm lifespan or healthspan extension clinical benefit induced by treatments leading to arterial stiffness regression.	III
Carotid IMT⁵⁸⁶⁻⁵⁸⁹	A quick, safe and bedside test that measures the thickness of the carotid artery wall at paired segments.	Associated with CV mortality	I	Associated with traditional CV factors	II	The extent of intervention effects on carotid IMT progression predicted the degree of CV risk reduction.	I
Carotid plaque parameters ***.^{590,591}	Carotid plaque parameters include plaque presence, number, thickness, area and volume.	Associated with all-cause and CV mortality	I	Associated with traditional CV factors	I	N/A	
CAC score⁵⁹²⁻⁵⁹⁴	CAC score measures the amount of calcified plaque in coronary arteries by computed tomography scan.	Associated with all-cause, CV, cancer and CHD mortality	I	Associated with traditional CV factors. CAC progression associated with risk of AMI. May predict incident cancer, CKD, COPD, and dementia.	I	N/A	

Sirtuins ⁵⁵⁸	Sirtuin proteins comprise a group of nutrient- and stress-responsive factors that regulate diverse cellular processes to promote health span.	Associated with lifespan	I	Associated with multiple mechanisms of CV disease	II	CR was associated with an increased expression of SIRT1 and SIRT1 was suggested as the main mediator of prolonged lifespan after CR.	
Hs-cTnT ^{595,596}	Newer type of biomarker which helps to diagnose heart injury and acute coronary syndrome earlier.	Associated with all-cause and CV mortality	I	Increased in older people. Associated with structural heart disease (left ventricular hypertrophy), CHD and HF.	I	N/A	
hs-CRP ^{218,597,598}	A marker of inflammation with established relationship with atherothrombotic disease.	Associated with all-cause and CV mortality	I	Increased in older populations. Improves CV disease prediction.	I	Achievement of on-treatment CRP <2 mg/L leads to significant reduction of both CV and all-cause mortality.	I
Serum BNP ⁵⁹⁹⁻⁶⁰¹	Method for detection of heart failure by measuring the amount of BNP or its prohormone NT-proBNP in the bloodstream.	Associated with all-cause mortality	I	Increased in aged individuals, age-related impairment of left atrial strain positively correlates with higher BNP levels. Marker of increased risk of HF in older adults. Inversely associated with walking speed, chair rise speed, balance time, and grip strength.	II	N/A	
IL-6 ^{601,602}	Important in ageing and age-related disease and has been called the “gerontologist’s	Independent predictor of all-	II	Increased in older populations.	II		

	cytokine". IL-6 plays a key role in the acute phase response, in metabolic control and in the pathogenesis of many chronic diseases.	cause mortality, CV and non-CV mortality		Inversely associated with walking speed, chair rise speed, balance time, and grip strength. Associated with iron deficiency, reduced LVEF and AF in HF patients.			
Urinary BNP ⁵⁵⁸	Method for detection of heart failure by measuring the amount of BNP in the urine.	Associated with all-cause mortality	II	May diagnose HF	II	N/A	
Urinary FPA ⁶⁰³	Urinary FPA is probably a valuable marker of low-grade activation of coagulation, particularly in chronic conditions.	Associated with CV mortality	II	Independent predictor of CV events in patients presenting with chest pain (AMI and angina pectoris)	II		

I - Evidence from large representative population samples. II - Evidence from small, well designed but not necessarily representative samples. III - Evidence from non-representative surveys, case reports.

* Visible age-related signs included male pattern baldness, earlobe crease, and xanthelasmata-alone or in combination.

** In this deep learning-based haematological ageing clock 20 blood biochemistry markers were included.

¶ Epigenetic clocks, based on a set of DNA methylation measures associated with chronological age.

¤ Epigenetic clocks, based on a set of DNA methylation measures associated with "clinical phenotypic age measures" (a panel of age associated molecular and physiological biomarkers, measured in blood).

¶ KDM applied to over 900 principal component transformed biomarkers (metabolites, proteins, genomics, and clinical measures). Abbreviations: DNA, deoxyribonucleic acid; CV, cardiovascular; CpG, 5'-C—phosphate—G—3'; CHD, coronary heart disease; CR, calorie restriction; KDM, Klemara and Doubal method; AI, artificial intelligence; CCS, chronic coronary syndrome; DM, diabetes mellitus; CMR, cardiovascular magnetic resonance; LV, left ventricular; LA, left atrium; ECG, electrocardiogram; HF, heart failure; CAD, coronary artery disease; AMI, acute myocardial infarction; AF, atrial fibrillation; CXR, chest X-ray; IHD, ischemic heart disease; PWV, pulse wave velocity; IMT, intima media thickness; CAC, coronary artery calcium; CKD, chronic kidney disease; COPD, chronic obstructive pulmonary disease; SIRT1, sirtuin 1; hs-cTnT, high-sensitive cardiac troponin T; hs-CRP, high-sensitivity C-reactive protein; BNP, B-type natriuretic peptide; NT-proBNP, N-terminal pro B-type natriuretic peptide; IL-6 interleukin-6; LVEF, left ventricular ejection fraction; FPA, fibrinopeptide.

References

- 1 Bloom, S. I. *et al.* Aging results in DNA damage and telomere dysfunction that is greater in endothelial versus vascular smooth muscle cells and is exacerbated in atheroprone regions. *Geroscience* **44**, 2741-2755, doi:10.1007/s11357-022-00681-6 (2022).
- 2 Durik, M. *et al.* Nucleotide excision DNA repair is associated with age-related vascular dysfunction. *Circulation* **126**, 468-478, doi:10.1161/CIRCULATIONAHA.112.104380 (2012).
- 3 Pieren, D. K. J. *et al.* Compromised DNA Repair Promotes the Accumulation of Regulatory T Cells With an Aging-Related Phenotype and Responsiveness. *Front Aging* **2**, doi:10.3389/fragi.2021.667193 (2021).
- 4 Ding, Q., Shao, C., Rose, P. & Zhu, Y. Z. Epigenetics and Vascular Senescence-Potential New Therapeutic Targets? *Front Pharmacol* **11**, 535395, doi:10.3389/fphar.2020.535395 (2020).
- 5 Lan, Y. *et al.* Long noncoding RNA MEG3 prevents vascular endothelial cell senescence by impairing miR-128-dependent Girdin downregulation. *Am J Physiol Cell Physiol* **316**, C830-C843, doi:10.1152/ajpcell.00262.2018 (2019).
- 6 Menghini, R. *et al.* MicroRNA 217 modulates endothelial cell senescence via silent information regulator 1. *Circulation* **120**, 1524-1532, doi:10.1161/CIRCULATIONAHA.109.864629 (2009).
- 7 Hofmann, P. *et al.* Long non-coding RNA H19 regulates endothelial cell aging via inhibition of STAT3 signalling. *Cardiovasc Res* **115**, 230-242, doi:10.1093/cvr/cvy206 (2019).
- 8 Hu, J. *et al.* Disrupted Binding of Cystathionine gamma-Lyase to p53 Promotes Endothelial Senescence. *Circ Res* **133**, 842-857, doi:10.1161/CIRCRESAHA.123.323084 (2023).
- 9 Kopacz, A. *et al.* Keap1 governs ageing-induced protein aggregation in endothelial cells. *Redox Biol* **34**, 101572, doi:10.1016/j.redox.2020.101572 (2020).
- 10 Zhang, L. *et al.* CD44 connects autophagy decline and ageing in the vascular endothelium. *Nat Commun* **14**, 5524, doi:10.1038/s41467-023-41346-y (2023).
- 11 LaRocca, T. J., Henson, G. D., Thorburn, A., Sindler, A. L., Pierce, G. L. & Seals, D. R. Translational evidence that impaired autophagy contributes to arterial ageing. *J Physiol* **590**, 3305-3316, doi:10.1113/jphysiol.2012.229690 (2012).
- 12 Wagner, J. U. G. *et al.* Aging impairs the neurovascular interface in the heart. *Science* **381**, 897-906, doi:10.1126/science.adc4961 (2023).
- 13 Donato, A. J. *et al.* Direct evidence of endothelial oxidative stress with aging in humans: relation to impaired endothelium-dependent dilation and upregulation of nuclear factor-kappaB. *Circ Res* **100**, 1659-1666, doi:10.1161/01.RES.0000269183.13937.e8 (2007).
- 14 Donato, A. J., Black, A. D., Jablonski, K. L., Gano, L. B. & Seals, D. R. Aging is associated with greater nuclear NF kappa B, reduced I kappa B alpha, and increased expression of proinflammatory cytokines in vascular endothelial cells of healthy humans. *Aging Cell* **7**, 805-812, doi:10.1111/j.1474-9726.2008.00438.x (2008).
- 15 Shimizu, S. *et al.* Aging impairs the ability of vascular endothelial stem cells to generate endothelial cells in mice. *Angiogenesis* **26**, 567-580, doi:10.1007/s10456-023-09891-8 (2023).
- 16 Dobner, S., Toth, F. & de Rooij, L. A high-resolution view of the heterogeneous aging endothelium. *Angiogenesis*, doi:10.1007/s10456-023-09904-6 (2024).
- 17 Liu, L. *et al.* Exercise reprograms the inflammatory landscape of multiple stem cell compartments during mammalian aging. *Cell Stem Cell* **30**, 689-705 e684, doi:10.1016/j.stem.2023.03.016 (2023).
- 18 Ma, S. *et al.* Single-cell transcriptomic atlas of primate cardiopulmonary aging. *Cell Res* **31**, 415-432, doi:10.1038/s41422-020-00412-6 (2021).
- 19 Catry, E. *et al.* Targeting the gut microbiota with inulin-type fructans: preclinical demonstration of a novel approach in the management of endothelial dysfunction. *Gut* **67**, 271-283, doi:10.1136/gutjnl-2016-313316 (2018).
- 20 Donato, A. J., Magerko, K. A., Lawson, B. R., Durrant, J. R., Lesniewski, L. A. & Seals, D. R. SIRT-1 and vascular endothelial dysfunction with ageing in mice and humans. *J Physiol* **589**, 4545-4554, doi:10.1113/jphysiol.2011.211219 (2011).

- 21 Ungvari, Z., Labinskyy, N., Gupte, S., Chander, P. N., Edwards, J. G. & Csiszar, A. Dysregulation of mitochondrial biogenesis in vascular endothelial and smooth muscle cells of aged rats. *Am J Physiol Heart Circ Physiol* **294**, H2121-2128, doi:10.1152/ajpheart.00012.2008 (2008).
- 22 Rossman, M. J. et al. Chronic Supplementation With a Mitochondrial Antioxidant (MitoQ) Improves Vascular Function in Healthy Older Adults. *Hypertension* **71**, 1056-1063, doi:10.1161/HYPERTENSIONAHA.117.10787 (2018).
- 23 Rossman, M. J. et al. Endothelial cell senescence with aging in healthy humans: prevention by habitual exercise and relation to vascular endothelial function. *Am J Physiol Heart Circ Physiol* **313**, H890-H895, doi:10.1152/ajpheart.00416.2017 (2017).
- 24 Kiss, T. et al. Single-cell RNA sequencing identifies senescent cerebromicrovascular endothelial cells in the aged mouse brain. *Geroscience* **42**, 429-444, doi:10.1007/s11357-020-00177-1 (2020).
- 25 Bloom, S. I., Islam, M. T., Lesniewski, L. A. & Donato, A. J. Mechanisms and consequences of endothelial cell senescence. *Nat Rev Cardiol* **20**, 38-51, doi:10.1038/s41569-022-00739-0 (2023).
- 26 From the American Association of Neurological Surgeons, A. S. o. N. C. et al. Multisociety Consensus Quality Improvement Revised Consensus Statement for Endovascular Therapy of Acute Ischemic Stroke. *Int J Stroke* **13**, 612-632, doi:10.1177/1747493018778713 (2018).
- 27 Demanelis, K. et al. Determinants of telomere length across human tissues. *Science* **369**, doi:10.1126/science.aaz6876 (2020).
- 28 Badi, I. et al. miR-34a Promotes Vascular Smooth Muscle Cell Calcification by Downregulating SIRT1 (Sirtuin 1) and Axl (AXL Receptor Tyrosine Kinase). *Arterioscler Thromb Vasc Biol* **38**, 2079-2090, doi:10.1161/ATVBAHA.118.311298 (2018).
- 29 Wang, L. et al. Cholesterol-induced HRD1 reduction accelerates vascular smooth muscle cell senescence via stimulation of endoplasmic reticulum stress-induced reactive oxygen species. *J Mol Cell Cardiol* **187**, 51-64, doi:10.1016/j.yjmcc.2023.12.007 (2024).
- 30 Grootaert, M. O. et al. Defective autophagy in vascular smooth muscle cells accelerates senescence and promotes neointima formation and atherogenesis. *Autophagy* **11**, 2014-2032, doi:10.1080/15548627.2015.1096485 (2015).
- 31 Lacolley, P., Regnault, V. & Avolio, A. P. Smooth muscle cell and arterial aging: basic and clinical aspects. *Cardiovasc Res* **114**, 513-528, doi:10.1093/cvr/cv009 (2018).
- 32 Spinetti, G., Wang, M., Monticone, R., Zhang, J., Zhao, D. & Lakatta, E. G. Rat aortic MCP-1 and its receptor CCR2 increase with age and alter vascular smooth muscle cell function. *Arterioscler Thromb Vasc Biol* **24**, 1397-1402, doi:10.1161/01.ATV.0000134529.65173.08 (2004).
- 33 Benson, T. W. et al. Gut Microbiota-Derived Trimethylamine N-Oxide Contributes to Abdominal Aortic Aneurysm Through Inflammatory and Apoptotic Mechanisms. *Circulation* **147**, 1079-1096, doi:10.1161/CIRCULATIONAHA.122.060573 (2023).
- 34 Chen, H. Z. et al. Age-Associated Sirtuin 1 Reduction in Vascular Smooth Muscle Links Vascular Senescence and Inflammation to Abdominal Aortic Aneurysm. *Circ Res* **119**, 1076-1088, doi:10.1161/CIRCRESAHA.116.308895 (2016).
- 35 Gorenne, I. et al. Vascular smooth muscle cell sirtuin 1 protects against DNA damage and inhibits atherosclerosis. *Circulation* **127**, 386-396, doi:10.1161/CIRCULATIONAHA.112.124404 (2013).
- 36 Thompson, A. M., Wagner, R. & Rzucidlo, E. M. Age-related loss of SirT1 expression results in dysregulated human vascular smooth muscle cell function. *Am J Physiol Heart Circ Physiol* **307**, H533-541, doi:10.1152/ajpheart.00871.2013 (2014).
- 37 Ragnauth, C. D. et al. Prelamin A acts to accelerate smooth muscle cell senescence and is a novel biomarker of human vascular aging. *Circulation* **121**, 2200-2210, doi:10.1161/CIRCULATIONAHA.109.902056 (2010).

- 38 Rodriguez-Menocal, L. et al. Aging increases p16 INK4a expression in vascular smooth-muscle cells. *Biosci Rep* **30**, 11-18, doi:10.1042/BSR20080128 (2009).
- 39 Han, J., Liu, J. Y., Swartz, D. D. & Andreadis, S. T. Molecular and functional effects of organismal ageing on smooth muscle cells derived from bone marrow mesenchymal stem cells. *Cardiovasc Res* **87**, 147-155, doi:10.1093/cvr/cvq024 (2010).
- 40 Iwao, T. et al. Senescence in brain pericytes attenuates blood-brain barrier function in vitro: A comparison of serially passaged and isolated pericytes from aged rat brains. *Biochem Biophys Res Commun* **645**, 154-163, doi:10.1016/j.bbrc.2023.01.037 (2023).
- 41 Anderson, R. et al. Length-independent telomere damage drives post-mitotic cardiomyocyte senescence. *EMBO J* **38**, doi:10.15252/embj.2018100492 (2019).
- 42 Ye, S., Zhou, X., Chen, P. & Lin, J. F. Folic acid attenuates remodeling and dysfunction in the aging heart through the ER stress pathway. *Life Sci* **264**, 118718, doi:10.1016/j.lfs.2020.118718 (2021).
- 43 Miyamoto, S. Autophagy and cardiac aging. *Cell Death Differ* **26**, 653-664, doi:10.1038/s41418-019-0286-9 (2019).
- 44 Chang, K. et al. TGFB-INHB/activin signaling regulates age-dependent autophagy and cardiac health through inhibition of MTORC2. *Autophagy* **16**, 1807-1822, doi:10.1080/15548627.2019.1704117 (2020).
- 45 Tang, X., Li, P. H. & Chen, H. Z. Cardiomyocyte Senescence and Cellular Communications Within Myocardial Microenvironments. *Front Endocrinol (Lausanne)* **11**, 280, doi:10.3389/fendo.2020.00280 (2020).
- 46 Li, T. et al. Pathological implication of CaMKII in NF-kappaB pathway and SASP during cardiomyocytes senescence. *Mech Ageing Dev* **209**, 111758, doi:10.1016/j.mad.2022.111758 (2023).
- 47 Kumar, V. et al. RelA-mediated signaling connects adaptation to chronic cardiomyocyte stress with myocardial and systemic inflammation in the ADCY8 model of accelerated aging. *Geroscience*, doi:10.1007/s11357-024-01121-3 (2024).
- 48 Gao, H. et al. Gut lumen-leaked microbial DNA causes myocardial inflammation and impairs cardiac contractility in ageing mouse heart. *Front Immunol* **14**, 1216344, doi:10.3389/fimmu.2023.1216344 (2023).
- 49 Abdellatif, M. et al. Fine-Tuning Cardiac Insulin-Like Growth Factor 1 Receptor Signaling to Promote Health and Longevity. *Circulation* **145**, 1853-1866, doi:10.1161/CIRCULATIONAHA.122.059863 (2022).
- 50 Ruiz-Meana, M. et al. Ryanodine Receptor Glycation Favors Mitochondrial Damage in the Senescent Heart. *Circulation* **139**, 949-964, doi:10.1161/CIRCULATIONAHA.118.035869 (2019).
- 51 Muller-Hocker, J. Cytochrome-c-oxidase deficient cardiomyocytes in the human heart--an age-related phenomenon. A histochemical ultracytochemical study. *Am J Pathol* **134**, 1167-1173 (1989).
- 52 Rouhi, L. et al. Deletion of the Lmna gene in fibroblasts causes senescence-associated dilated cardiomyopathy by activating the double-stranded DNA damage response and induction of senescence-associated secretory phenotype. *J Cardiovasc Aging* **2**, doi:10.20517/jca.2022.14 (2022).
- 53 Vidal, R. et al. Transcriptional heterogeneity of fibroblasts is a hallmark of the aging heart. *JCI Insight* **4**, doi:10.1172/jci.insight.131092 (2019).
- 54 Ock, S., Ham, W., Kang, C. W., Kang, H., Lee, W. S. & Kim, J. IGF-1 protects against angiotensin II-induced cardiac fibrosis by targeting alphaSMA. *Cell Death Dis* **12**, 688, doi:10.1038/s41419-021-03965-5 (2021).
- 55 Azar, A., Lawrence, I., Jofre, S., Mell, J. & Sell, C. Distinct patterns of gene expression in human cardiac fibroblasts exposed to rapamycin treatment or methionine restriction. *Ann N Y Acad Sci* **1418**, 95-105, doi:10.1111/nyas.13566 (2018).

- 56 Vue, Z. *et al.* Three-dimensional mitochondria reconstructions of murine cardiac muscle changes in size across aging. *Am J Physiol Heart Circ Physiol* **325**, H965-H982, doi:10.1152/ajpheart.00202.2023 (2023).
- 57 Sawaki, D. *et al.* Visceral Adipose Tissue Drives Cardiac Aging Through Modulation of Fibroblast Senescence by Osteopontin Production. *Circulation* **138**, 809-822, doi:10.1161/CIRCULATIONAHA.117.031358 (2018).
- 58 Hu, W. S., Chen, J. Y., Liao, W. Y., Chang, C. H. & Chen, T. S. Regulation of ROS/inflammasome Axis is Essential for Cardiac Regeneration in Aging Rats Receiving Transplantation of Mesenchymal Stem Cells. *Curr Stem Cell Res Ther*, doi:10.2174/011574888X276612231121065203 (2023).
- 59 Vas, V., Senger, K., Dorr, K., Niebel, A. & Geiger, H. Aging of the microenvironment influences clonality in hematopoiesis. *PLoS One* **7**, e42080, doi:10.1371/journal.pone.0042080 (2012).
- 60 Wang, J., Lu, X., Sakk, V., Klein, C. A. & Rudolph, K. L. Senescence and apoptosis block hematopoietic activation of quiescent hematopoietic stem cells with short telomeres. *Blood* **124**, 3237-3240, doi:10.1182/blood-2014-04-568055 (2014).
- 61 Vaziri, H., Dragowska, W., Allsopp, R. C., Thomas, T. E., Harley, C. B. & Lansdorp, P. M. Evidence for a mitotic clock in human hematopoietic stem cells: loss of telomeric DNA with age. *Proc Natl Acad Sci U S A* **91**, 9857-9860, doi:10.1073/pnas.91.21.9857 (1994).
- 62 Mejia-Ramirez, E. & Florian, M. C. Understanding intrinsic hematopoietic stem cell aging. *Haematologica* **105**, 22-37, doi:10.3324/haematol.2018.211342 (2020).
- 63 Beerman, I. *et al.* Proliferation-dependent alterations of the DNA methylation landscape underlie hematopoietic stem cell aging. *Cell Stem Cell* **12**, 413-425, doi:10.1016/j.stem.2013.01.017 (2013).
- 64 Chapple, R. H. *et al.* ERalpha promotes murine hematopoietic regeneration through the Ire1alpha-mediated unfolded protein response. *eLife* **7**, doi:10.7554/eLife.31159 (2018).
- 65 Warr, M. R. *et al.* FOXO3A directs a protective autophagy program in haematopoietic stem cells. *Nature* **494**, 323-327, doi:10.1038/nature11895 (2013).
- 66 Yeganeh, A. *et al.* Age-related defects in autophagy alter the secretion of paracrine factors from bone marrow mononuclear cells. *Aging (Albany NY)* **13**, 14687-14708, doi:10.18632/aging.203127 (2021).
- 67 Conboy, I. M., Conboy, M. J., Wagers, A. J., Girma, E. R., Weissman, I. L. & Rando, T. A. Rejuvenation of aged progenitor cells by exposure to a young systemic environment. *Nature* **433**, 760-764, doi:10.1038/nature03260 (2005).
- 68 Sun, D. *et al.* Epigenomic profiling of young and aged HSCs reveals concerted changes during aging that reinforce self-renewal. *Cell Stem Cell* **14**, 673-688, doi:10.1016/j.stem.2014.03.002 (2014).
- 69 Kovtonyuk, L. V. *et al.* IL-1 mediates microbiome-induced inflammaging of hematopoietic stem cells in mice. *Blood* **139**, 44-58, doi:10.1182/blood.2021011570 (2022).
- 70 Tang, D. *et al.* Dietary restriction improves repopulation but impairs lymphoid differentiation capacity of hematopoietic stem cells in early aging. *J Exp Med* **213**, 535-553, doi:10.1084/jem.20151100 (2016).
- 71 Ito, K. *et al.* Reactive oxygen species act through p38 MAPK to limit the lifespan of hematopoietic stem cells. *Nat Med* **12**, 446-451, doi:10.1038/nm1388 (2006).
- 72 Mohrin, M. *et al.* Stem cell aging. A mitochondrial UPR-mediated metabolic checkpoint regulates hematopoietic stem cell aging. *Science* **347**, 1374-1377, doi:10.1126/science.aaa2361 (2015).
- 73 Chang, J. *et al.* Clearance of senescent cells by ABT263 rejuvenates aged hematopoietic stem cells in mice. *Nat Med* **22**, 78-83, doi:10.1038/nm.4010 (2016).
- 74 Rossi, D. J. *et al.* Cell intrinsic alterations underlie hematopoietic stem cell aging. *Proc Natl Acad Sci U S A* **102**, 9194-9199, doi:10.1073/pnas.0503280102 (2005).

- 75 Leins, H. *et al.* Aged murine hematopoietic stem cells drive aging-associated immune remodeling. *Blood* **132**, 565-576, doi:10.1182/blood-2018-02-831065 (2018).
- 76 Najarro, K. *et al.* Telomere Length as an Indicator of the Robustness of B- and T-Cell Response to Influenza in Older Adults. *J Infect Dis* **212**, 1261-1269, doi:10.1093/infdis/jiv202 (2015).
- 77 Dalzini, A., Petrara, M. R., Ballin, G., Zanchetta, M., Giaquinto, C. & De Rossi, A. Biological Aging and Immune Senescence in Children with Perinatally Acquired HIV. *J Immunol Res* **2020**, 8041616, doi:10.1155/2020/8041616 (2020).
- 78 Ucar, D. *et al.* The chromatin accessibility signature of human immune aging stems from CD8(+) T cells. *J Exp Med* **214**, 3123-3144, doi:10.1084/jem.20170416 (2017).
- 79 Carre, C. *et al.* Endoplasmic reticulum stress response and bile acid signatures associate with multi-strain seroresponsiveness during elderly influenza vaccination. *iScience* **24**, 102970, doi:10.1016/j.isci.2021.102970 (2021).
- 80 Zhang, H. *et al.* Polyamines Control eIF5A Hypusination, TFEB Translation, and Autophagy to Reverse B Cell Senescence. *Mol Cell* **76**, 110-125 e119, doi:10.1016/j.molcel.2019.08.005 (2019).
- 81 Li, K. *et al.* B cells from old mice induce the generation of inflammatory T cells through metabolic pathways. *Mech Ageing Dev* **209**, 111742, doi:10.1016/j.mad.2022.111742 (2023).
- 82 Kawamoto, S. *et al.* Bacterial induction of B cell senescence promotes age-related changes in the gut microbiota. *Nat Cell Biol* **25**, 865-876, doi:10.1038/s41556-023-01145-5 (2023).
- 83 Ebersole, J. L. *et al.* Transcriptome Analysis of B Cell Immune Functions in Periodontitis: Mucosal Tissue Responses to the Oral Microbiome in Aging. *Front Immunol* **7**, 272, doi:10.3389/fimmu.2016.00272 (2016).
- 84 Krambs, J. R., Monligh, D. A., Gao, F., Schuettpelz, L. G. & Link, D. C. Microbiota Signals Suppress B Lymphopoiesis With Aging in Mice. *Front Immunol* **12**, 767267, doi:10.3389/fimmu.2021.767267 (2021).
- 85 Dowery, R. *et al.* Peripheral B cells repress B-cell regeneration in aging through a TNF-alpha/IGFBP-1/IGF-1 immune-endocrine axis. *Blood* **138**, 1817-1829, doi:10.1182/blood.2021012428 (2021).
- 86 El-Naseery, N. I., Mousa, H. S. E., Noreldin, A. E., El-Far, A. H. & Elewa, Y. H. A. Aging-associated immunosenescence via alterations in splenic immune cell populations in rat. *Life Sci* **241**, 117168, doi:10.1016/j.lfs.2019.117168 (2020).
- 87 Frasca, D. Senescent B cells in aging and age-related diseases: Their role in the regulation of antibody responses. *Exp Gerontol* **107**, 55-58, doi:10.1016/j.exger.2017.07.002 (2018).
- 88 Ross, J. B. *et al.* Depleting myeloid-biased haematopoietic stem cells rejuvenates aged immunity. *Nature* **628**, 162-170, doi:10.1038/s41586-024-07238-x (2024).
- 89 Chebly, A., Khalil, C., Kuzyk, A., Beylot-Barry, M. & Chevret, E. T-cell lymphocytes' aging clock: telomeres, telomerase and aging. *Biogerontology* **25**, 279-288, doi:10.1007/s10522-023-10075-6 (2024).
- 90 Sanderson, S. L. & Simon, A. K. In aged primary T cells, mitochondrial stress contributes to telomere attrition measured by a novel imaging flow cytometry assay. *Aging Cell* **16**, 1234-1243, doi:10.1111/acel.12640 (2017).
- 91 Ponnappan, U., Zhong, M. & Trebilcock, G. U. Decreased proteasome-mediated degradation in T cells from the elderly: A role in immune senescence. *Cell Immunol* **192**, 167-174, doi:10.1006/cimm.1998.1418 (1999).
- 92 Ponnappan, U. Ubiquitin-proteasome pathway is compromised in CD45RO+ and CD45RA+ T lymphocyte subsets during aging. *Exp Gerontol* **37**, 359-367, doi:10.1016/s0531-5565(01)00203-0 (2002).
- 93 Das, R., Ponnappan, S. & Ponnappan, U. Redox regulation of the proteasome in T lymphocytes during aging. *Free Radic Biol Med* **42**, 541-551, doi:10.1016/j.freeradbiomed.2006.11.020 (2007).

- 94 Bharath, L. P. *et al.* Metformin Enhances Autophagy and Normalizes Mitochondrial Function to Alleviate Aging-Associated Inflammation. *Cell Metab* **32**, 44-55 e46, doi:10.1016/j.cmet.2020.04.015 (2020).
- 95 Bektas, A. *et al.* Age-associated changes in human CD4(+) T cells point to mitochondrial dysfunction consequent to impaired autophagy. *Aging (Albany NY)* **11**, 9234-9263, doi:10.18632/aging.102438 (2019).
- 96 Wang, T. W. *et al.* Blocking PD-L1-PD-1 improves senescence surveillance and ageing phenotypes. *Nature* **611**, 358-364, doi:10.1038/s41586-022-05388-4 (2022).
- 97 Henson, S. M., Macaulay, R., Riddell, N. E., Nunn, C. J. & Akbar, A. N. Blockade of PD-1 or p38 MAP kinase signaling enhances senescent human CD8(+) T-cell proliferation by distinct pathways. *Eur J Immunol* **45**, 1441-1451, doi:10.1002/eji.201445312 (2015).
- 98 Bapat, S. P. *et al.* Depletion of fat-resident Treg cells prevents age-associated insulin resistance. *Nature* **528**, 137-141, doi:10.1038/nature16151 (2015).
- 99 Zanni, F. *et al.* Marked increase with age of type 1 cytokines within memory and effector/cytotoxic CD8+ T cells in humans: a contribution to understand the relationship between inflammation and immunosenescence. *Exp Gerontol* **38**, 981-987, doi:10.1016/s0531-5565(03)00160-8 (2003).
- 100 Fagnoni, F. F. *et al.* Expansion of cytotoxic CD8+ CD28- T cells in healthy ageing people, including centenarians. *Immunology* **88**, 501-507, doi:10.1046/j.1365-2567.1996.d01-689.x (1996).
- 101 Sbierski-Kind, J. *et al.* T cell phenotypes associated with insulin resistance: results from the Berlin Aging Study II. *Immun Ageing* **17**, 40, doi:10.1186/s12979-020-00211-y (2020).
- 102 Wu, D. *et al.* T reg-specific insulin receptor deletion prevents diet-induced and age-associated metabolic syndrome. *J Exp Med* **217**, doi:10.1084/jem.20191542 (2020).
- 103 Desdin-Mico, G. *et al.* T cells with dysfunctional mitochondria induce multimorbidity and premature senescence. *Science* **368**, 1371-1376, doi:10.1126/science.aax0860 (2020).
- 104 Ron-Harel, N. *et al.* Defective respiration and one-carbon metabolism contribute to impaired naive T cell activation in aged mice. *Proc Natl Acad Sci U S A* **115**, 13347-13352, doi:10.1073/pnas.1804149115 (2018).
- 105 Yousefzadeh, M. J. *et al.* Tissue specificity of senescent cell accumulation during physiologic and accelerated aging of mice. *Aging Cell* **19**, e13094, doi:10.1111/acel.13094 (2020).
- 106 Martinez-Zamudio, R. I., Dewald, H. K., Vasilopoulos, T., Gittens-Williams, L., Fitzgerald-Bocarsly, P. & Herbig, U. Senescence-associated beta-galactosidase reveals the abundance of senescent CD8+ T cells in aging humans. *Aging Cell* **20**, e13344, doi:10.1111/acel.13344 (2021).
- 107 Denman, C. J. *et al.* Membrane-bound IL-21 promotes sustained ex vivo proliferation of human natural killer cells. *PLOS One* **7**, e30264, doi:10.1371/journal.pone.0030264 (2012).
- 108 Sagiv, A., Biran, A., Yon, M., Simon, J., Lowe, S. W. & Krizhanovsky, V. Granule exocytosis mediates immune surveillance of senescent cells. *Oncogene* **32**, 1971-1977, doi:10.1038/onc.2012.206 (2013).
- 109 Hazeldine, J., Hampson, P. & Lord, J. M. Reduced release and binding of perforin at the immunological synapse underlies the age-related decline in natural killer cell cytotoxicity. *Aging Cell* **11**, 751-759, doi:10.1111/j.1474-9726.2012.00839.x (2012).
- 110 Almeida-Oliveira, A. *et al.* Age-related changes in natural killer cell receptors from childhood through old age. *Hum Immunol* **72**, 319-329, doi:10.1016/j.humimm.2011.01.009 (2011).
- 111 Mariani, E. *et al.* Different IL-8 production by T and NK lymphocytes in elderly subjects. *Mech Ageing Dev* **122**, 1383-1395, doi:10.1016/s0047-6374(01)00270-6 (2001).
- 112 Mariani, E. *et al.* RANTES and MIP-1alpha production by T lymphocytes, monocytes and NK cells from nonagenarian subjects. *Exp Gerontol* **37**, 219-226, doi:10.1016/s0531-5565(01)00187-5 (2002).

- 113 Krishnaraj, R. & Bhooma, T. Cytokine sensitivity of human NK cells during immunosenescence.
2. IL2-induced interferon gamma secretion. *Immunol Lett* **50**, 59-63, doi:10.1016/0165-2478(96)02519-9 (1996).
- 114 Krishnaraj, R. Senescence and cytokines modulate the NK cell expression. *Mech Ageing Dev* **96**, 89-101, doi:10.1016/s0047-6374(97)00045-6 (1997).
- 115 Finamore, A. et al. Supplementation with Bifidobacterium longum Bar33 and Lactobacillus helveticus Bar13 mixture improves immunity in elderly humans (over 75 years) and aged mice. *Nutrition* **63-64**, 184-192, doi:10.1016/j.nut.2019.02.005 (2019).
- 116 Nieman, D. C. et al. Physical activity and immune function in elderly women. *Med Sci Sports Exerc* **25**, 823-831, doi:10.1249/00005768-199307000-00011 (1993).
- 117 Yan, H., Kuroiwa, A., Tanaka, H., Shindo, M., Kiyonaga, A. & Nagayama, A. Effect of moderate exercise on immune senescence in men. *Eur J Appl Physiol* **86**, 105-111, doi:10.1007/s004210100521 (2001).
- 118 Fairey, A. S., Courneya, K. S., Field, C. J., Bell, G. J., Jones, L. W. & Mackey, J. R. Randomized controlled trial of exercise and blood immune function in postmenopausal breast cancer survivors. *J Appl Physiol* (1985) **98**, 1534-1540, doi:10.1152/japplphysiol.00566.2004 (2005).
- 119 Shinkai, S. et al. Physical activity and immune senescence in men. *Med Sci Sports Exerc* **27**, 1516-1526 (1995).
- 120 Campbell, P. T. et al. Effect of exercise on in vitro immune function: a 12-month randomized, controlled trial among postmenopausal women. *J Appl Physiol* (1985) **104**, 1648-1655, doi:10.1152/japplphysiol.01349.2007 (2008).
- 121 Levy, S. M. et al. Persistently low natural killer cell activity, age, and environmental stress as predictors of infectious morbidity. *Nat Immun Cell Growth Regul* **10**, 289-307 (1991).
- 122 Ogata, K. et al. Natural killer cells in the late decades of human life. *Clin Immunol Immunopathol* **84**, 269-275, doi:10.1006/clin.1997.4401 (1997).
- 123 Ogata, K. et al. Association between natural killer cell activity and infection in immunologically normal elderly people. *Clin Exp Immunol* **124**, 392-397, doi:10.1046/j.1365-2249.2001.01571.x (2001).
- 124 Jacinto, T. A. et al. Increased ROS production and DNA damage in monocytes are biomarkers of aging and atherosclerosis. *Biol Res* **51**, 33, doi:10.1186/s40659-018-0182-7 (2018).
- 125 Merino, A., Buendia, P., Martin-Malo, A., Aljama, P., Ramirez, R. & Carracedo, J. Senescent CD14+CD16+ monocytes exhibit proinflammatory and proatherosclerotic activity. *J Immunol* **186**, 1809-1815, doi:10.4049/jimmunol.1001866 (2011).
- 126 Shchukina, I. et al. Enhanced epigenetic profiling of classical human monocytes reveals a specific signature of healthy aging in the DNA methylome. *Nat Aging* **1**, 124-141, doi:10.1038/s43587-020-00002-6 (2021).
- 127 Mytych, J. et al. Towards Age-Related Anti-Inflammatory Therapy: Klotho Suppresses Activation of ER and Golgi Stress Response in Senescent Monocytes. *Cells* **9**, doi:10.3390/cells9020261 (2020).
- 128 Ding, J. et al. The association between aging-related monocyte transcriptional networks and comorbidity burden: the Multi-Ethnic Study of Atherosclerosis (MESA). *Geroscience* **45**, 197-207, doi:10.1007/s11357-022-00608-1 (2023).
- 129 Goodridge, H. S. Aging of classical monocyte subsets. *Aging (Albany NY)* **15**, 290-292, doi:10.18632/aging.204493 (2023).
- 130 Seidler, S., Zimmermann, H. W., Bartneck, M., Trautwein, C. & Tacke, F. Age-dependent alterations of monocyte subsets and monocyte-related chemokine pathways in healthy adults. *BMC Immunol* **11**, 30, doi:10.1186/1471-2172-11-30 (2010).
- 131 Wang, C. et al. Transcriptional characteristics and functional validation of three monocyte subsets during aging. *Immun Ageing* **20**, 50, doi:10.1186/s12979-023-00377-1 (2023).

- 132 Puchta, A. et al. TNF Drives Monocyte Dysfunction with Age and Results in Impaired Anti-pneumococcal Immunity. *PLoS Pathog* **12**, e1005368, doi:10.1371/journal.ppat.1005368 (2016).
- 133 Nakamura, S., Mori, K., Okuma, H., Sekine, T., Miyazaki, A. & Tsuchiya, K. Age-associated decline of monocyte insulin sensitivity in diabetic and healthy individuals. *Diab Vasc Dis Res* **18**, 1479164121989281, doi:10.1177/1479164121989281 (2021).
- 134 Walrand, S., Guillet, C., Boirie, Y. & Vasson, M. P. Insulin differentially regulates monocyte and polymorphonuclear neutrophil functions in healthy young and elderly humans. *J Clin Endocrinol Metab* **91**, 2738-2748, doi:10.1210/jc.2005-1619 (2006).
- 135 De Silva, N. S. et al. Nuclear envelope disruption triggers hallmarks of aging in lung alveolar macrophages. *Nat Aging* **3**, 1251-1268, doi:10.1038/s43587-023-00488-w (2023).
- 136 Khalil, H. et al. Aging is associated with hypermethylation of autophagy genes in macrophages. *Epigenetics* **11**, 381-388, doi:10.1080/15592294.2016.1144007 (2016).
- 137 Wang, H. et al. BRD4 contributes to LPS-induced macrophage senescence and promotes progression of atherosclerosis-associated lipid uptake. *Aging (Albany NY)* **12**, 9240-9259, doi:10.18632/aging.103200 (2020).
- 138 Wong, C. K., Smith, C. A., Sakamoto, K., Kaminski, N., Koff, J. L. & Goldstein, D. R. Aging Impairs Alveolar Macrophage Phagocytosis and Increases Influenza-Induced Mortality in Mice. *J Immunol* **199**, 1060-1068, doi:10.4049/jimmunol.1700397 (2017).
- 139 Song, Y., Shen, H., Du, W. & Goldstein, D. R. Inhibition of x-box binding protein 1 reduces tunicamycin-induced apoptosis in aged murine macrophages. *Aging Cell* **12**, 794-801, doi:10.1111/acel.12105 (2013).
- 140 Stranks, A. J. et al. Autophagy Controls Acquisition of Aging Features in Macrophages. *J Innate Immun* **7**, 375-391, doi:10.1159/000370112 (2015).
- 141 Agius, E. et al. Decreased TNF-alpha synthesis by macrophages restricts cutaneous immunosurveillance by memory CD4+ T cells during aging. *J Exp Med* **206**, 1929-1940, doi:10.1084/jem.20090896 (2009).
- 142 Aprahamian, T., Takemura, Y., Goukassian, D. & Walsh, K. Ageing is associated with diminished apoptotic cell clearance in vivo. *Clin Exp Immunol* **152**, 448-455, doi:10.1111/j.1365-2249.2008.03658.x (2008).
- 143 Arnardottir, H. H., Dalli, J., Colas, R. A., Shinohara, M. & Serhan, C. N. Aging delays resolution of acute inflammation in mice: reprogramming the host response with novel nano-proresolving medicines. *J Immunol* **193**, 4235-4244, doi:10.4049/jimmunol.1401313 (2014).
- 144 Thevaranjan, N. et al. Age-Associated Microbial Dysbiosis Promotes Intestinal Permeability, Systemic Inflammation, and Macrophage Dysfunction. *Cell Host Microbe* **21**, 455-466 e454, doi:10.1016/j.chom.2017.03.002 (2017).
- 145 Ahmadi, S. et al. A human-origin probiotic cocktail ameliorates aging-related leaky gut and inflammation via modulating the microbiota/taurine/tight junction axis. *JCI Insight* **5**, doi:10.1172/jci.insight.132055 (2020).
- 146 Kuhn, F. et al. Intestinal alkaline phosphatase targets the gut barrier to prevent aging. *JCI Insight* **5**, doi:10.1172/jci.insight.134049 (2020).
- 147 Minhas, P. S. et al. Macrophage de novo NAD(+) synthesis specifies immune function in aging and inflammation. *Nat Immunol* **20**, 50-63, doi:10.1038/s41590-018-0255-3 (2019).
- 148 Tannahill, G. M. et al. Succinate is an inflammatory signal that induces IL-1beta through HIF-1alpha. *Nature* **496**, 238-242, doi:10.1038/nature11986 (2013).
- 149 Hall, B. M. et al. Aging of mice is associated with p16(INK4a)- and beta-galactosidase-positive macrophage accumulation that can be induced in young mice by senescent cells. *Aging (Albany NY)* **8**, 1294-1315, doi:10.18632/aging.100991 (2016).
- 150 Childs, B. G., Baker, D. J., Wijshake, T., Conover, C. A., Campisi, J. & van Deursen, J. M. Senescent intimal foam cells are deleterious at all stages of atherosclerosis. *Science* **354**, 472-477, doi:10.1126/science.aaf6659 (2016).

- 151 Cudejko, C. *et al.* p16INK4a deficiency promotes IL-4-induced polarization and inhibits proinflammatory signaling in macrophages. *Blood* **118**, 2556-2566, doi:10.1182/blood-2010-10-313106 (2011).
- 152 Lu, R. J. *et al.* Multi-omic profiling of primary mouse neutrophils predicts a pattern of sex and age-related functional regulation. *Nat Aging* **1**, 715-733, doi:10.1038/s43587-021-00086-8 (2021).
- 153 Pereira, F. S. M. *et al.* Association Between the Length of Leukocyte Telomeres and Functional Performance of Older Adults: Observational Study. *Rejuvenation Res* **27**, 44-50, doi:10.1089/rej.2023.0050 (2024).
- 154 Xu, F. *et al.* Aging-related Atg5 defect impairs neutrophil extracellular traps formation. *Immunology* **151**, 417-432, doi:10.1111/imm.12740 (2017).
- 155 Lagnado, A. *et al.* Neutrophils induce paracrine telomere dysfunction and senescence in ROS-dependent manner. *EMBO J* **40**, e106048, doi:10.15252/embj.2020106048 (2021).
- 156 Fulop, T., Jr. *et al.* Changes in apoptosis of human polymorphonuclear granulocytes with aging. *Mech Ageing Dev* **96**, 15-34, doi:10.1016/s0047-6374(96)01881-7 (1997).
- 157 Barkaway, A. *et al.* Age-related changes in the local milieu of inflamed tissues cause aberrant neutrophil trafficking and subsequent remote organ damage. *Immunity* **54**, 1494-1510 e1497, doi:10.1016/j.jimmuni.2021.04.025 (2021).
- 158 Zhang, D. *et al.* Neutrophil ageing is regulated by the microbiome. *Nature* **525**, 528-532, doi:10.1038/nature15367 (2015).
- 159 Burgess, W. *et al.* The immune-endocrine loop during aging: role of growth hormone and insulin-like growth factor-I. *Neuroimmunomodulation* **6**, 56-68, doi:10.1159/000026365 (1999).
- 160 Pastorek, M. *et al.* Mitochondria-induced formation of neutrophil extracellular traps is enhanced in the elderly via Toll-like receptor 9. *J Leukoc Biol* **114**, 651-665, doi:10.1093/jleuko/qiad101 (2023).
- 161 Li, C. J. *et al.* Senescent immune cells release grancalcin to promote skeletal aging. *Cell Metab* **33**, 1957-1973 e1956, doi:10.1016/j.cmet.2021.08.009 (2021).
- 162 Mollica, L., Fleury, I., Belisle, C., Provost, S., Roy, D. C. & Busque, L. No association between telomere length and blood cell counts in elderly individuals. *J Gerontol A Biol Sci Med Sci* **64**, 965-967, doi:10.1093/gerona/glp065 (2009).
- 163 Allan, H. E., Vadgama, A., Armstrong, P. C. & Warner, T. D. What can we learn from senescent platelets, their transcriptomes and proteomes? *Platelets* **34**, 2200838, doi:10.1080/09537104.2023.2200838 (2023).
- 164 Daniele, S. *et al.* alpha-Synuclein Aggregated with Tau and beta-Amyloid in Human Platelets from Healthy Subjects: Correlation with Physical Exercise. *Front Aging Neurosci* **10**, 17, doi:10.3389/fnagi.2018.00017 (2018).
- 165 de Sousa, D. M. B. *et al.* The platelet transcriptome and proteome in Alzheimer's disease and aging: an exploratory cross-sectional study. *Front Mol Biosci* **10**, 1196083, doi:10.3389/fmoleb.2023.1196083 (2023).
- 166 Park, C. *et al.* Platelet factors are induced by longevity factor klotho and enhance cognition in young and aging mice. *Nat Aging* **3**, 1067-1078, doi:10.1038/s43587-023-00468-0 (2023).
- 167 Giani, E., Masi, I. & Galli, C. Platelets from aged rats aggregate more, but are more sensitive to prostacyclin. *Prostaglandins Leukot Med* **20**, 237-246, doi:10.1016/0262-1746(85)90145-3 (1985).
- 168 Gleerup, G. & Winther, K. The effect of ageing on human platelet sensitivity to serotonin. *Eur J Clin Invest* **18**, 504-506, doi:10.1111/j.1365-2362.1988.tb01047.x (1988).
- 169 Davison-Castillo, P. *et al.* TNF-alpha-driven inflammation and mitochondrial dysfunction define the platelet hyperreactivity of aging. *Blood* **134**, 727-740, doi:10.1182/blood.2019000200 (2019).

- 170 Oleksowicz, L., Mrowiec, Z., Zuckerman, D., Isaacs, R., Dutcher, J. & Puszkin, E. Platelet activation induced by interleukin-6: evidence for a mechanism involving arachidonic acid metabolism. *Thromb Haemost* **72**, 302-308 (1994).
- 171 Corica, F. et al. Reduced intraplatelet magnesium concentrations in elderly patients with non-insulin dependent diabetes mellitus (NIDDM). *Arch Gerontol Geriatr* **25**, 255-262, doi:10.1016/s0167-4943(97)00018-6 (1997).
- 172 Fisar, Z., Hroudova, J., Zverova, M., Jirak, R., Raboch, J. & Kitzlerova, E. Age-Dependent Alterations in Platelet Mitochondrial Respiration. *Biomedicines* **11**, doi:10.3390/biomedicines11061564 (2023).
- 173 Pascabio, D. M., Worthington, A. K., Smith-Berdan, S. & Forsberg, E. C. Megakaryocyte progenitor cell function is enhanced upon aging despite the functional decline of aged hematopoietic stem cells. *Stem Cell Reports* **16**, 1598-1613, doi:10.1016/j.stemcr.2021.04.016 (2021).
- 174 Kasagi, F., Yamada, M., Sasaki, H. & Fujita, S. Biologic score and mortality based on a 30-year mortality follow-up: radiation effects research foundation adult health study. *J Gerontol A Biol Sci Med Sci* **64**, 865-870, doi:10.1093/gerona/glp025 (2009).
- 175 Delbosc, S. et al. Erythrocyte Efferocytosis by the Arterial Wall Promotes Oxidation in Early-Stage Atheroma in Humans. *Front Cardiovasc Med* **4**, 43, doi:10.3389/fcvm.2017.00043 (2017).
- 176 Tripathi, S. S., Kumar, R., Bissoyi, A. & Rizvi, S. I. Baicalein May Act as a Caloric Restriction Mimetic Candidate to Improve the Antioxidant Profile in a Natural Rodent Model of Aging. *Rejuvenation Res* **25**, 70-78, doi:10.1089/rej.2021.0071 (2022).
- 177 Kim, D., Won, C. W. & Park, Y. Association Between Erythrocyte Levels of n-3 Polyunsaturated Fatty Acids and Risk of Frailty in Community-Dwelling Older Adults: The Korean Frailty and Aging Cohort Study. *J Gerontol A Biol Sci Med Sci* **76**, 499-504, doi:10.1093/gerona/glaa042 (2021).
- 178 Klei, T. R. L. et al. The Gardos effect drives erythrocyte senescence and leads to Lu/BCAM and CD44 adhesion molecule activation. *Blood Adv* **4**, 6218-6229, doi:10.1182/bloodadvances.2020003077 (2020).
- 179 Schwarz, A., Maeda, A. & Schwarz, T. Alteration of the migratory behavior of UV-induced regulatory T cells by tissue-specific dendritic cells. *J Immunol* **178**, 877-886, doi:10.4049/jimmunol.178.2.877 (2007).
- 180 Yamazaki, S. et al. Ultraviolet B-Induced Maturation of CD11b-Type Langerin(-) Dendritic Cells Controls the Expansion of Foxp3(+) Regulatory T Cells in the Skin. *J Immunol* **200**, 119-129, doi:10.4049/jimmunol.1701056 (2018).
- 181 Anderson, R. et al. eIF2A-knockout mice reveal decreased life span and metabolic syndrome. *FASEB J* **35**, e21990, doi:10.1096/fj.202101105R (2021).
- 182 Wong, C. P., Magnusson, K. R. & Ho, E. Aging is associated with altered dendritic cells subset distribution and impaired proinflammatory cytokine production. *Exp Gerontol* **45**, 163-169, doi:10.1016/j.exger.2009.11.005 (2010).
- 183 Agrawal, A., Agrawal, S., Cao, J. N., Su, H., Osann, K. & Gupta, S. Altered innate immune functioning of dendritic cells in elderly humans: a role of phosphoinositide 3-kinase-signaling pathway. *J Immunol* **178**, 6912-6922, doi:10.4049/jimmunol.178.11.6912 (2007).
- 184 Bashir, H., Singh, S., Singh, R. P., Agrewala, J. N. & Kumar, R. Age-mediated gut microbiota dysbiosis promotes the loss of dendritic cells tolerance. *Aging Cell* **22**, e13838, doi:10.1111/ace.13838 (2023).
- 185 Hernandez-Garcia, E. et al. Conventional type 1 dendritic cells protect against age-related adipose tissue dysfunction and obesity. *Cell Mol Immunol* **19**, 260-275, doi:10.1038/s41423-021-00812-7 (2022).

- 186 Chouquet, C. A. et al. Loss of Phagocytic and Antigen Cross-Presenting Capacity in Aging Dendritic Cells Is Associated with Mitochondrial Dysfunction. *J Immunol* **195**, 2624-2632, doi:10.4049/jimmunol.1501006 (2015).
- 187 Elsayed, R. et al. Microbially-Induced Exosomes from Dendritic Cells Promote Paracrine Immune Senescence: Novel Mechanism of Bone Degenerative Disease in Mice. *Aging Dis* **14**, 136-151, doi:10.14336/AD.2022.0623 (2023).
- 188 Xiao, J., Zhou, H., Wu, N. & Wu, L. The non-canonical Wnt pathway negatively regulates dendritic cell differentiation by inhibiting the expansion of Flt3(+) lymphocyte-primed multipotent precursors. *Cell Mol Immunol* **13**, 593-604, doi:10.1038/cmi.2015.39 (2016).
- 189 Matsumoto, T., Baker, D. J., d'Uscio, L. V., Mozammel, G., Katusic, Z. S. & van Deursen, J. M. Aging-associated vascular phenotype in mutant mice with low levels of BubR1. *Stroke* **38**, 1050-1056, doi:10.1161/01.STR.0000257967.86132.01 (2007).
- 190 Morgan, R. G. et al. Induced Trf2 deletion leads to aging vascular phenotype in mice associated with arterial telomere uncapping, senescence signaling, and oxidative stress. *J Mol Cell Cardiol* **127**, 74-82, doi:10.1016/j.jmcc.2018.11.014 (2019).
- 191 Olive, M. et al. Cardiovascular pathology in Hutchinson-Gilford progeria: correlation with the vascular pathology of aging. *Arterioscler Thromb Vasc Biol* **30**, 2301-2309, doi:10.1161/ATVBAHA.110.209460 (2010).
- 192 Xu, S. et al. SIRT6 protects against endothelial dysfunction and atherosclerosis in mice. *Aging (Albany NY)* **8**, 1064-1082, doi:10.18632/aging.100975 (2016).
- 193 Madrigal-Matute, J. et al. Protective role of chaperone-mediated autophagy against atherosclerosis. *Proc Natl Acad Sci U S A* **119**, e2121133119, doi:10.1073/pnas.2121133119 (2022).
- 194 Henderson, J. M., Weber, C. & Santovito, D. Beyond Self-Recycling: Cell-Specific Role of Autophagy in Atherosclerosis. *Cells* **10**, doi:10.3390/cells10030625 (2021).
- 195 Eisenberg, T. et al. Cardioprotection and lifespan extension by the natural polyamine spermidine. *Nat Med* **22**, 1428-1438, doi:10.1038/nm.4222 (2016).
- 196 LaRocca, T. J., Goscia-Ryan, R. A., Hearon, C. M., Jr. & Seals, D. R. The autophagy enhancer spermidine reverses arterial aging. *Mech Ageing Dev* **134**, 314-320, doi:10.1016/j.mad.2013.04.004 (2013).
- 197 Hsieh, P. N. et al. A conserved KLF-autophagy pathway modulates nematode lifespan and mammalian age-associated vascular dysfunction. *Nat Commun* **8**, 914, doi:10.1038/s41467-017-00899-5 (2017).
- 198 Lesniewski, L. A. et al. Dietary rapamycin supplementation reverses age-related vascular dysfunction and oxidative stress, while modulating nutrient-sensing, cell cycle, and senescence pathways. *Aging Cell* **16**, 17-26, doi:10.1111/acel.12524 (2017).
- 199 Kiss, T. et al. Old blood from heterochronic parabionts accelerates vascular aging in young mice: transcriptomic signature of pathologic smooth muscle remodeling. *Geroscience* **44**, 953-981, doi:10.1007/s11357-022-00519-1 (2022).
- 200 Kiss, T. et al. Circulating anti-geronic factors from heterochronic parabionts promote vascular rejuvenation in aged mice: transcriptional footprint of mitochondrial protection, attenuation of oxidative stress, and rescue of endothelial function by young blood. *Geroscience* **42**, 727-748, doi:10.1007/s11357-020-00180-6 (2020).
- 201 Ballak, D. B. et al. Short-term interleukin-37 treatment improves vascular endothelial function, endurance exercise capacity, and whole-body glucose metabolism in old mice. *Aging Cell* **19**, e13074, doi:10.1111/acel.13074 (2020).
- 202 Williams, B. et al. 2018 ESC/ESH Guidelines for the management of arterial hypertension. *Eur Heart J* **39**, 3021-3104, doi:10.1093/eurheartj/ehy339 (2018).
- 203 Gocmez, S. S. et al. Etanercept improves aging-induced cognitive deficits by reducing inflammation and vascular dysfunction in rats. *Physiol Behav* **224**, 113019, doi:10.1016/j.physbeh.2020.113019 (2020).

- 204 Marin-Aguilar, F. *et al.* NLRP3 inflammasome suppression improves longevity and prevents cardiac aging in male mice. *Aging Cell* **19**, e13050, doi:10.1111/acel.13050 (2020).
- 205 Saeedi Saravi, S. S. *et al.* Long-term dietary n3 fatty acid prevents aging-related cardiac diastolic and vascular dysfunction. *Vascul Pharmacol* **150**, 107175, doi:10.1016/j.vph.2023.107175 (2023).
- 206 Touyz, R. M. Gut Dysbiosis-Induced Hypertension Is Ameliorated by Intermittent Fasting. *Circ Res* **128**, 1255-1257, doi:10.1161/CIRCRESAHA.121.319147 (2021).
- 207 Colman, R. J. *et al.* Caloric restriction delays disease onset and mortality in rhesus monkeys. *Science* **325**, 201-204, doi:10.1126/science.1173635 (2009).
- 208 Holloszy, J. O. & Fontana, L. Caloric restriction in humans. *Exp Gerontol* **42**, 709-712, doi:10.1016/j.exger.2007.03.009 (2007).
- 209 Goldhamer, A., Lisle, D., Parpia, B., Anderson, S. V. & Campbell, T. C. Medically supervised water-only fasting in the treatment of hypertension. *J Manipulative Physiol Ther* **24**, 335-339, doi:10.1067/mmmt.2001.115263 (2001).
- 210 Foote, K. *et al.* Restoring mitochondrial DNA copy number preserves mitochondrial function and delays vascular aging in mice. *Aging Cell* **17**, e12773, doi:10.1111/acel.12773 (2018).
- 211 Abdellatif, M. *et al.* Nicotinamide for the treatment of heart failure with preserved ejection fraction. *Sci Transl Med* **13**, doi:10.1126/scitranslmed.abd7064 (2021).
- 212 Oshima, J., Sidorova, J. M. & Monnat, R. J., Jr. Werner syndrome: Clinical features, pathogenesis and potential therapeutic interventions. *Ageing Res Rev* **33**, 105-114, doi:10.1016/j.arr.2016.03.002 (2017).
- 213 Huang, S. *et al.* The spectrum of WRN mutations in Werner syndrome patients. *Hum Mutat* **27**, 558-567, doi:10.1002/humu.20337 (2006).
- 214 Torisu, K. *et al.* Intact endothelial autophagy is required to maintain vascular lipid homeostasis. *Aging Cell* **15**, 187-191, doi:10.1111/acel.12423 (2016).
- 215 Sladitschek-Martens, H. L. *et al.* YAP/TAZ activity in stromal cells prevents ageing by controlling cGAS-STING. *Nature* **607**, 790-798, doi:10.1038/s41586-022-04924-6 (2022).
- 216 Raber, L. *et al.* Effect of Alirocumab Added to High-Intensity Statin Therapy on Coronary Atherosclerosis in Patients With Acute Myocardial Infarction: The PACMAN-AMI Randomized Clinical Trial. *JAMA* **327**, 1771-1781, doi:10.1001/jama.2022.5218 (2022).
- 217 Ridker, P. M. *et al.* Effect of interleukin-1beta inhibition with canakinumab on incident lung cancer in patients with atherosclerosis: exploratory results from a randomised, double-blind, placebo-controlled trial. *Lancet* **390**, 1833-1842, doi:10.1016/S0140-6736(17)32247-X (2017).
- 218 Ridker, P. M. *et al.* Antiinflammatory Therapy with Canakinumab for Atherosclerotic Disease. *N Engl J Med* **377**, 1119-1131, doi:10.1056/NEJMoa1707914 (2017).
- 219 Kiouptsi, K. *et al.* The Microbiota Promotes Arterial Thrombosis in Low-Density Lipoprotein Receptor-Deficient Mice. *mBio* **10**, doi:10.1128/mBio.02298-19 (2019).
- 220 Lindskog Jonsson, A. *et al.* Impact of Gut Microbiota and Diet on the Development of Atherosclerosis in Apoe(-/-) Mice. *Arterioscler Thromb Vasc Biol* **38**, 2318-2326, doi:10.1161/ATVBAHA.118.311233 (2018).
- 221 Schriner, S. E. *et al.* Extension of murine life span by overexpression of catalase targeted to mitochondria. *Science* **308**, 1909-1911, doi:10.1126/science.1106653 (2005).
- 222 Rauscher, F. M. *et al.* Aging, progenitor cell exhaustion, and atherosclerosis. *Circulation* **108**, 457-463, doi:10.1161/01.CIR.0000082924.75945.48 (2003).
- 223 Bhayadia, R., Schmidt, B. M., Melk, A. & Homme, M. Senescence-Induced Oxidative Stress Causes Endothelial Dysfunction. *J Gerontol A Biol Sci Med Sci* **71**, 161-169, doi:10.1093/gerona/glv008 (2016).
- 224 Liao, W. L. *et al.* Brahma-related gene 1 inhibits proliferation and migration of human aortic smooth muscle cells by directly up-regulating Ras-related associated with diabetes in the pathophysiological processes of aortic dissection. *J Thorac Cardiovasc Surg* **150**, 1292-1301 e1292, doi:10.1016/j.jtcvs.2015.08.010 (2015).

- 225 Davies, H. A. *et al.* Idiopathic degenerative thoracic aneurysms are associated with increased aortic medial amyloid. *Amyloid* **26**, 148-155, doi:10.1080/13506129.2019.1625323 (2019).
- 226 Osonoi, Y. *et al.* Defective autophagy in vascular smooth muscle cells enhances cell death and atherosclerosis. *Autophagy* **14**, 1991-2006, doi:10.1080/15548627.2018.1501132 (2018).
- 227 Isselbacher, E. M. *et al.* 2022 ACC/AHA Guideline for the Diagnosis and Management of Aortic Disease: A Report of the American Heart Association/American College of Cardiology Joint Committee on Clinical Practice Guidelines. *Circulation* **146**, e334-e482, doi:10.1161/CIR.0000000000001106 (2022).
- 228 Ding, Y. N. *et al.* SIRT6 is an epigenetic repressor of thoracic aortic aneurysms via inhibiting inflammation and senescence. *Signal Transduct Target Ther* **8**, 255, doi:10.1038/s41392-023-01456-x (2023).
- 229 Tian, Z. *et al.* Gut microbiome dysbiosis contributes to abdominal aortic aneurysm by promoting neutrophil extracellular trap formation. *Cell Host Microbe* **30**, 1450-1463 e1458, doi:10.1016/j.chom.2022.09.004 (2022).
- 230 Oller, J. *et al.* Extracellular Tuning of Mitochondrial Respiration Leads to Aortic Aneurysm. *Circulation* **143**, 2091-2109, doi:10.1161/CIRCULATIONAHA.120.051171 (2021).
- 231 Ocampo, A. *et al.* In Vivo Amelioration of Age-Associated Hallmarks by Partial Reprogramming. *Cell* **167**, 1719-1733 e1712, doi:10.1016/j.cell.2016.11.052 (2016).
- 232 Beckmann, B. M. *et al.* Laminopathy presenting as familial atrial fibrillation. *Int J Cardiol* **145**, 394-396, doi:10.1016/j.ijcard.2010.04.024 (2010).
- 233 Saj, M. *et al.* Variants of the lamin A/C (LMNA) gene in non-valvular atrial fibrillation patients: a possible pathogenic role of the Thr528Met mutation. *Mol Diagn Ther* **16**, 99-107, doi:10.1007/BF03256434 (2012).
- 234 Wiersma, M. *et al.* Endoplasmic Reticulum Stress Is Associated With Autophagy and Cardiomyocyte Remodeling in Experimental and Human Atrial Fibrillation. *J Am Heart Assoc* **6**, doi:10.1161/JAHA.117.006458 (2017).
- 235 Barbhaiya, C. R. *et al.* Electrophysiologic assessment of conduction abnormalities and atrial arrhythmias associated with amyloid cardiomyopathy. *Heart Rhythm* **13**, 383-390, doi:10.1016/j.hrthm.2015.09.016 (2016).
- 236 Xiao, H. D. *et al.* Mice with cardiac-restricted angiotensin-converting enzyme (ACE) have atrial enlargement, cardiac arrhythmia, and sudden death. *Am J Pathol* **165**, 1019-1032, doi:10.1016/S0002-9440(10)63363-9 (2004).
- 237 Liao, J. *et al.* Interleukin-6-Mediated-Ca(2+) Handling Abnormalities Contributes to Atrial Fibrillation in Sterile Pericarditis Rats. *Front Immunol* **12**, 758157, doi:10.3389/fimmu.2021.758157 (2021).
- 238 Gollob, M. H. *et al.* Somatic mutations in the connexin 40 gene (GJA5) in atrial fibrillation. *N Engl J Med* **354**, 2677-2688, doi:10.1056/NEJMoa052800 (2006).
- 239 Hodgson-Zingman, D. M. *et al.* Atrial natriuretic peptide frameshift mutation in familial atrial fibrillation. *N Engl J Med* **359**, 158-165, doi:10.1056/NEJMoa0706300 (2008).
- 240 Healey, J. S. *et al.* Prevention of atrial fibrillation with angiotensin-converting enzyme inhibitors and angiotensin receptor blockers: a meta-analysis. *J Am Coll Cardiol* **45**, 1832-1839, doi:10.1016/j.jacc.2004.11.070 (2005).
- 241 Saba, S. *et al.* Atrial contractile dysfunction, fibrosis, and arrhythmias in a mouse model of cardiomyopathy secondary to cardiac-specific overexpression of tumor necrosis factor-alpha. *Am J Physiol Heart Circ Physiol* **289**, H1456-1467, doi:10.1152/ajpheart.00733.2004 (2005).
- 242 Correction to: Enhanced Cardiomyocyte NLRP3 Inflammasome Signaling Promotes Atrial Fibrillation. *Circulation* **139**, e889, doi:10.1161/CIR.0000000000000694 (2019).
- 243 Zhang, Y. *et al.* Gut microbiota dysbiosis promotes age-related atrial fibrillation by lipopolysaccharide and glucose-induced activation of NLRP3-inflammasome. *Cardiovasc Res* **118**, 785-797, doi:10.1093/cvr/cvab114 (2022).

- 244 Chan, Y. H. *et al.* Atrial fibrillation and its arrhythmogenesis associated with insulin resistance. *Cardiovasc Diabetol* **18**, 125, doi:10.1186/s12933-019-0928-8 (2019).
- 245 Liu, C. *et al.* Mitochondrial Dysfunction Contributes to Aging-Related Atrial Fibrillation. *Oxid Med Cell Longev* **2021**, 5530293, doi:10.1155/2021/5530293 (2021).
- 246 Cook, A. & Giunti, P. Friedreich's ataxia: clinical features, pathogenesis and management. *Br Med Bull* **124**, 19-30, doi:10.1093/bmb/ldx034 (2017).
- 247 Limongelli, G., Tome-Esteban, M., Dejthevaporn, C., Rahman, S., Hanna, M. G. & Elliott, P. M. Prevalence and natural history of heart disease in adults with primary mitochondrial respiratory chain disease. *Eur J Heart Fail* **12**, 114-121, doi:10.1093/eurjhf/hfp186 (2010).
- 248 Triana-Martinez, F. *et al.* Identification and characterization of Cardiac Glycosides as senolytic compounds. *Nat Commun* **10**, 4731, doi:10.1038/s41467-019-12888-x (2019).
- 249 Yap, L. *et al.* Pluripotent stem cell-derived committed cardiac progenitors remuscularize damaged ischemic hearts and improve their function in pigs. *NPJ Regen Med* **8**, 26, doi:10.1038/s41536-023-00302-6 (2023).
- 250 North, B. J. *et al.* SIRT2 induces the checkpoint kinase BubR1 to increase lifespan. *EMBO J* **33**, 1438-1453, doi:10.15252/embj.201386907 (2014).
- 251 Kuo, H. C. *et al.* A defect in the Kv channel-interacting protein 2 (KChIP2) gene leads to a complete loss of I_(to) and confers susceptibility to ventricular tachycardia. *Cell* **107**, 801-813, doi:10.1016/s0092-8674(01)00588-8 (2001).
- 252 Matsumura, H. *et al.* H558R, a common SCN5A polymorphism, modifies the clinical phenotype of Brugada syndrome by modulating DNA methylation of SCN5A promoters. *J Biomed Sci* **24**, 91, doi:10.1186/s12929-017-0397-x (2017).
- 253 Schmidt, E. K., Mustonen, T., Kiuru-Enari, S., Kivela, T. T. & Atula, S. Finnish gelsolin amyloidosis causes significant disease burden but does not affect survival: FIN-GAR phase II study. *Orphanet J Rare Dis* **15**, 19, doi:10.1186/s13023-020-1300-5 (2020).
- 254 Byrne, S., Dionisi-Vici, C., Smith, L., Gautel, M. & Jungbluth, H. Vici syndrome: a review. *Orphanet J Rare Dis* **11**, 21, doi:10.1186/s13023-016-0399-x (2016).
- 255 Monnerat, G. *et al.* Macrophage-dependent IL-1beta production induces cardiac arrhythmias in diabetic mice. *Nat Commun* **7**, 13344, doi:10.1038/ncomms13344 (2016).
- 256 Li, D. S. *et al.* Knockout of interleukin-17A diminishes ventricular arrhythmia susceptibility in diabetic mice via inhibiting NF-kappaB-mediated electrical remodeling. *Acta Pharmacol Sin* **43**, 307-315, doi:10.1038/s41401-021-00659-8 (2022).
- 257 Meng, G. *et al.* Gut microbe-derived metabolite trimethylamine N-oxide activates the cardiac autonomic nervous system and facilitates ischemia-induced ventricular arrhythmia via two different pathways. *EBioMedicine* **44**, 656-664, doi:10.1016/j.ebiom.2019.03.066 (2019).
- 258 Sanchez, G. *et al.* High-Fat-Diet-Induced Obesity Produces Spontaneous Ventricular Arrhythmias and Increases the Activity of Ryanodine Receptors in Mice. *Int J Mol Sci* **19**, doi:10.3390/ijms19020533 (2018).
- 259 Tsou, A. Y. *et al.* Mortality in Friedreich ataxia. *J Neurol Sci* **307**, 46-49, doi:10.1016/j.jns.2011.05.023 (2011).
- 260 Baker, D. J. *et al.* Increased expression of BubR1 protects against aneuploidy and cancer and extends healthy lifespan. *Nat Cell Biol* **15**, 96-102, doi:10.1038/ncb2643 (2013).
- 261 Zhu, X. *et al.* Fine-Tuning of PGC1alpha Expression Regulates Cardiac Function and Longevity. *Circ Res* **125**, 707-719, doi:10.1161/CIRCRESAHA.119.315529 (2019).
- 262 Boon, R. A. *et al.* MicroRNA-34a regulates cardiac ageing and function. *Nature* **495**, 107-110, doi:10.1038/nature11919 (2013).
- 263 Brayson, D. *et al.* Prelamin A mediates myocardial inflammation in dilated and HIV-associated cardiomyopathies. *JCI Insight* **4**, doi:10.1172/jci.insight.126315 (2019).
- 264 Sharifi-Sanjani, M. *et al.* Cardiomyocyte-Specific Telomere Shortening is a Distinct Signature of Heart Failure in Humans. *J Am Heart Assoc* **6**, doi:10.1161/JAHA.116.005086 (2017).

- 265 Jeong, M. Y. *et al.* Histone deacetylase activity governs diastolic dysfunction through a nongenomic mechanism. *Sci Transl Med* **10**, doi:10.1126/scitranslmed.aao0144 (2018).
- 266 Lin, Y. H. *et al.* HDAC6 modulates myofibril stiffness and diastolic function of the heart. *J Clin Invest* **132**, doi:10.1172/JCI148333 (2022).
- 267 Pyo, J. O. *et al.* Overexpression of Atg5 in mice activates autophagy and extends lifespan. *Nat Commun* **4**, 2300, doi:10.1038/ncomms3300 (2013).
- 268 Blackwood, E. A. *et al.* ATF6 Regulates Cardiac Hypertrophy by Transcriptional Induction of the mTORC1 Activator, Rheb. *Circ Res* **124**, 79-93, doi:10.1161/CIRCRESAHA.118.313854 (2019).
- 269 Maurer, M. S. *et al.* Tafamidis Treatment for Patients with Transthyretin Amyloid Cardiomyopathy. *N Engl J Med* **379**, 1007-1016, doi:10.1056/NEJMoa1805689 (2018).
- 270 Quarles, E. *et al.* Rapamycin persistently improves cardiac function in aged, male and female mice, even following cessation of treatment. *Aging Cell* **19**, e13086, doi:10.1111/acel.13086 (2020).
- 271 Fernandez, A. F. *et al.* Disruption of the beclin 1-BCL2 autophagy regulatory complex promotes longevity in mice. *Nature* **558**, 136-140, doi:10.1038/s41586-018-0162-7 (2018).
- 272 Flynn, J. M. *et al.* Late-life rapamycin treatment reverses age-related heart dysfunction. *Aging Cell* **12**, 851-862, doi:10.1111/acel.12109 (2013).
- 273 Miyata, R., Hayashi, M. & Itoh, E. Pathological changes in cardiac muscle and cerebellar cortex in Vici syndrome. *Am J Med Genet A* **164A**, 3203-3205, doi:10.1002/ajmg.a.36753 (2014).
- 274 Anker, S. D. *et al.* Empagliflozin in Heart Failure with a Preserved Ejection Fraction. *N Engl J Med* **385**, 1451-1461, doi:10.1056/NEJMoa2107038 (2021).
- 275 Pantiya, P. *et al.* Chronic D-galactose administration induces natural aging characteristics, in rat's brain and heart. *Toxicology* **492**, 153553, doi:10.1016/j.tox.2023.153553 (2023).
- 276 Zhang, H., Alder, N. N., Wang, W., Szeto, H., Marcinek, D. J. & Rabinovitch, P. S. Reduction of elevated proton leak rejuvenates mitochondria in the aged cardiomyocyte. *eLife* **9**, doi:10.7554/eLife.60827 (2020).
- 277 Gevaert, A. B. *et al.* Endothelial Senescence Contributes to Heart Failure With Preserved Ejection Fraction in an Aging Mouse Model. *Circ Heart Fail* **10**, doi:10.1161/CIRCHEARTFAILURE.116.003806 (2017).
- 278 Kurz, D. J. *et al.* Degenerative aortic valve stenosis, but not coronary disease, is associated with shorter telomere length in the elderly. *Arterioscler Thromb Vasc Biol* **26**, e114-117, doi:10.1161/01.ATV.0000222961.24912.69 (2006).
- 279 Gosev, I. *et al.* Epigenome alterations in aortic valve stenosis and its related left ventricular hypertrophy. *Clin Epigenetics* **9**, 106, doi:10.1186/s13148-017-0406-7 (2017).
- 280 Salah, S., Hegazy, R., Ammar, R., Sheba, H. & Abdelrahman, L. MEFV gene mutations and cardiac phenotype in children with familial Mediterranean fever: a cohort study. *Pediatr Rheumatol Online J* **12**, 5, doi:10.1186/1546-0096-12-5 (2014).
- 281 Xiong, J. *et al.* A Metabolic Basis for Endothelial-to-Mesenchymal Transition. *Mol Cell* **69**, 689-698 e687, doi:10.1016/j.molcel.2018.01.010 (2018).
- 282 Oh, K. S. *et al.* Cellular senescence evaluated by P16INK4a immunohistochemistry is a prevalent phenomenon in advanced calcific aortic valve disease. *Cardiovasc Pathol* **52**, 107318, doi:10.1016/j.carpath.2021.107318 (2021).
- 283 Matsumoto, Y. *et al.* Reduced number and function of endothelial progenitor cells in patients with aortic valve stenosis: a novel concept for valvular endothelial cell repair. *Eur Heart J* **30**, 346-355, doi:10.1093/eurheartj/ehn501 (2009).
- 284 Hartman, T. K., Wengenack, T. M., Poduslo, J. F. & van Deursen, J. M. Mutant mice with small amounts of BubR1 display accelerated age-related gliosis. *Neurobiol Aging* **28**, 921-927, doi:10.1016/j.neurobiolaging.2006.05.012 (2007).
- 285 Garcia-Castillo, H., Vasquez-Velasquez, A. I., Rivera, H. & Barros-Nunez, P. Clinical and genetic heterogeneity in patients with mosaic variegated aneuploidy: delineation of clinical subtypes. *Am J Med Genet A* **146A**, 1687-1695, doi:10.1002/ajmg.a.32315 (2008).

- 286 Jaskelioff, M. *et al.* Telomerase reactivation reverses tissue degeneration in aged telomerase-deficient mice. *Nature* **469**, 102-106, doi:10.1038/nature09603 (2011).
- 287 Bernardes de Jesus, B. *et al.* Telomerase gene therapy in adult and old mice delays aging and increases longevity without increasing cancer. *EMBO Mol Med* **4**, 691-704, doi:10.1002/emmm.201200245 (2012).
- 288 Yang, J. H. *et al.* Loss of epigenetic information as a cause of mammalian aging. *Cell* **186**, 305-326 e327, doi:10.1016/j.cell.2022.12.027 (2023).
- 289 Kumar, S. *et al.* MicroRNA-455-3p improves synaptic, cognitive functions and extends lifespan: Relevance to Alzheimer's disease. *Redox Biol* **48**, 102182, doi:10.1016/j.redox.2021.102182 (2021).
- 290 Pao, P. C. *et al.* HDAC1 modulates OGG1-initiated oxidative DNA damage repair in the aging brain and Alzheimer's disease. *Nat Commun* **11**, 2484, doi:10.1038/s41467-020-16361-y (2020).
- 291 Bobkova, N. V. *et al.* Exogenous Hsp70 delays senescence and improves cognitive function in aging mice. *Proc Natl Acad Sci U S A* **112**, 16006-16011, doi:10.1073/pnas.1516131112 (2015).
- 292 Hafycz, J. M., Strus, E. & Naidoo, N. Reducing ER stress with chaperone therapy reverses sleep fragmentation and cognitive decline in aged mice. *Aging Cell* **21**, e13598, doi:10.1111/ace.13598 (2022).
- 293 Degenhardt, K. *et al.* Medin aggregation causes cerebrovascular dysfunction in aging wild-type mice. *Proc Natl Acad Sci U S A* **117**, 23925-23931, doi:10.1073/pnas.2011133117 (2020).
- 294 Wirth, M. *et al.* The effect of spermidine on memory performance in older adults at risk for dementia: A randomized controlled trial. *Cortex* **109**, 181-188, doi:10.1016/j.cortex.2018.09.014 (2018).
- 295 Villeda, S. A. *et al.* Young blood reverses age-related impairments in cognitive function and synaptic plasticity in mice. *Nat Med* **20**, 659-663, doi:10.1038/nm.3569 (2014).
- 296 Villeda, S. A. *et al.* The ageing systemic milieu negatively regulates neurogenesis and cognitive function. *Nature* **477**, 90-94, doi:10.1038/nature10357 (2011).
- 297 Katsimpardi, L. *et al.* Vascular and neurogenic rejuvenation of the aging mouse brain by young systemic factors. *Science* **344**, 630-634, doi:10.1126/science.1251141 (2014).
- 298 Yang, S. *et al.* Chondroitin 6-sulphate is required for neuroplasticity and memory in ageing. *Mol Psychiatry* **26**, 5658-5668, doi:10.1038/s41380-021-01208-9 (2021).
- 299 Hajjar, I. *et al.* Effects of Candesartan vs Lisinopril on Neurocognitive Function in Older Adults With Executive Mild Cognitive Impairment: A Randomized Clinical Trial. *JAMA Netw Open* **3**, e2012252, doi:10.1001/jamanetworkopen.2020.122252 (2020).
- 300 Minhas, P. S. *et al.* Restoring metabolism of myeloid cells reverses cognitive decline in ageing. *Nature* **590**, 122-128, doi:10.1038/s41586-020-03160-0 (2021).
- 301 Fielder, E. *et al.* Anti-inflammatory treatment rescues memory deficits during aging in nfkb1(-/-) mice. *Aging Cell* **19**, e13188, doi:10.1111/ace.13188 (2020).
- 302 Lin, S. W. *et al.* Lactobacillus plantarum GKM3 Promotes Longevity, Memory Retention, and Reduces Brain Oxidation Stress in SAMP8 Mice. *Nutrients* **13**, doi:10.3390/nu13082860 (2021).
- 303 Boehme, M. *et al.* Microbiota from young mice counteracts selective age-associated behavioral deficits. *Nat Aging* **1**, 666-676, doi:10.1038/s43587-021-00093-9 (2021).
- 304 Singh, R. *et al.* Late-onset intermittent fasting dietary restriction as a potential intervention to retard age-associated brain function impairments in male rats. *Age (Dordr)* **34**, 917-933, doi:10.1007/s11357-011-9289-2 (2012).
- 305 Witte, A. V., Fobker, M., Gellner, R., Knecht, S. & Floel, A. Caloric restriction improves memory in elderly humans. *Proc Natl Acad Sci U S A* **106**, 1255-1260, doi:10.1073/pnas.0808587106 (2009).

- 306 Shabalina, I. G. *et al.* Improved health-span and lifespan in mtDNA mutator mice treated with the mitochondrially targeted antioxidant SkQ1. *Aging (Albany NY)* **9**, 315-339, doi:10.18632/aging.101174 (2017).
- 307 Tarantini, S. *et al.* Treatment with the mitochondrial-targeted antioxidant peptide SS-31 rescues neurovascular coupling responses and cerebrovascular endothelial function and improves cognition in aged mice. *Aging Cell* **17**, doi:10.1111/acel.12731 (2018).
- 308 Zhang, X. *et al.* Rejuvenation of the aged brain immune cell landscape in mice through p16-positive senescent cell clearance. *Nat Commun* **13**, 5671, doi:10.1038/s41467-022-33226-8 (2022).
- 309 Ogrodnik, M. *et al.* Whole-body senescent cell clearance alleviates age-related brain inflammation and cognitive impairment in mice. *Aging Cell* **20**, e13296, doi:10.1111/acel.13296 (2021).
- 310 Bussian, T. J., Aziz, A., Meyer, C. F., Swenson, B. L., van Deursen, J. M. & Baker, D. J. Clearance of senescent glial cells prevents tau-dependent pathology and cognitive decline. *Nature* **562**, 578-582, doi:10.1038/s41586-018-0543-y (2018).
- 311 Rodriguez-Matellan, A., Alcazar, N., Hernandez, F., Serrano, M. & Avila, J. In Vivo Reprogramming Ameliorates Aging Features in Dentate Gyrus Cells and Improves Memory in Mice. *Stem Cell Reports* **15**, 1056-1066, doi:10.1016/j.stemcr.2020.09.010 (2020).
- 312 El Hajjar, J. *et al.* Heterochromatic genome instability and neurodegeneration sharing similarities with Alzheimer's disease in old Bmi1+/- mice. *Sci Rep* **9**, 594, doi:10.1038/s41598-018-37444-3 (2019).
- 313 Thadathil, N., Delotterie, D. F., Xiao, J., Hori, R., McDonald, M. P. & Khan, M. M. DNA Double-Strand Break Accumulation in Alzheimer's Disease: Evidence from Experimental Models and Postmortem Human Brains. *Mol Neurobiol* **58**, 118-131, doi:10.1007/s12035-020-02109-8 (2021).
- 314 Shim, H. S. *et al.* Telomerase Reverse Transcriptase Preserves Neuron Survival and Cognition in Alzheimer's Disease Models. *Nat Aging* **1**, 1162-1174, doi:10.1038/s43587-021-00146-z (2021).
- 315 Whittemore, K. *et al.* Telomerase gene therapy ameliorates the effects of neurodegeneration associated to short telomeres in mice. *Aging (Albany NY)* **11**, 2916-2948, doi:10.18632/aging.101982 (2019).
- 316 Xiong, X. *et al.* Epigenomic dissection of Alzheimer's disease pinpoints causal variants and reveals epigenome erosion. *Cell* **186**, 4422-4437 e4421, doi:10.1016/j.cell.2023.08.040 (2023).
- 317 Bobkova, N. V. *et al.* Therapeutic effect of exogenous hsp70 in mouse models of Alzheimer's disease. *J Alzheimers Dis* **38**, 425-435, doi:10.3233/JAD-130779 (2014).
- 318 Wagner, J. *et al.* Medin co-aggregates with vascular amyloid-beta in Alzheimer's disease. *Nature* **612**, 123-131, doi:10.1038/s41586-022-05440-3 (2022).
- 319 van Dyck, C. H. *et al.* Lecanemab in Early Alzheimer's Disease. *N Engl J Med* **388**, 9-21, doi:10.1056/NEJMoa2212948 (2023).
- 320 Middeldorp, J. *et al.* Preclinical Assessment of Young Blood Plasma for Alzheimer Disease. *JAMA Neurol* **73**, 1325-1333, doi:10.1001/jamaneurol.2016.3185 (2016).
- 321 Zhao, Y. *et al.* Young blood plasma reduces Alzheimer's disease-like brain pathologies and ameliorates cognitive impairment in 3xTg-AD mice. *Alzheimers Res Ther* **12**, 70, doi:10.1186/s13195-020-00639-w (2020).
- 322 Reisberg, B. *et al.* Memantine in moderate-to-severe Alzheimer's disease. *N Engl J Med* **348**, 1333-1341, doi:10.1056/NEJMoa013128 (2003).
- 323 Sha, S. J. *et al.* Safety, Tolerability, and Feasibility of Young Plasma Infusion in the Plasma for Alzheimer Symptom Amelioration Study: A Randomized Clinical Trial. *JAMA Neurol* **76**, 35-40, doi:10.1001/jamaneurol.2018.3288 (2019).

- 324 Decourt, B., Lahiri, D. K. & Sabbagh, M. N. Targeting Tumor Necrosis Factor Alpha for Alzheimer's Disease. *Curr Alzheimer Res* **14**, 412-425, doi:10.2174/1567205013666160930110551 (2017).
- 325 Tobinick, E. L. & Gross, H. Rapid cognitive improvement in Alzheimer's disease following perispinal etanercept administration. *J Neuroinflammation* **5**, 2, doi:10.1186/1742-2094-5-2 (2008).
- 326 Whittaker, D. S. *et al.* Circadian modulation by time-restricted feeding rescues brain pathology and improves memory in mouse models of Alzheimer's disease. *Cell Metab* **35**, 1704-1721 e1706, doi:10.1016/j.cmet.2023.07.014 (2023).
- 327 Kashiwaya, Y. *et al.* A ketone ester diet exhibits anxiolytic and cognition-sparing properties, and lessens amyloid and tau pathologies in a mouse model of Alzheimer's disease. *Neurobiol Aging* **34**, 1530-1539, doi:10.1016/j.neurobiolaging.2012.11.023 (2013).
- 328 Halagappa, V. K. *et al.* Intermittent fasting and caloric restriction ameliorate age-related behavioral deficits in the triple-transgenic mouse model of Alzheimer's disease. *Neurobiol Dis* **26**, 212-220, doi:10.1016/j.nbd.2006.12.019 (2007).
- 329 Nitzan, K. *et al.* Mitochondrial Transfer Ameliorates Cognitive Deficits, Neuronal Loss, and Gliosis in Alzheimer's Disease Mice. *J Alzheimers Dis* **72**, 587-604, doi:10.3233/JAD-190853 (2019).
- 330 Zhang, P. *et al.* Senolytic therapy alleviates Abeta-associated oligodendrocyte progenitor cell senescence and cognitive deficits in an Alzheimer's disease model. *Nat Neurosci* **22**, 719-728, doi:10.1038/s41593-019-0372-9 (2019).
- 331 Reinitz, F. *et al.* Inhibiting USP16 rescues stem cell aging and memory in an Alzheimer's model. *Elife* **11**, doi:10.7554/elife.66037 (2022).
- 332 Ambroziak, W. *et al.* Genomic instability in the PARK2 locus is associated with Parkinson's disease. *J Appl Genet* **56**, 451-461, doi:10.1007/s13353-015-0282-9 (2015).
- 333 Scheffold, A. *et al.* Telomere shortening leads to an acceleration of synucleinopathy and impaired microglia response in a genetic mouse model. *Acta Neuropathol Commun* **4**, 87, doi:10.1186/s40478-016-0364-x (2016).
- 334 Wan, T., Weir, E. J., Johnson, M., Korolchuk, V. I. & Saretzki, G. C. Increased telomerase improves motor function and alpha-synuclein pathology in a transgenic mouse model of Parkinson's disease associated with enhanced autophagy. *Prog Neurobiol* **199**, 101953, doi:10.1016/j.pneurobio.2020.101953 (2021).
- 335 Horvath, S. & Ritz, B. R. Increased epigenetic age and granulocyte counts in the blood of Parkinson's disease patients. *Aging (Albany NY)* **7**, 1130-1142, doi:10.18632/aging.100859 (2015).
- 336 Taipa, R. *et al.* DJ-1 linked parkinsonism (PARK7) is associated with Lewy body pathology. *Brain* **139**, 1680-1687, doi:10.1093/brain/aww080 (2016).
- 337 Martinez-Vicente, M. *et al.* Dopamine-modified alpha-synuclein blocks chaperone-mediated autophagy. *J Clin Invest* **118**, 777-788, doi:10.1172/JCI32806 (2008).
- 338 Parker, J. E. *et al.* Safety of Plasma Infusions in Parkinson's Disease. *Mov Disord* **35**, 1905-1913, doi:10.1002/mds.28198 (2020).
- 339 Fahn, S. *et al.* Levodopa and the progression of Parkinson's disease. *N Engl J Med* **351**, 2498-2508, doi:10.1056/NEJMoa033447 (2004).
- 340 Garretti, F., Agalliu, D., Lindestam Arlehamn, C. S., Sette, A. & Sulzer, D. Autoimmunity in Parkinson's Disease: The Role of alpha-Synuclein-Specific T Cells. *Front Immunol* **10**, 303, doi:10.3389/fimmu.2019.00303 (2019).
- 341 Du, Y. *et al.* Minocycline prevents nigrostriatal dopaminergic neurodegeneration in the MPTP model of Parkinson's disease. *Proc Natl Acad Sci U S A* **98**, 14669-14674, doi:10.1073/pnas.251341998 (2001).

- 342 Sampson, T. R. *et al.* Gut Microbiota Regulate Motor Deficits and Neuroinflammation in a Model of Parkinson's Disease. *Cell* **167**, 1469-1480 e1412, doi:10.1016/j.cell.2016.11.018 (2016).
- 343 Duan, W. & Mattson, M. P. Dietary restriction and 2-deoxyglucose administration improve behavioral outcome and reduce degeneration of dopaminergic neurons in models of Parkinson's disease. *J Neurosci Res* **57**, 195-206, doi:10.1002/(SICI)1097-4547(19990715)57:2<195::AID-JNR5>3.0.CO;2-P (1999).
- 344 Pirooznia, S. K. *et al.* PARIS induced defects in mitochondrial biogenesis drive dopamine neuron loss under conditions of parkin or PINK1 deficiency. *Mol Neurodegener* **15**, 17, doi:10.1186/s13024-020-00363-x (2020).
- 345 Chinta, S. J. *et al.* Cellular Senescence Is Induced by the Environmental Neurotoxin Paraquat and Contributes to Neuropathology Linked to Parkinson's Disease. *Cell Rep* **22**, 930-940, doi:10.1016/j.celrep.2017.12.092 (2018).
- 346 Bjorklund, L. M. *et al.* Embryonic stem cells develop into functional dopaminergic neurons after transplantation in a Parkinson rat model. *Proc Natl Acad Sci U S A* **99**, 2344-2349, doi:10.1073/pnas.022438099 (2002).
- 347 Liberale, L. *et al.* Endothelial SIRT6 blunts stroke size and neurological deficit by preserving blood-brain barrier integrity: a translational study. *Eur Heart J* **41**, 1575-1587, doi:10.1093/euroheartj/ehz712 (2020).
- 348 Silvera, V. M., Gordon, L. B., Orbach, D. B., Campbell, S. E., Machan, J. T. & Ullrich, N. J. Imaging characteristics of cerebrovascular arteriopathy and stroke in Hutchinson-Gilford progeria syndrome. *AJNR Am J Neuroradiol* **34**, 1091-1097, doi:10.3174/ajnr.A3341 (2013).
- 349 Hennekam, R. C. Hutchinson-Gilford progeria syndrome: review of the phenotype. *Am J Med Genet A* **140**, 2603-2624, doi:10.1002/ajmg.a.31346 (2006).
- 350 Christoffersen, M., Frikke-Schmidt, R., Nordestgaard, B. G. & Tybjaerg-Hansen, A. Genetic variation in WRN and ischemic stroke: General population studies and meta-analyses. *Exp Gerontol* **89**, 69-77, doi:10.1016/j.exger.2017.01.005 (2017).
- 351 Langley, B., Brochier, C. & Rivieccio, M. A. Targeting histone deacetylases as a multifaceted approach to treat the diverse outcomes of stroke. *Stroke* **40**, 2899-2905, doi:10.1161/STROKEAHA.108.540229 (2009).
- 352 Wang, Z. & Yang, W. Impaired capacity to restore proteostasis in the aged brain after ischemia: Implications for translational brain ischemia research. *Neurochem Int* **127**, 87-93, doi:10.1016/j.neuint.2018.12.018 (2019).
- 353 Papadakis, M. *et al.* Tsc1 (hamartin) confers neuroprotection against ischemia by inducing autophagy. *Nat Med* **19**, 351-357, doi:10.1038/nm.3097 (2013).
- 354 Simon, R. & Shiraishi, K. N-methyl-D-aspartate antagonist reduces stroke size and regional glucose metabolism. *Ann Neurol* **27**, 606-611, doi:10.1002/ana.410270604 (1990).
- 355 Yusuf, S. *et al.* Cholesterol Lowering in Intermediate-Risk Persons without Cardiovascular Disease. *N Engl J Med* **374**, 2021-2031, doi:10.1056/NEJMoa1600176 (2016).
- 356 Liberale, L. *et al.* Post-ischaemic administration of the murine Canakinumab-surrogate antibody improves outcome in experimental stroke. *Eur Heart J* **39**, 3511-3517, doi:10.1093/eurheartj/ehy286 (2018).
- 357 Chen, X. *et al.* Combination Therapy with Low-Dose IVIG and a C1-esterase Inhibitor Ameliorates Brain Damage and Functional Deficits in Experimental Ischemic Stroke. *Neuromolecular Med* **20**, 63-72, doi:10.1007/s12017-017-8474-6 (2018).
- 358 Li, H. *et al.* Alterations of gut microbiota contribute to the progression of unruptured intracranial aneurysms. *Nat Commun* **11**, 3218, doi:10.1038/s41467-020-16990-3 (2020).
- 359 Xu, K. *et al.* Rapid gut dysbiosis induced by stroke exacerbates brain infarction in turn. *Gut*, doi:10.1136/gutjnl-2020-323263 (2021).

- 360 Bellastella, G. *et al.* Glucagon-Like Peptide-1 Receptor Agonists and Prevention of Stroke Systematic Review of Cardiovascular Outcome Trials With Meta-Analysis. *Stroke* **51**, 666-669, doi:10.1161/STROKEAHA.119.027557 (2020).
- 361 Liu, F., Lu, J., Manaenko, A., Tang, J. & Hu, Q. Mitochondria in Ischemic Stroke: New Insight and Implications. *Aging Dis* **9**, 924-937, doi:10.14336/AD.2017.1126 (2018).
- 362 Tomas-Loba, A. *et al.* Telomerase reverse transcriptase delays aging in cancer-resistant mice. *Cell* **135**, 609-622, doi:10.1016/j.cell.2008.09.034 (2008).
- 363 Saraswati, S., Martinez, P., Grana-Castro, O. & Blasco, M. A. Short and dysfunctional telomeres sensitize the kidneys to develop fibrosis. *Nat Aging* **1**, 269-283, doi:10.1038/s43587-021-00040-8 (2021).
- 364 Li, L. *et al.* REGgamma deficiency promotes premature aging via the casein kinase 1 pathway. *Proc Natl Acad Sci U S A* **110**, 11005-11010, doi:10.1073/pnas.1308497110 (2013).
- 365 Rowczenio, D. *et al.* Amyloidogenicity and clinical phenotype associated with five novel mutations in apolipoprotein A-I. *Am J Pathol* **179**, 1978-1987, doi:10.1016/j.ajpath.2011.06.024 (2011).
- 366 Yamamoto, T. *et al.* Time-dependent dysregulation of autophagy: Implications in aging and mitochondrial homeostasis in the kidney proximal tubule. *Autophagy* **12**, 801-813, doi:10.1080/15548627.2016.1159376 (2016).
- 367 Davidsohn, N. *et al.* A single combination gene therapy treats multiple age-related diseases. *Proc Natl Acad Sci U S A* **116**, 23505-23511, doi:10.1073/pnas.1910073116 (2019).
- 368 The, E.-K. C. G. *et al.* Empagliflozin in Patients with Chronic Kidney Disease. *N Engl J Med* **388**, 117-127, doi:10.1056/NEJMoa2204233 (2023).
- 369 Barcena, C. *et al.* Healthspan and lifespan extension by fecal microbiota transplantation into progeroid mice. *Nat Med* **25**, 1234-1242, doi:10.1038/s41591-019-0504-5 (2019).
- 370 Mitchell, J. R. *et al.* Short-term dietary restriction and fasting precondition against ischemia reperfusion injury in mice. *Aging Cell* **9**, 40-53, doi:10.1111/j.1474-9726.2009.00532.x (2010).
- 371 Baker, D. J. *et al.* Naturally occurring p16(INK4a)-positive cells shorten healthy lifespan. *Nature* **530**, 184-189, doi:10.1038/nature16932 (2016).
- 372 Hickson, L. J. *et al.* Senolytics decrease senescent cells in humans: Preliminary report from a clinical trial of Dasatinib plus Quercetin in individuals with diabetic kidney disease. *EBioMedicine* **47**, 446-456, doi:10.1016/j.ebiom.2019.08.069 (2019).
- 373 Alder, J. K. & Armanios, M. Telomere-mediated lung disease. *Physiol Rev* **102**, 1703-1720, doi:10.1152/physrev.00046.2021 (2022).
- 374 Armanios, M. Telomeres and age-related disease: how telomere biology informs clinical paradigms. *J Clin Invest* **123**, 996-1002, doi:10.1172/JCI66370 (2013).
- 375 Nunes, H. *et al.* Is telomeropathy the explanation for combined pulmonary fibrosis and emphysema syndrome?: report of a family with TERT mutation. *Am J Respir Crit Care Med* **189**, 753-754, doi:10.1164/rccm.201309-1724LE (2014).
- 376 Ito, K. *et al.* Decreased histone deacetylase activity in chronic obstructive pulmonary disease. *N Engl J Med* **352**, 1967-1976, doi:10.1056/NEJMoa041892 (2005).
- 377 Miravitles, M. *et al.* European Respiratory Society statement: diagnosis and treatment of pulmonary disease in alpha(1)-antitrypsin deficiency. *Eur Respir J* **50**, doi:10.1183/13993003.00610-2017 (2017).
- 378 Mizumura, K. *et al.* Mitophagy-dependent necroptosis contributes to the pathogenesis of COPD. *J Clin Invest* **124**, 3987-4003, doi:10.1172/JCI74985 (2014).
- 379 Yao, H. *et al.* SIRT1 protects against emphysema via FOXO3-mediated reduction of premature senescence in mice. *J Clin Invest* **122**, 2032-2045, doi:10.1172/JCI60132 (2012).
- 380 Chapman, K. R. *et al.* Long-acting antimuscarinic therapy in patients with chronic obstructive pulmonary disease receiving beta-blockers. *Respir Res* **22**, 272, doi:10.1186/s12931-021-01861-2 (2021).

- 381 Yao, H. *et al.* SIRT1 redresses the imbalance of tissue inhibitor of matrix metalloproteinase-1 and matrix metalloproteinase-9 in the development of mouse emphysema and human COPD. *Am J Physiol Lung Cell Mol Physiol* **305**, L615-624, doi:10.1152/ajplung.00249.2012 (2013).
- 382 Lai, H. C. *et al.* Gut microbiota modulates COPD pathogenesis: role of anti-inflammatory Parabacteroides goldsteinii lipopolysaccharide. *Gut* **71**, 309-321, doi:10.1136/gutjnl-2020-322599 (2022).
- 383 Suga, T. *et al.* Disruption of the klotho gene causes pulmonary emphysema in mice. Defect in maintenance of pulmonary integrity during postnatal life. *Am J Respir Cell Mol Biol* **22**, 26-33, doi:10.1165/ajrcmb.22.1.3554 (2000).
- 384 Jang, Y. O. *et al.* High-fiber diets attenuate emphysema development via modulation of gut microbiota and metabolism. *Sci Rep* **11**, 7008, doi:10.1038/s41598-021-86404-x (2021).
- 385 Johnson, J. B. *et al.* Alternate day calorie restriction improves clinical findings and reduces markers of oxidative stress and inflammation in overweight adults with moderate asthma. *Free Radic Biol Med* **42**, 665-674, doi:10.1016/j.freeradbiomed.2006.12.005 (2007).
- 386 Cloonan, S. M. & Choi, A. M. Mitochondria in lung disease. *J Clin Invest* **126**, 809-820, doi:10.1172/JCI81113 (2016).
- 387 Kurozumi, M., Matsushita, T., Hosokawa, M. & Takeda, T. Age-related changes in lung structure and function in the senescence-accelerated mouse (SAM): SAM-P/1 as a new murine model of senile hyperinflation of lung. *Am J Respir Crit Care Med* **149**, 776-782, doi:10.1164/ajrccm.149.3.8118649 (1994).
- 388 Ryan, D. M. *et al.* Smoking dysregulates the human airway basal cell transcriptome at COPD risk locus 19q13.2. *PLoS One* **9**, e88051, doi:10.1371/journal.pone.0088051 (2014).
- 389 Paschalaki, K. E. *et al.* Dysfunction of endothelial progenitor cells from smokers and chronic obstructive pulmonary disease patients due to increased DNA damage and senescence. *Stem Cells* **31**, 2813-2826, doi:10.1002/stem.1488 (2013).
- 390 Povedano, J. M. *et al.* Therapeutic effects of telomerase in mice with pulmonary fibrosis induced by damage to the lungs and short telomeres. *eLife* **7**, doi:10.7554/eLife.31299 (2018).
- 391 Le Saux, C. J. *et al.* A novel telomerase activator suppresses lung damage in a murine model of idiopathic pulmonary fibrosis. *PLoS One* **8**, e58423, doi:10.1371/journal.pone.0058423 (2013).
- 392 Calado, R. T. & Young, N. S. Telomere diseases. *N Engl J Med* **361**, 2353-2365, doi:10.1056/NEJMra0903373 (2009).
- 393 Coward, W. R., Watts, K., Feghali-Bostwick, C. A., Knox, A. & Pang, L. Defective histone acetylation is responsible for the diminished expression of cyclooxygenase 2 in idiopathic pulmonary fibrosis. *Mol Cell Biol* **29**, 4325-4339, doi:10.1128/MCB.01776-08 (2009).
- 394 Seibold, M. A. *et al.* A common MUC5B promoter polymorphism and pulmonary fibrosis. *N Engl J Med* **364**, 1503-1512, doi:10.1056/NEJMoa1013660 (2011).
- 395 Lawson, W. E. *et al.* Genetic mutations in surfactant protein C are a rare cause of sporadic cases of IPF. *Thorax* **59**, 977-980, doi:10.1136/thx.2004.026336 (2004).
- 396 Hill, C. *et al.* Autophagy inhibition-mediated epithelial-mesenchymal transition augments local myofibroblast differentiation in pulmonary fibrosis. *Cell Death Dis* **10**, 591, doi:10.1038/s41419-019-1820-x (2019).
- 397 Richeldi, L. *et al.* Efficacy and safety of nintedanib in idiopathic pulmonary fibrosis. *N Engl J Med* **370**, 2071-2082, doi:10.1056/NEJMoa1402584 (2014).
- 398 Sosulski, M. L., Gongora, R., Feghali-Bostwick, C., Lasky, J. A. & Sanchez, C. G. Sirtuin 3 Dereulation Promotes Pulmonary Fibrosis. *J Gerontol A Biol Sci Med Sci* **72**, 595-602, doi:10.1093/gerona/glw151 (2017).
- 399 King, T. E., Jr. *et al.* A phase 3 trial of pirfenidone in patients with idiopathic pulmonary fibrosis. *N Engl J Med* **370**, 2083-2092, doi:10.1056/NEJMoa1402582 (2014).
- 400 Yang, D., Xing, Y., Song, X. & Qian, Y. The impact of lung microbiota dysbiosis on inflammation. *Immunology* **159**, 156-166, doi:10.1111/imm.13139 (2020).

- 401 Han, Y. *et al.* Mefenidone Ameliorates Bleomycin-Induced Pulmonary Fibrosis in Mice. *Front Pharmacol* **12**, 713572, doi:10.3389/fphar.2021.713572 (2021).
- 402 Bueno, M. *et al.* PINK1 deficiency impairs mitochondrial homeostasis and promotes lung fibrosis. *J Clin Invest* **125**, 521-538, doi:10.1172/JCI74942 (2015).
- 403 Calhoun, C. *et al.* Senescent Cells Contribute to the Physiological Remodeling of Aged Lungs. *J Gerontol A Biol Sci Med Sci* **71**, 153-160, doi:10.1093/gerona/glu241 (2016).
- 404 Schafer, M. J. *et al.* Cellular senescence mediates fibrotic pulmonary disease. *Nat Commun* **8**, 14532, doi:10.1038/ncomms14532 (2017).
- 405 Bustos, M. L. *et al.* Aging mesenchymal stem cells fail to protect because of impaired migration and antiinflammatory response. *Am J Respir Crit Care Med* **189**, 787-798, doi:10.1164/rccm.201306-1043OC (2014).
- 406 Benkafadar, N. *et al.* ROS-Induced Activation of DNA Damage Responses Drives Senescence-Like State in Postmitotic Cochlear Cells: Implication for Hearing Preservation. *Mol Neurobiol* **56**, 5950-5969, doi:10.1007/s12035-019-1493-6 (2019).
- 407 Menardo, J. *et al.* Oxidative stress, inflammation, and autophagic stress as the key mechanisms of premature age-related hearing loss in SAMP8 mouse Cochlea. *Antioxid Redox Signal* **16**, 263-274, doi:10.1089/ars.2011.4037 (2012).
- 408 Shastry, S. *et al.* A novel syndrome of mandibular hypoplasia, deafness, and progeroid features associated with lipodystrophy, undescended testes, and male hypogonadism. *J Clin Endocrinol Metab* **95**, E192-197, doi:10.1210/jc.2010-0419 (2010).
- 409 Watanabe, K. & Bloch, W. Histone methylation and acetylation indicates epigenetic change in the aged cochlea of mice. *Eur Arch Otorhinolaryngol* **270**, 1823-1830, doi:10.1007/s00405-012-2222-1 (2013).
- 410 Wu, X. *et al.* Reduced expression of Connexin26 and its DNA promoter hypermethylation in the inner ear of mimetic aging rats induced by d-galactose. *Biochem Biophys Res Commun* **452**, 340-346, doi:10.1016/j.bbrc.2014.08.063 (2014).
- 411 Peixoto Pinheiro, B., Adel, Y., Knipper, M., Muller, M. & Lowenheim, H. Auditory Threshold Variability in the SAMP8 Mouse Model of Age-Related Hearing Loss: Functional Loss and Phenotypic Change Precede Outer Hair Cell Loss. *Front Aging Neurosci* **13**, 708190, doi:10.3389/fnagi.2021.708190 (2021).
- 412 Choo, O. S. *et al.* Effect of statin on age-related hearing loss via drug repurposing. *Biochim Biophys Acta Mol Cell Res* **1869**, 119331, doi:10.1016/j.bbamcr.2022.119331 (2022).
- 413 Someya, S. *et al.* Sirt3 mediates reduction of oxidative damage and prevention of age-related hearing loss under caloric restriction. *Cell* **143**, 802-812, doi:10.1016/j.cell.2010.10.002 (2010).
- 414 Xiong, H. *et al.* Activation of miR-34a/SIRT1/p53 signaling contributes to cochlear hair cell apoptosis: implications for age-related hearing loss. *Neurobiol Aging* **36**, 1692-1701, doi:10.1016/j.neurobiolaging.2014.12.034 (2015).
- 415 Cho, S. I., Jo, E. R. & Song, H. Urolithin A attenuates auditory cell senescence by activating mitophagy. *Sci Rep* **12**, 7704, doi:10.1038/s41598-022-11894-2 (2022).
- 416 Someya, S. *et al.* Age-related hearing loss in C57BL/6J mice is mediated by Bak-dependent mitochondrial apoptosis. *Proc Natl Acad Sci U S A* **106**, 19432-19437, doi:10.1073/pnas.0908786106 (2009).
- 417 Sarzi, E. *et al.* The human OPA1delTTAG mutation induces premature age-related systemic neurodegeneration in mouse. *Brain* **135**, 3599-3613, doi:10.1093/brain/aws303 (2012).
- 418 Seidman, M. D., Khan, M. J., Bai, U., Shirwany, N. & Quirk, W. S. Biologic activity of mitochondrial metabolites on aging and age-related hearing loss. *Am J Otol* **21**, 161-167, doi:10.1016/s0196-0709(00)80003-4 (2000).
- 419 Trifunovic, A. *et al.* Somatic mtDNA mutations cause aging phenotypes without affecting reactive oxygen species production. *Proc Natl Acad Sci U S A* **102**, 17993-17998, doi:10.1073/pnas.0508886102 (2005).

- 420 Sharma, P., Sharma, N. & Sharma, D. A Narrative Review on Fanconi Anemia: Genetic and Diagnostic Considerations. *Glob Med Genet* **9**, 237-241, doi:10.1055/s-0042-1751303 (2022).
- 421 Dow, C. T. & Harley, C. B. Evaluation of an oral telomerase activator for early age-related macular degeneration - a pilot study. *Clin Ophthalmol* **10**, 243-249, doi:10.2147/OPTH.S100042 (2016).
- 422 Lu, Y. *et al.* Reprogramming to recover youthful epigenetic information and restore vision. *Nature* **588**, 124-129, doi:10.1038/s41586-020-2975-4 (2020).
- 423 Kozhevnikova, O. S., Telegina, D. V., Devyatkin, V. A. & Kolosova, N. G. Involvement of the autophagic pathway in the progression of AMD-like retinopathy in senescence-accelerated OXYS rats. *Biogerontology* **19**, 223-235, doi:10.1007/s10522-018-9751-y (2018).
- 424 McLaughlin, T. *et al.* Loss of XBP1 accelerates age-related decline in retinal function and neurodegeneration. *Mol Neurodegener* **13**, 16, doi:10.1186/s13024-018-0250-z (2018).
- 425 Sun, Q. *et al.* Inhibition of Sumoylation Alleviates Oxidative Stress-induced Retinal Pigment Epithelial Cell Senescence and Represses Proinflammatory Gene Expression. *Curr Mol Med* **18**, 575-583, doi:10.2174/1566524019666190107154250 (2018).
- 426 Jadeja, R. N. *et al.* Loss of NAMPT in aging retinal pigment epithelium reduces NAD(+) availability and promotes cellular senescence. *Aging (Albany NY)* **10**, 1306-1323, doi:10.18632/aging.101469 (2018).
- 427 Haines, J. L. *et al.* Complement factor H variant increases the risk of age-related macular degeneration. *Science* **308**, 419-421, doi:10.1126/science.1110359 (2005).
- 428 Tarallo, V. *et al.* DICER1 loss and Alu RNA induce age-related macular degeneration via the NLRP3 inflammasome and MyD88. *Cell* **149**, 847-859, doi:10.1016/j.cell.2012.03.036 (2012).
- 429 Hu, S. J. *et al.* Upregulation of P2RX7 in Cx3cr1-Deficient Mononuclear Phagocytes Leads to Increased Interleukin-1beta Secretion and Photoreceptor Neurodegeneration. *J Neurosci* **35**, 6987-6996, doi:10.1523/JNEUROSCI.3955-14.2015 (2015).
- 430 Edwards, A. O., Ritter, R., 3rd, Abel, K. J., Manning, A., Panhuysen, C. & Farrer, L. A. Complement factor H polymorphism and age-related macular degeneration. *Science* **308**, 421-424, doi:10.1126/science.1110189 (2005).
- 431 Rowan, S. *et al.* Involvement of a gut-retina axis in protection against dietary glycemia-induced age-related macular degeneration. *Proc Natl Acad Sci U S A* **114**, E4472-E4481, doi:10.1073/pnas.1702302114 (2017).
- 432 Zhang, M. *et al.* Dysregulated metabolic pathways in age-related macular degeneration. *Sci Rep* **10**, 2464, doi:10.1038/s41598-020-59244-4 (2020).
- 433 Zhao, C. *et al.* mTOR-mediated dedifferentiation of the retinal pigment epithelium initiates photoreceptor degeneration in mice. *J Clin Invest* **121**, 369-383, doi:10.1172/JCI44303 (2011).
- 434 Mao, H. *et al.* Mitochondrial oxidative stress in the retinal pigment epithelium leads to localized retinal degeneration. *Invest Ophthalmol Vis Sci* **55**, 4613-4627, doi:10.1167/iovs.14-14633 (2014).
- 435 Baker, D. J. *et al.* BubR1 insufficiency causes early onset of aging-associated phenotypes and infertility in mice. *Nat Genet* **36**, 744-749, doi:10.1038/ng1382 (2004).
- 436 Zhao, L. *et al.* Lanosterol reverses protein aggregation in cataracts. *Nature* **523**, 607-611, doi:10.1038/nature14650 (2015).
- 437 Makley, L. N. *et al.* Pharmacological chaperone for alpha-crystallin partially restores transparency in cataract models. *Science* **350**, 674-677, doi:10.1126/science.aac9145 (2015).
- 438 Sekijima, Y. in *GeneReviews((R))* (eds M. P. Adam *et al.*) (1993).
- 439 Sidjanin, D. J., Park, A. K., Ronchetti, A., Martins, J. & Jackson, W. T. TBC1D20 mediates autophagy as a key regulator of autophagosome maturation. *Autophagy* **12**, 1759-1775, doi:10.1080/15548627.2016.1199300 (2016).
- 440 Liegel, R. P. *et al.* Loss-of-function mutations in TBC1D20 cause cataracts and male infertility in blind sterile mice and Warburg micro syndrome in humans. *Am J Hum Genet* **93**, 1001-1014, doi:10.1016/j.ajhg.2013.10.011 (2013).

- 441 Hales, A. M., Chamberlain, C. G., Murphy, C. R. & McAvoy, J. W. Estrogen protects lenses against cataract induced by transforming growth factor-beta (TGFbeta). *J Exp Med* **185**, 273-280, doi:10.1084/jem.185.2.273 (1997).
- 442 Blakytny, R. & Harding, J. J. Prevention of cataract in diabetic rats by aspirin, paracetamol (acetaminophen) and ibuprofen. *Exp Eye Res* **54**, 509-518, doi:10.1016/0014-4835(92)90129-g (1992).
- 443 Wolf, N. et al. Age-related cataract progression in five mouse models for anti-oxidant protection or hormonal influence. *Exp Eye Res* **81**, 276-285, doi:10.1016/j.exer.2005.01.024 (2005).
- 444 Baker, D. J. et al. Clearance of p16Ink4a-positive senescent cells delays ageing-associated disorders. *Nature* **479**, 232-236, doi:10.1038/nature10600 (2011).
- 445 Massip, L. et al. Vitamin C restores healthy aging in a mouse model for Werner syndrome. *FASEB J* **24**, 158-172, doi:10.1096/fj.09-137133 (2010).
- 446 Rudolph, K. L., Chang, S., Millard, M., Schreiber-Agus, N. & DePinho, R. A. Inhibition of experimental liver cirrhosis in mice by telomerase gene delivery. *Science* **287**, 1253-1258, doi:10.1126/science.287.5456.1253 (2000).
- 447 Castoldi, F. et al. Autophagy-mediated metabolic effects of aspirin. *Cell Death Discov* **6**, 129, doi:10.1038/s41420-020-00365-0 (2020).
- 448 Mehdipour, M. et al. Rejuvenation of three germ layers tissues by exchanging old blood plasma with saline-albumin. *Aging (Albany NY)* **12**, 8790-8819, doi:10.18632/aging.103418 (2020).
- 449 Grunewald, M. et al. Counteracting age-related VEGF signaling insufficiency promotes healthy aging and extends life span. *Science* **373**, doi:10.1126/science.abc8479 (2021).
- 450 Yan, J. et al. Liraglutide, Sitagliptin, and Insulin Glargine Added to Metformin: The Effect on Body Weight and Intrahepatic Lipid in Patients With Type 2 Diabetes Mellitus and Nonalcoholic Fatty Liver Disease. *Hepatology* **69**, 2414-2426, doi:10.1002/hep.30320 (2019).
- 451 Krishnan, S. et al. Gut Microbiota-Derived Tryptophan Metabolites Modulate Inflammatory Response in Hepatocytes and Macrophages. *Cell Rep* **23**, 1099-1111, doi:10.1016/j.celrep.2018.03.109 (2018).
- 452 Depommier, C. et al. Supplementation with Akkermansia muciniphila in overweight and obese human volunteers: a proof-of-concept exploratory study. *Nat Med* **25**, 1096-1103, doi:10.1038/s41591-019-0495-2 (2019).
- 453 Perry, R. J. et al. Reversal of hypertriglyceridemia, fatty liver disease, and insulin resistance by a liver-targeted mitochondrial uncoupler. *Cell Metab* **18**, 740-748, doi:10.1016/j.cmet.2013.10.004 (2013).
- 454 Samuel, V. T. et al. Inhibition of protein kinase Cepsilon prevents hepatic insulin resistance in nonalcoholic fatty liver disease. *J Clin Invest* **117**, 739-745, doi:10.1172/JCI30400 (2007).
- 455 Goedeke, L. et al. Sex- and strain-specific effects of mitochondrial uncoupling on age-related metabolic diseases in high-fat diet-fed mice. *Aging Cell* **21**, e13539, doi:10.1111/acel.13539 (2022).
- 456 Goedeke, L. et al. Controlled-release mitochondrial protonophore (CRMP) reverses dyslipidemia and hepatic steatosis in dysmetabolic nonhuman primates. *Sci Transl Med* **11**, doi:10.1126/scitranslmed.aay0284 (2019).
- 457 Ogrodnik, M. et al. Cellular senescence drives age-dependent hepatic steatosis. *Nat Commun* **8**, 15691, doi:10.1038/ncomms15691 (2017).
- 458 Wang, W. et al. A genome-wide CRISPR-based screen identifies KAT7 as a driver of cellular senescence. *Sci Transl Med* **13**, doi:10.1126/scitranslmed.abd2655 (2021).
- 459 Munoz-Lorente, M. A., Cano-Martin, A. C. & Blasco, M. A. Mice with hyper-long telomeres show less metabolic aging and longer lifespans. *Nat Commun* **10**, 4723, doi:10.1038/s41467-019-12664-x (2019).

- 460 Schratz, K. E. Extrahematopoietic manifestations of the short telomere syndromes. *Hematology Am Soc Hematol Educ Program* **2020**, 115-122, doi:10.1182/hematology.2020000170 (2020).
- 461 Zhang, B. *et al.* Multi-omic rejuvenation and life span extension on exposure to youthful circulation. *Nat Aging* **3**, 948-964, doi:10.1038/s43587-023-00451-9 (2023).
- 462 Moncsek, A. *et al.* Targeting senescent cholangiocytes and activated fibroblasts with B-cell lymphoma-extra large inhibitors ameliorates fibrosis in multidrug resistance 2 gene knockout (Mdr2(-/-)) mice. *Hepatology* **67**, 247-259, doi:10.1002/hep.29464 (2018).
- 463 Rudolph, K. L. *et al.* Longevity, stress response, and cancer in aging telomerase-deficient mice. *Cell* **96**, 701-712, doi:10.1016/s0092-8674(00)80580-2 (1999).
- 464 Beier, F., Foronda, M., Martinez, P. & Blasco, M. A. Conditional TRF1 knockout in the hematopoietic compartment leads to bone marrow failure and recapitulates clinical features of dyskeratosis congenita. *Blood* **120**, 2990-3000, doi:10.1182/blood-2012-03-418038 (2012).
- 465 Colla, S. *et al.* Telomere dysfunction drives aberrant hematopoietic differentiation and myelodysplastic syndrome. *Cancer Cell* **27**, 644-657, doi:10.1016/j.ccr.2015.04.007 (2015).
- 466 Vulliamy, T., Marrone, A., Dokal, I. & Mason, P. J. Association between aplastic anaemia and mutations in telomerase RNA. *Lancet* **359**, 2168-2170, doi:10.1016/S0140-6736(02)09087-6 (2002).
- 467 Kantarjian, H. *et al.* Decitabine improves patient outcomes in myelodysplastic syndromes: results of a phase III randomized study. *Cancer* **106**, 1794-1803, doi:10.1002/cncr.21792 (2006).
- 468 Fenaux, P. *et al.* Efficacy of azacitidine compared with that of conventional care regimens in the treatment of higher-risk myelodysplastic syndromes: a randomised, open-label, phase III study. *Lancet Oncol* **10**, 223-232, doi:10.1016/S1470-2045(09)70003-8 (2009).
- 469 Clarke, M. L. *et al.* MYB insufficiency disrupts proteostasis in hematopoietic stem cells, leading to age-related neoplasia. *Blood* **141**, 1858-1870, doi:10.1182/blood.2022019138 (2023).
- 470 Allen, J. E. *et al.* Identification of TRAIL-inducing compounds highlights small molecule ONC201/TIC10 as a unique anti-cancer agent that activates the TRAIL pathway. *Mol Cancer* **14**, 99, doi:10.1186/s12943-015-0346-9 (2015).
- 471 Townsley, D. M. *et al.* Danazol Treatment for Telomere Diseases. *N Engl J Med* **374**, 1922-1931, doi:10.1056/NEJMoa1515319 (2016).
- 472 Bertoli, S. *et al.* Dexamethasone in hyperleukocytic acute myeloid leukemia. *Haematologica* **103**, 988-998, doi:10.3324/haematol.2017.184267 (2018).
- 473 Calcinotto, A. *et al.* Microbiota-driven interleukin-17-producing cells and eosinophils synergize to accelerate multiple myeloma progression. *Nat Commun* **9**, 4832, doi:10.1038/s41467-018-07305-8 (2018).
- 474 Descamps, O., Riondel, J., Ducros, V. & Roussel, A. M. Mitochondrial production of reactive oxygen species and incidence of age-associated lymphoma in OF1 mice: effect of alternate-day fasting. *Mech Ageing Dev* **126**, 1185-1191, doi:10.1016/j.mad.2005.06.007 (2005).
- 475 Hattori, A. *et al.* Cancer progression by reprogrammed BCAA metabolism in myeloid leukaemia. *Nature* **545**, 500-504, doi:10.1038/nature22314 (2017).
- 476 Raffel, S. *et al.* BCAT1 restricts alphaKG levels in AML stem cells leading to IDHmut-like DNA hypermethylation. *Nature* **551**, 384-388, doi:10.1038/nature24294 (2017).
- 477 Stein, E. M. *et al.* Enasidenib in mutant IDH2 relapsed or refractory acute myeloid leukemia. *Blood* **130**, 722-731, doi:10.1182/blood-2017-04-779405 (2017).
- 478 Bar, C. *et al.* Telomerase gene therapy rescues telomere length, bone marrow aplasia, and survival in mice with aplastic anemia. *Blood* **127**, 1770-1779, doi:10.1182/blood-2015-08-667485 (2016).
- 479 Yamaguchi, H. *et al.* Mutations of the human telomerase RNA gene (TERC) in aplastic anemia and myelodysplastic syndrome. *Blood* **102**, 916-918, doi:10.1182/blood-2003-01-0335 (2003).

- 480 Roichman, A. *et al.* Restoration of energy homeostasis by SIRT6 extends healthy lifespan. *Nat Commun* **12**, 3208, doi:10.1038/s41467-021-23545-7 (2021).
- 481 Dong, S. *et al.* Chaperone-mediated autophagy sustains haematopoietic stem-cell function. *Nature* **591**, 117-123, doi:10.1038/s41586-020-03129-z (2021).
- 482 Moist, L. M. *et al.* Clinical practice guidelines for evidence-based use of erythropoietic-stimulating agents. *Kidney Int Suppl*, S12-18, doi:10.1038/ki.2008.270 (2008).
- 483 Shanmugam, N. K., Chen, K. & Cherayil, B. J. Commensal Bacteria-induced Interleukin 1beta (IL-1beta) Secreted by Macrophages Up-regulates Hepcidin Expression in Hepatocytes by Activating the Bone Morphogenetic Protein Signaling Pathway. *J Biol Chem* **290**, 30637-30647, doi:10.1074/jbc.M115.689190 (2015).
- 484 Trifunovic, A. *et al.* Premature ageing in mice expressing defective mitochondrial DNA polymerase. *Nature* **429**, 417-423, doi:10.1038/nature02517 (2004).
- 485 Harrison, D. E. Long-term erythropoietic repopulating ability of old, young, and fetal stem cells. *J Exp Med* **157**, 1496-1504, doi:10.1084/jem.157.5.1496 (1983).
- 486 Galmozzi, A. *et al.* Inhibition of class I histone deacetylases unveils a mitochondrial signature and enhances oxidative metabolism in skeletal muscle and adipose tissue. *Diabetes* **62**, 732-742, doi:10.2337/db12-0548 (2013).
- 487 Frohlich, J. & Vinciguerra, M. Candidate rejuvenating factor GDF11 and tissue fibrosis: friend or foe? *Geroscience* **42**, 1475-1498, doi:10.1007/s11357-020-00279-w (2020).
- 488 Zhang, Y. J. *et al.* Efficacy and safety of empagliflozin for type 2 diabetes mellitus: Meta-analysis of randomized controlled trials. *Medicine (Baltimore)* **97**, e12843, doi:10.1097/MD.00000000000012843 (2018).
- 489 Pedersen, C. R., Hagemann, I., Bock, T. & Buschard, K. Intermittent feeding and fasting reduces diabetes incidence in BB rats. *Autoimmunity* **30**, 243-250, doi:10.3109/08916939908993805 (1999).
- 490 Halberg, N. *et al.* Effect of intermittent fasting and refeeding on insulin action in healthy men. *J Appl Physiol* (1985) **99**, 2128-2136, doi:10.1152/japplphysiol.00683.2005 (2005).
- 491 Tavallaie, M. *et al.* Moderation of mitochondrial respiration mitigates metabolic syndrome of aging. *Proc Natl Acad Sci U S A* **117**, 9840-9850, doi:10.1073/pnas.1917948117 (2020).
- 492 Caldeira da Silva, C. C., Cerqueira, F. M., Barbosa, L. F., Medeiros, M. H. & Kowaltowski, A. J. Mild mitochondrial uncoupling in mice affects energy metabolism, redox balance and longevity. *Aging Cell* **7**, 552-560, doi:10.1111/j.1474-9726.2008.00407.x (2008).
- 493 Tomaru, U. *et al.* Decreased proteasomal activity causes age-related phenotypes and promotes the development of metabolic abnormalities. *Am J Pathol* **180**, 963-972, doi:10.1016/j.ajpath.2011.11.012 (2012).
- 494 Min, J. N., Whaley, R. A., Sharpless, N. E., Lockyer, P., Portbury, A. L. & Patterson, C. CHIP deficiency decreases longevity, with accelerated aging phenotypes accompanied by altered protein quality control. *Mol Cell Biol* **28**, 4018-4025, doi:10.1128/MCB.00296-08 (2008).
- 495 Zhu, L. *et al.* Variants in ALDH1A2 reveal an anti-inflammatory role for retinoic acid and a new class of disease-modifying drugs in osteoarthritis. *Sci Transl Med* **14**, eabm4054, doi:10.1126/scitranslmed.abm4054 (2022).
- 496 Novais, E. J. *et al.* Long-term treatment with senolytic drugs Dasatinib and Quercetin ameliorates age-dependent intervertebral disc degeneration in mice. *Nat Commun* **12**, 5213, doi:10.1038/s41467-021-25453-2 (2021).
- 497 Jeon, O. H. *et al.* Local clearance of senescent cells attenuates the development of post-traumatic osteoarthritis and creates a pro-regenerative environment. *Nat Med* **23**, 775-781, doi:10.1038/nm.4324 (2017).
- 498 Su, X. *et al.* TA63 prevents premature aging by promoting adult stem cell maintenance. *Cell Stem Cell* **5**, 64-75, doi:10.1016/j.stem.2009.04.003 (2009).
- 499 Mostoslavsky, R. *et al.* Genomic instability and aging-like phenotype in the absence of mammalian SIRT6. *Cell* **124**, 315-329, doi:10.1016/j.cell.2005.11.044 (2006).

- 500 Baht, G. S. *et al.* Exposure to a youthful circulation rejuvenates bone repair through modulation of beta-catenin. *Nat Commun* **6**, 7131, doi:10.1038/ncomms8131 (2015).
- 501 McCloskey, E. V. *et al.* Clodronate reduces the incidence of fractures in community-dwelling elderly women unselected for osteoporosis: results of a double-blind, placebo-controlled randomized study. *J Bone Miner Res* **22**, 135-141, doi:10.1359/jbmr.061008 (2007).
- 502 Sciorati, C. *et al.* Pharmacological blockade of TNFalpha prevents sarcopenia and prolongs survival in aging mice. *Aging (Albany NY)* **12**, 23497-23508, doi:10.18632/aging.202200 (2020).
- 503 Chrzanowska, K. H., Gregorek, H., Dembowska-Baginska, B., Kalina, M. A. & Digweed, M. Nijmegen breakage syndrome (NBS). *Orphanet J Rare Dis* **7**, 13, doi:10.1186/1750-1172-7-13 (2012).
- 504 Liu, L., Blasco, M., Trimarchi, J. & Keefe, D. An essential role for functional telomeres in mouse germ cells during fertilization and early development. *Dev Biol* **249**, 74-84, doi:10.1006/dbio.2002.0735 (2002).
- 505 Armanios, M. & Blackburn, E. H. The telomere syndromes. *Nat Rev Genet* **13**, 693-704, doi:10.1038/nrg3246 (2012).
- 506 Wang, Y. Y. *et al.* Decreased Klotho Expression Causes Accelerated Decline of Male Fecundity through Oxidative Injury in Murine Testis. *Antioxidants (Basel)* **12**, doi:10.3390/antiox12091671 (2023).
- 507 Blake, K. D. & Prasad, C. CHARGE syndrome. *Orphanet J Rare Dis* **1**, 34, doi:10.1186/1750-1172-1-34 (2006).
- 508 Xu, L. *et al.* Fecal microbiota transplantation from young donor mice improves ovarian function in aged mice. *J Genet Genomics* **49**, 1042-1052, doi:10.1016/j.jgg.2022.05.006 (2022).
- 509 Kim, H. S. *et al.* SIRT2 maintains genome integrity and suppresses tumorigenesis through regulating APC/C activity. *Cancer Cell* **20**, 487-499, doi:10.1016/j.ccr.2011.09.004 (2011).
- 510 Hofstatter, E. W. *et al.* Increased epigenetic age in normal breast tissue from luminal breast cancer patients. *Clin Epigenetics* **10**, 112, doi:10.1186/s13148-018-0534-8 (2018).
- 511 Feng, Y. X. *et al.* Epithelial-to-mesenchymal transition activates PERK-eIF2alpha and sensitizes cells to endoplasmic reticulum stress. *Cancer Discov* **4**, 702-715, doi:10.1158/2159-8290.CD-13-0945 (2014).
- 512 Stein, M. N. *et al.* First-in-Human Clinical Trial of Oral ONC201 in Patients with Refractory Solid Tumors. *Clin Cancer Res* **23**, 4163-4169, doi:10.1158/1078-0432.CCR-16-2658 (2017).
- 513 Reck, M. *et al.* Docetaxel plus nintedanib versus docetaxel plus placebo in patients with previously treated non-small-cell lung cancer (LUME-Lung 1): a phase 3, double-blind, randomised controlled trial. *Lancet Oncol* **15**, 143-155, doi:10.1016/S1470-2045(13)70586-2 (2014).
- 514 Choi, I. J. *et al.* Family History of Gastric Cancer and Helicobacter pylori Treatment. *N Engl J Med* **382**, 427-436, doi:10.1056/NEJMoa1909666 (2020).
- 515 Ulgherait, M. *et al.* Circadian regulation of mitochondrial uncoupling and lifespan. *Nat Commun* **11**, 1927, doi:10.1038/s41467-020-15617-x (2020).
- 516 Larizza, L., Roversi, G. & Volpi, L. Rothmund-Thomson syndrome. *Orphanet J Rare Dis* **5**, 2, doi:10.1186/1750-1172-5-2 (2010).
- 517 Flores, I., Cayuela, M. L. & Blasco, M. A. Effects of telomerase and telomere length on epidermal stem cell behavior. *Science* **309**, 1253-1256, doi:10.1126/science.1115025 (2005).
- 518 Gronskov, K., Ek, J. & Brondum-Nielsen, K. Oculocutaneous albinism. *Orphanet J Rare Dis* **2**, 43, doi:10.1186/1750-1172-2-43 (2007).
- 519 Browder, K. C. *et al.* In vivo partial reprogramming alters age-associated molecular changes during physiological aging in mice. *Nat Aging* **2**, 243-253, doi:10.1038/s43587-022-00183-2 (2022).

- 520 Fitzgerald, K. N. *et al.* Potential reversal of epigenetic age using a diet and lifestyle intervention: a pilot randomized clinical trial. *Aging (Albany NY)* **13**, 9419-9432, doi:10.18632/aging.202913 (2021).
- 521 Aversa, Z. *et al.* Calorie restriction reduces biomarkers of cellular senescence in humans. *Aging Cell* **23**, e14038, doi:10.1111/acel.14038 (2024).
- 522 Kraus, W. E. *et al.* 2 years of calorie restriction and cardiometabolic risk (CALERIE): exploratory outcomes of a multicentre, phase 2, randomised controlled trial. *Lancet Diabetes Endocrinol* **7**, 673-683, doi:10.1016/S2213-8587(19)30151-2 (2019).
- 523 Lefevre, M. *et al.* Caloric restriction alone and with exercise improves CVD risk in healthy non-obese individuals. *Atherosclerosis* **203**, 206-213, doi:10.1016/j.atherosclerosis.2008.05.036 (2009).
- 524 Christensen, P. *et al.* Men and women respond differently to rapid weight loss: Metabolic outcomes of a multi-centre intervention study after a low-energy diet in 2500 overweight, individuals with pre-diabetes (PREVIEW). *Diabetes Obes Metab* **20**, 2840-2851, doi:10.1111/dom.13466 (2018).
- 525 Kitzman, D. W. *et al.* Effect of Caloric Restriction or Aerobic Exercise Training on Peak Oxygen Consumption and Quality of Life in Obese Older Patients With Heart Failure With Preserved Ejection Fraction: A Randomized Clinical Trial. *JAMA* **315**, 36-46, doi:10.1001/jama.2015.17346 (2016).
- 526 Fontana, L. *et al.* Calorie restriction or exercise: effects on coronary heart disease risk factors. A randomized, controlled trial. *Am J Physiol Endocrinol Metab* **293**, E197-202, doi:10.1152/ajpendo.00102.2007 (2007).
- 527 Wei, M. *et al.* Fasting-mimicking diet and markers/risk factors for aging, diabetes, cancer, and cardiovascular disease. *Sci Transl Med* **9**, doi:10.1126/scitranslmed.aai8700 (2017).
- 528 Willcox, B. J. *et al.* Caloric restriction, the traditional Okinawan diet, and healthy aging: the diet of the world's longest-lived people and its potential impact on morbidity and life span. *Ann NY Acad Sci* **1114**, 434-455, doi:10.1196/annals.1396.037 (2007).
- 529 Fontana, L., Meyer, T. E., Klein, S. & Holloszy, J. O. Long-term calorie restriction is highly effective in reducing the risk for atherosclerosis in humans. *Proc Natl Acad Sci U S A* **101**, 6659-6663, doi:10.1073/pnas.0308291101 (2004).
- 530 Bhella, P. S. *et al.* Impact of lifelong exercise "dose" on left ventricular compliance and distensibility. *J Am Coll Cardiol* **64**, 1257-1266, doi:10.1016/j.jacc.2014.03.062 (2014).
- 531 Mandsager, K., Harb, S., Cremer, P., Phelan, D., Nissen, S. E. & Jaber, W. Association of Cardiorespiratory Fitness With Long-term Mortality Among Adults Undergoing Exercise Treadmill Testing. *JAMA Netw Open* **1**, e183605, doi:10.1001/jamanetworkopen.2018.3605 (2018).
- 532 Kokkinos, P. *et al.* Cardiorespiratory Fitness and Mortality Risk Across the Spectra of Age, Race, and Sex. *J Am Coll Cardiol* **80**, 598-609, doi:10.1016/j.jacc.2022.05.031 (2022).
- 533 Arbab-Zadeh, A. *et al.* Effect of aging and physical activity on left ventricular compliance. *Circulation* **110**, 1799-1805, doi:10.1161/01.CIR.0000142863.71285.74 (2004).
- 534 Look, A. R. G. *et al.* Cardiovascular effects of intensive lifestyle intervention in type 2 diabetes. *N Engl J Med* **369**, 145-154, doi:10.1056/NEJMoa1212914 (2013).
- 535 Ryan, D. H. *et al.* Look AHEAD (Action for Health in Diabetes): design and methods for a clinical trial of weight loss for the prevention of cardiovascular disease in type 2 diabetes. *Control Clin Trials* **24**, 610-628, doi:10.1016/s0197-2456(03)00064-3 (2003).
- 536 Bawamia, B. *et al.* Activation of telomerase by TA-65 enhances immunity and reduces inflammation post myocardial infarction. *Geroscience* **45**, 2689-2705, doi:10.1007/s11357-023-00794-6 (2023).
- 537 Goldberg, R. B. *et al.* Effects of Long-term Metformin and Lifestyle Interventions on Cardiovascular Events in the Diabetes Prevention Program and Its Outcome Study. *Circulation* **145**, 1632-1641, doi:10.1161/CIRCULATIONAHA.121.056756 (2022).

- 538 Wilding, J. P. H. *et al.* Once-Weekly Semaglutide in Adults with Overweight or Obesity. *N Engl J Med* **384**, 989-1002, doi:10.1056/NEJMoa2032183 (2021).
- 539 Lincoff, A. M. *et al.* Semaglutide and Cardiovascular Outcomes in Obesity without Diabetes. *N Engl J Med* **389**, 2221-2232, doi:10.1056/NEJMoa2307563 (2023).
- 540 Stahli, B. E. *et al.* Mammalian Target of Rapamycin Inhibition in Patients With ST-Segment Elevation Myocardial Infarction. *J Am Coll Cardiol* **80**, 1802-1814, doi:10.1016/j.jacc.2022.08.747 (2022).
- 541 Matsumoto, M., Kitada, Y. & Naito, Y. Endothelial Function is improved by Inducing Microbial Polyamine Production in the Gut: A Randomized Placebo-Controlled Trial. *Nutrients* **11**, doi:10.3390/nu11051188 (2019).
- 542 Yi, L. *et al.* The efficacy and safety of beta-nicotinamide mononucleotide (NMN) supplementation in healthy middle-aged adults: a randomized, multicenter, double-blind, placebo-controlled, parallel-group, dose-dependent clinical trial. *Geroscience* **45**, 29-43, doi:10.1007/s11357-022-00705-1 (2023).
- 543 Akbari, M. *et al.* The Effects of Resveratrol Supplementation on Endothelial Function and Blood Pressures Among Patients with Metabolic Syndrome and Related Disorders: A Systematic Review and Meta-Analysis of Randomized Controlled Trials. *High Blood Press Cardiovasc Prev* **26**, 305-319, doi:10.1007/s40292-019-00324-6 (2019).
- 544 Nidorf, S. M. *et al.* Colchicine in Patients with Chronic Coronary Disease. *N Engl J Med* **383**, 1838-1847, doi:10.1056/NEJMoa2021372 (2020).
- 545 Diguet, N. *et al.* Nicotinamide Riboside Preserves Cardiac Function in a Mouse Model of Dilated Cardiomyopathy. *Circulation* **137**, 2256-2273, doi:10.1161/CIRCULATIONAHA.116.026099 (2018).
- 546 de Picciotto, N. E. *et al.* Nicotinamide mononucleotide supplementation reverses vascular dysfunction and oxidative stress with aging in mice. *Aging Cell* **15**, 522-530, doi:10.1111/acel.12461 (2016).
- 547 Harrison, D. E. *et al.* 17-a-estradiol late in life extends lifespan in aging UM-HET3 male mice; nicotinamide riboside and three other drugs do not affect lifespan in either sex. *Aging Cell* **20**, e13328, doi:10.1111/acel.13328 (2021).
- 548 Zhang, H. *et al.* NAD(+) repletion improves mitochondrial and stem cell function and enhances life span in mice. *Science* **352**, 1436-1443, doi:10.1126/science.aaf2693 (2016).
- 549 Montegut, L. *et al.* High plasma concentrations of acyl-coenzyme A binding protein (ACBP) predispose to cardiovascular disease: Evidence for a phylogenetically conserved proaging function of ACBP. *Aging Cell* **22**, e13751, doi:10.1111/acel.13751 (2023).
- 550 Harrison, D. E. *et al.* Rapamycin fed late in life extends lifespan in genetically heterogeneous mice. *Nature* **460**, 392-395, doi:10.1038/nature08221 (2009).
- 551 Ximerakis, M. *et al.* Heterochronic parabiosis reprograms the mouse brain transcriptome by shifting aging signatures in multiple cell types. *Nat Aging* **3**, 327-345, doi:10.1038/s43587-023-00373-6 (2023).
- 552 Xu, M. *et al.* Senolytics improve physical function and increase lifespan in old age. *Nat Med* **24**, 1246-1256, doi:10.1038/s41591-018-0092-9 (2018).
- 553 Singh, P. *et al.* Taurine deficiency as a driver of aging. *Science* **380**, eabn9257, doi:10.1126/science.abn9257 (2023).
- 554 Widjaja, A. A. *et al.* Inhibition of IL-11 signalling extends mammalian healthspan and lifespan. *Nature* **632**, 157-165, doi:10.1038/s41586-024-07701-9 (2024).
- 555 Mamoshina, P. *et al.* Population Specific Biomarkers of Human Aging: A Big Data Study Using South Korean, Canadian, and Eastern European Patient Populations. *J Gerontol A Biol Sci Med Sci* **73**, 1482-1490, doi:10.1093/gerona/gly005 (2018).
- 556 Moqri, M. *et al.* Biomarkers of aging for the identification and evaluation of longevity interventions. *Cell* **186**, 3758-3775, doi:10.1016/j.cell.2023.08.003 (2023).

- 557 Macdonald-Dunlop, E. *et al.* A catalogue of omics biological ageing clocks reveals substantial commonality and associations with disease risk. *Aging (Albany NY)* **14**, 623-659, doi:10.18632/aging.203847 (2022).
- 558 Bao, H. *et al.* Biomarkers of aging. *Sci China Life Sci* **66**, 893-1066, doi:10.1007/s11427-023-2305-0 (2023).
- 559 Ganz, P. *et al.* Development and Validation of a Protein-Based Risk Score for Cardiovascular Outcomes Among Patients With Stable Coronary Heart Disease. *Jama* **315**, 2532-2541, doi:10.1001/jama.2016.5951 (2016).
- 560 Sayed, N. *et al.* An inflammatory aging clock (iAge) based on deep learning tracks multimorbidity, immunosenescence, frailty and cardiovascular aging. *Nat Aging* **1**, 598-615, doi:10.1038/s43587-021-00082-y (2021).
- 561 Alpert, A. *et al.* A clinically meaningful metric of immune age derived from high-dimensional longitudinal monitoring. *Nat Med* **25**, 487-495, doi:10.1038/s41591-019-0381-y (2019).
- 562 Raisi-Estabragh, Z. *et al.* Estimation of biological heart age using cardiovascular magnetic resonance radiomics. *Sci Rep* **12**, 12805, doi:10.1038/s41598-022-16639-9 (2022).
- 563 Salih, A. M. *et al.* Image-Based Biological Heart Age Estimation Reveals Differential Aging Patterns Across Cardiac Chambers. *J Magn Reson Imaging* **58**, 1797-1812, doi:10.1002/jmri.28675 (2023).
- 564 Ganau, A. *et al.* Echocardiographic heart ageing patterns predict cardiovascular and non-cardiovascular events and reflect biological age: the SardiNIA study. *Eur J Prev Cardiol*, doi:10.1093/eurjpc/zwad254 (2023).
- 565 Lima, E. M. *et al.* Deep neural network-estimated electrocardiographic age as a mortality predictor. *Nat Commun* **12**, 5117, doi:10.1038/s41467-021-25351-7 (2021).
- 566 Zhu, Z. *et al.* Retinal age gap as a predictive biomarker of stroke risk. *BMC Med* **20**, 466, doi:10.1186/s12916-022-02620-w (2022).
- 567 Zhu, Z. *et al.* Retinal age gap as a predictive biomarker for mortality risk. *Br J Ophthalmol* **107**, 547-554, doi:10.1136/bjophthalmol-2021-319807 (2023).
- 568 Raghu, V. K., Weiss, J., Hoffmann, U., Aerts, H. & Lu, M. T. Deep Learning to Estimate Biological Age From Chest Radiographs. *JACC Cardiovasc Imaging* **14**, 2226-2236, doi:10.1016/j.jcmg.2021.01.008 (2021).
- 569 Christoffersen, M., Frikkie-Schmidt, R., Schnohr, P., Jensen, G. B., Nordestgaard, B. G. & Tybjærg-Hansen, A. Visible age-related signs and risk of ischemic heart disease in the general population: a prospective cohort study. *Circulation* **129**, 990-998, doi:10.1161/circulationaha.113.001696 (2014).
- 570 Boutouyrie, P. *et al.* Aortic stiffness is an independent predictor of primary coronary events in hypertensive patients: a longitudinal study. *Hypertension* **39**, 10-15, doi:10.1161/hy0102.099031 (2002).
- 571 Mattace-Raso, F. U. *et al.* Arterial stiffness and risk of coronary heart disease and stroke: the Rotterdam Study. *Circulation* **113**, 657-663, doi:10.1161/circulationaha.105.555235 (2006).
- 572 Vlachopoulos, C., Aznaouridis, K. & Stefanadis, C. Prediction of cardiovascular events and all-cause mortality with arterial stiffness: a systematic review and meta-analysis. *J Am Coll Cardiol* **55**, 1318-1327, doi:10.1016/j.jacc.2009.10.061 (2010).
- 573 Ben-Shlomo, Y. *et al.* Aortic pulse wave velocity improves cardiovascular event prediction: an individual participant meta-analysis of prospective observational data from 17,635 subjects. *J Am Coll Cardiol* **63**, 636-646, doi:10.1016/j.jacc.2013.09.063 (2014).
- 574 van Sloten, T. T. *et al.* Local stiffness of the carotid and femoral artery is associated with incident cardiovascular events and all-cause mortality: the Hoorn study. *J Am Coll Cardiol* **63**, 1739-1747, doi:10.1016/j.jacc.2013.12.041 (2014).
- 575 Sutton-Tyrrell, K. *et al.* Elevated aortic pulse wave velocity, a marker of arterial stiffness, predicts cardiovascular events in well-functioning older adults. *Circulation* **111**, 3384-3390, doi:10.1161/circulationaha.104.483628 (2005).

- 576 Belizna, C. *et al.* Arterial stiffness and stroke in sickle cell disease. *Stroke* **43**, 1129-1130, doi:10.1161/strokeaha.111.635383 (2012).
- 577 Boonyasirinant, T. *et al.* Aortic stiffness is increased in hypertrophic cardiomyopathy with myocardial fibrosis: novel insights in vascular function from magnetic resonance imaging. *J Am Coll Cardiol* **54**, 255-262, doi:10.1016/j.jacc.2009.03.060 (2009).
- 578 Kang, S. *et al.* Relationship of arterial stiffness and early mild diastolic heart failure in general middle and aged population. *Eur Heart J* **31**, 2799-2807, doi:10.1093/eurheartj/ehq296 (2010).
- 579 Oh, J. *et al.* Association of Morning Hypertension Subtype With Vascular Target Organ Damage and Central Hemodynamics. *J Am Heart Assoc* **6**, doi:10.1161/jaha.116.005424 (2017).
- 580 Chirinos, J. A. *et al.* Impact of Diabetes Mellitus on Ventricular Structure, Arterial Stiffness, and Pulsatile Hemodynamics in Heart Failure With Preserved Ejection Fraction. *J Am Heart Assoc* **8**, e011457, doi:10.1161/jaha.118.011457 (2019).
- 581 Vasu, S. *et al.* Abnormal stress-related measures of arterial stiffness in middle-aged and elderly men and women with impaired fasting glucose at risk for a first episode of symptomatic heart failure. *J Am Heart Assoc* **4**, e000991, doi:10.1161/jaha.114.000991 (2015).
- 582 Cereda, C. W. *et al.* Endothelial dysfunction and arterial stiffness in ischemic stroke: the role of sleep-disordered breathing. *Stroke* **44**, 1175-1178, doi:10.1161/strokeaha.111.000112 (2013).
- 583 Kingwell, B. A., Waddell, T. K., Medley, T. L., Cameron, J. D. & Dart, A. M. Large artery stiffness predicts ischemic threshold in patients with coronary artery disease. *J Am Coll Cardiol* **40**, 773-779, doi:10.1016/s0735-1097(02)02009-0 (2002).
- 584 Guerin, A. P., Blacher, J., Pannier, B., Marchais, S. J., Safar, M. E. & London, G. M. Impact of aortic stiffness attenuation on survival of patients in end-stage renal failure. *Circulation* **103**, 987-992, doi:10.1161/01.cir.103.7.987 (2001).
- 585 Cardoso, C. R. L. & Salles, G. F. Prognostic Value of Changes in Aortic Stiffness for Cardiovascular Outcomes and Mortality in Resistant Hypertension: a Cohort Study. *Hypertension* **79**, 447-456, doi:10.1161/hypertensionaha.121.18498 (2022).
- 586 Kosmopoulos, M. *et al.* The relationship between telomere length and putative markers of vascular ageing: A systematic review and meta-analysis. *Mech Ageing Dev* **201**, 111604, doi:10.1016/j.mad.2021.111604 (2022).
- 587 Ojeda-Rodriguez, A. *et al.* Association between telomere length and intima-media thickness of both common carotid arteries in patients with coronary heart disease: From the CORDIOPREV randomized controlled trial. *Atherosclerosis* **380**, 117193, doi:10.1016/j.atherosclerosis.2023.117193 (2023).
- 588 Paul, T. K. *et al.* Framingham risk score is associated with femoral artery intima-media thickness in asymptomatic young adults (the Bogalusa heart study). *Atherosclerosis* **213**, 627-631, doi:10.1016/j.atherosclerosis.2010.09.026 (2010).
- 589 Willeit, P. *et al.* Carotid Intima-Media Thickness Progression as Surrogate Marker for Cardiovascular Risk: Meta-Analysis of 119 Clinical Trials Involving 100 667 Patients. *Circulation* **142**, 621-642, doi:10.1186/1468-6708-6-3 (2020).
- 590 Noflatscher, M. *et al.* Influence of Traditional Cardiovascular Risk Factors on Carotid and Femoral Atherosclerotic Plaque Volume as Measured by Three-Dimensional Ultrasound. *J Clin Med* **8**, doi:10.3390/jcm8010032 (2018).
- 591 Li, W. *et al.* Joint effects of carotid plaques and renal impairment on the risk of cardiovascular disease and all-cause death in a community-based population: The Kailuan cohort study. *Front Cardiovasc Med* **9**, 943718, doi:10.3389/fcvm.2022.943718 (2022).
- 592 Peng, A. W. *et al.* Long-Term All-Cause and Cause-Specific Mortality in Asymptomatic Patients With CAC \geq 1,000: Results From the CAC Consortium. *JACC Cardiovasc Imaging* **13**, 83-93, doi:10.1016/j.jcmg.2019.02.005 (2020).

- 593 Greenland, P. *et al.* 2010 ACCF/AHA guideline for assessment of cardiovascular risk in asymptomatic adults: executive summary: a report of the American College of Cardiology Foundation/American Heart Association Task Force on Practice Guidelines. *Circulation* **122**, 2748-2764, doi:10.1161/CIR.0b013e3182051bab (2010).
- 594 Kronmal, R. A. *et al.* Risk factors for the progression of coronary artery calcification in asymptomatic subjects: results from the Multi-Ethnic Study of Atherosclerosis (MESA). *Circulation* **115**, 2722-2730, doi:10.1161/circulationaha.106.674143 (2007).
- 595 de Lemos, J. A. *et al.* Association of troponin T detected with a highly sensitive assay and cardiac structure and mortality risk in the general population. *Jama* **304**, 2503-2512, doi:10.1001/jama.2010.1768 (2010).
- 596 Saunders, J. T. *et al.* Cardiac troponin T measured by a highly sensitive assay predicts coronary heart disease, heart failure, and mortality in the Atherosclerosis Risk in Communities Study. *Circulation* **123**, 1367-1376, doi:10.1161/circulationaha.110.005264 (2011).
- 597 Saeed, A. *et al.* Short-Term Global Cardiovascular Disease Risk Prediction in Older Adults. *J Am Coll Cardiol* **71**, 2527-2536, doi:10.1016/j.jacc.2018.02.050 (2018).
- 598 Li, Y. *et al.* Hs-CRP and all-cause, cardiovascular, and cancer mortality risk: A meta-analysis. *Atherosclerosis* **259**, 75-82, doi:10.1016/j.atherosclerosis.2017.02.003 (2017).
- 599 Yoshida, Y. *et al.* Alteration of Cardiac Performance and Serum B-Type Natriuretic Peptide Level in Healthy Aging. *J Am Coll Cardiol* **74**, 1789-1800, doi:10.1016/j.jacc.2019.07.080 (2019).
- 600 York, M. K. *et al.* B-Type Natriuretic Peptide Levels and Mortality in Patients With and Without Heart Failure. *J Am Coll Cardiol* **71**, 2079-2088, doi:10.1016/j.jacc.2018.02.071 (2018).
- 601 Kuh, D., Cooper, R., Sattar, N., Welsh, P., Hardy, R. & Ben-Shlomo, Y. Systemic Inflammation and Cardio-Renal Organ Damage Biomarkers in Middle Age Are Associated With Physical Capability Up to 9 Years Later. *Circulation* **139**, 1988-1999, doi:10.1161/circulationaha.118.037332 (2019).
- 602 Markousis-Mavrogenis, G. *et al.* The clinical significance of interleukin-6 in heart failure: results from the BIOSTAT-CHF study. *Eur J Heart Fail* **21**, 965-973, doi:10.1002/ejhf.1482 (2019).
- 603 Sonel, A., Sasseen, B. M., Fineberg, N., Bang, N. & Wilensky, R. L. Prospective study correlating fibrinopeptide A, troponin I, myoglobin, and myosin light chain levels with early and late ischemic events in consecutive patients presenting to the emergency department with chest pain. *Circulation* **102**, 1107-1113, doi:10.1161/01.cir.102.10.1107 (2000).