

How to live for centuries: common denominators of organisms with exceptional longevity

Stefania E. Kapsetaki¹, Nektarios Tavernarakis^{1,2}

¹Institute of Molecular Biology and Biotechnology, Foundation for Research and Technology-Hellas, Heraklion 70013, Crete, Greece

²Division of Basic Sciences, School of Medicine, University of Crete, Heraklion 71003, Crete, Greece

Correspondence to: Nektarios Tavernarakis; email: tavernarakis@imbb.forth.gr

Keywords: aging, healthspan, interspecific variation, intraspecific variation, stemness

Received: February 2, 2026

Accepted: August 6, 2026

Published: September 8, 2026

Copyright: © 2026 Kapsetaki and Tavernarakis. This is an open access article distributed under the terms of the [Creative Commons Attribution License](https://creativecommons.org/licenses/by/4.0/) (CC BY 4.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

ABSTRACT

Organisms vary in their lifespan. Understanding this variation may help us live healthier and longer. Here, we focus on species living twice as long as humans, or more. Out of the 101 multicellular species with a maximum lifespan of 250+ years, 11 are animals and 90 are plants. We surveyed the genetic, transcriptional, proteomic, metabolomic, regeneration-, stress-, and cancer-related components of intraspecific and interspecific lifespan variation, across these species. We examined whether the mechanisms regulating intraspecific lifespan variation across these species are the same or different from mechanisms regulating interspecific lifespan variation. We identified several similarities: both types of variation include mechanisms related to DNA maintenance, stemness, and stress management. Such mechanisms are also typical of early developmental stages and germ cells. Nonetheless, caution should be exercised when attempting to draw robust conclusions based on available data, given the lack of in-depth molecular studies on the healthspan and lifespan across thousands of individuals and species, the methodological variation across published studies, and our partial understanding of the interplay between physiology and the environment across species.

INTRODUCTION

Each species consists of numerous individuals, with some living less than others. Over a dozen different definitions of a species have, historically, been put forward [1–3]. More recently, a unified concept of a species has been introduced, such that a species is a separately evolving population consisting of sub-populations [4]. We use these conventions to refer to specific species in this article. Certain molecular processes shape the lifespan of each species and each individual. Deciphering these processes, which underlie the diversity in lifespan across and within species can help us better understand how they evolved and why. Such knowledge may inform the design of effective strategies towards increasing human and other species' healthspan and lifespan. Healthspan broadly refers to

the time that an organism is healthy, i.e., does not suffer from diseases or disabilities, whereas lifespan refers to the time that the organism exists from birth to death [5]. Several studies have investigated the molecular biology of lifespan variation within and across different groups of species (reviewed in [6–11]). However, a survey of recent developments regarding the molecular components regulating the variation in lifespan within and across extremely long-lived (e.g., of at least 250 years old) animal and plant species is lacking.

Here, we consider 101 species that live past 250 years, including the longest-living beings on Earth (Figure 1). Among these species, 90 are plants and only 11 are animals. Some barely reach an age of 250 years, while others live for up to 80,000 years. Few, such as the freshwater planarian and the *Turritopsis dohrnii* jellyfish,

are considered potentially immortal [12, 13]. There are only a handful of species, that live for over 10,000 years. These are mostly animals (four out of the six species, with the rest being plants). None of these species are mammals; indeed, no known mammal lives for over 250 years.

We reviewed the literature on the molecular and cellular mechanisms implicated in lifespan variation within and among extremely long-lived multicellular species. We aimed to examine whether the lifespan-extending mechanisms that extremely long-lived organisms have

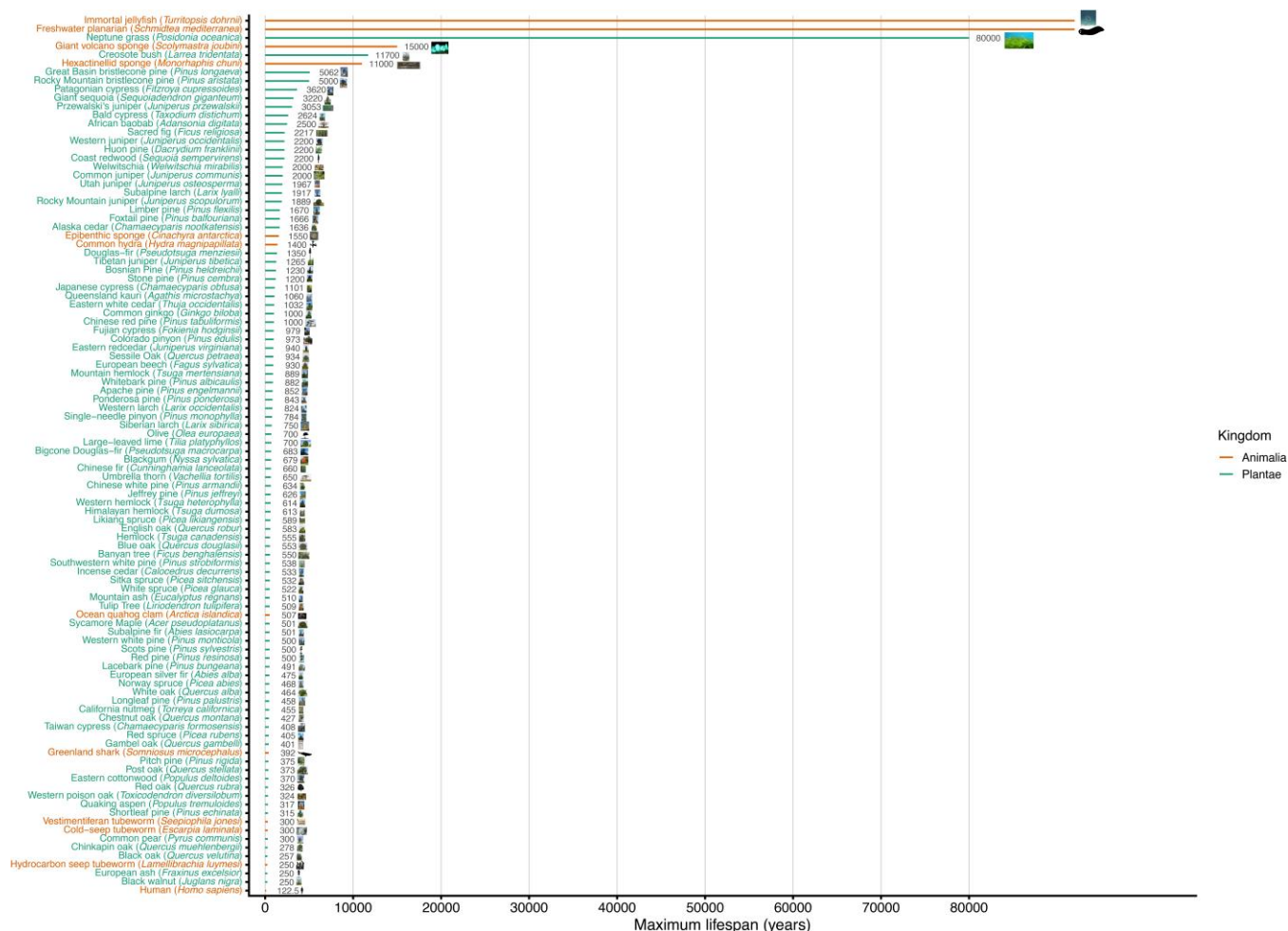


Figure 1. 101 species with a maximum lifespan of at least 250 years. The maximum lifespan of humans is noted for comparison with other species. We obtained most maximum lifespan values from the AnAge database [14]. For the following species, maximum lifespan values were found in the indicated references: *Juglans nigra*, *Fraxinus excelsior*, *Pinus sylvestris*, *Tsuga canadensis*, *Nyssa sylvatica*, *Olea europaea*, *Fagus sylvatica*, *Agathis microstachya*, *Pinus cembra*, *Juniperus communis*, *Dacrydium franklinii*, *Quercus velutina*, *Quercus montana*, *Quercus rubra*, *Quercus alba*, *Pinus rigida*, *Pyrus communis* [15], *Fitzroya cupressoides* [16], *Juniperus przewalskii* [17], *Pinus heldreichii*, *Quercus petraea* [18], *Acer pseudoplatanus* [19], *Liriodendron tulipifera* [20], *Quercus douglasii*, *Taxodium distichum* [21], *Posidonia oceanica* [22], *Pinus monophylla*, *Pseudotsuga macrocarpa* [23], *Pinus monticola*, *Pinus resinosa*, *Pinus aristata* [24], *Hydra magnipapillata* [25], *Monorhaphis chuni* [26], *Pinus tabuliformis* [27], *Welwitschia mirabilis* [28], *Ficus benghalensis* [29], and *Larrea tridentata* [30]. For the remaining species shown on the figure that lack maximum lifespan values in the AnAge database, we found lifespan data in ref. [31]. Numbers on the side of each bar show the value of maximum lifespan for each species. The freshwater planarian *Schmidtea mediterranea* does not have a reported maximum lifespan in the AnAge database, and is considered potentially immortal [32]. Likewise, the jellyfish *Turritopsis dohrnii* is considered potentially immortal [13]. We obtained the species images included from Phylopic, or from online resources under the creative commons license. Due to unavailability of Creative Commons Licensed images for *Escarpiia laminata*, *Cinachya antarctica* and *Juniperus tibetica*, we used Creative Commons Licensed images of their sister species *Escarpiia southwardae*, *Cinachya australiensis* and *Juniperus virginiana*, respectively. The image of *Scolymastra joubini* is taken from ref. [33]. The image of *Juniperus tibetica* is taken from https://www.inaturalist.org/taxa/135889-Juniperus-tibetica/browse_photos. The figure was generated using R.

evolved over millennia are distinct, or bear discernible similarities. Our analysis incorporated available genetic, genomic, transcriptomic and metabolomic data, focusing on findings relevant to telomere maintenance, stress resistance, regeneration, and tumorigenesis processes. These analyses have yielded a wealth of information about the genes and molecules shaping organismal physiology and the adaptations that underlie the variation in lifespan among extremely long-lived individuals.

Vulnerability to cellular, tissue and organismal stress may also change the lifespan of organisms through a complex network of molecular and cellular interactions [34–37]. For example, oxidative stress responses involve over 70 molecular pathways, including apoptosis, mitophagy, FoxO signaling, JAK-STAT signaling, Toll-like receptor signaling, and HIF-1 signaling [38]. In addition, the ability of cells to regenerate gradually diminishes in aging organisms. This decline is thought to be an evolutionary adaptation preventing uncontrollable cell overproliferation, e.g., cancer [39]. Thus, understanding potential intraspecific and interspecific variations in the regulation of regenerative downfall can aid in understanding intraspecific and interspecific variations in lifespan of extremely long-lived individuals. It is important to examine these mechanisms in extremely long-lived species, given their known role in lifespan variation across long-lived species (reviewed in [10]). For example – across various species of mammals, birds, or fish – long-lived species have a higher ratio of tumor suppressor gene:oncogene copies [40], lower somatic mutation rates [41], less telomere shortening [42], shorter telomeres [43, 44], longer telomeres [45], no significant difference in telomere length compared to shorter-lived species [46], higher concentrations of antioxidants [47], lower concentrations of reactive oxygen species [47], higher resistance to stress [48], fewer polyunsaturated triacylglycerols [49], and increased levels of mono-unsaturated fatty acids [50].

Finally, we discuss the translatability and the implications of studies involving species of exceptional longevity on human health and aging. We also note that the mechanisms regulating lifespan variation within and between ≥ 250 -year-old species are somewhat similar. Key features of extremely long-lived organisms are enhanced regeneration and DNA maintenance capacity, as well as better resistance to stress, oxidative damage, and cancer. Further studies with many more individuals and species, consistent methodology, and inclusion of environmental parameters could strengthen our understanding of intra- and inter-species lifespan variation.

Components of intraspecific lifespan variation

Genomic analyses

Studies of the genetic variation among individuals of extremely long-lived species found some genetic variation, selection for stress-related genes, and chromosomal rearrangements, depending on the species (Table 1). For example, a genomic region of 12 cytochrome b sequences was sequenced from six populations of ocean quahog clams; each population from a different region in northern Europe. Large genetic variation (80%) was detected within the populations, but negligible variation was observed between the populations (2%), indicating that geographic location had a marginal effect on genetic variation [51]. Among the genomes of 545 trees of the species *Ginkgo biloba* from 51 populations across the world, specific genes were selected over evolutionary time. These genes are involved in managing abiotic (e.g., low temperatures, high concentrations of salt, and dehydration) and biotic stress. There was no correlation between diversity at the genetic level versus variation in morphology, or their geographical distance [52]. In freshwater planarians, large chromosomal rearrangements possibly regulate lifespan. Comparison of the asexual (which is also considered immortal) and sexual *Schmidtea mediterranea* show that the former has a translocated chromosome, whereas the latter does not have this rearrangement [53, 54]. Specifically, in the asexual strain, a fragment from one of the 1st pair of chromosomes has moved to one of the 3rd pair of chromosomes [53]. It is worth noting that these studies are an important initial step in understanding what regulates the variation in lifespan within extremely long-lived species. However, they do not establish a direct causal link between specific genetic components and lifespan regulation, within these species.

Transcriptomics

Comparisons of transcriptomic data between individuals, within two potentially immortal species, show that differences in the regulation of DNA maintenance and stemness pathways may play an important role in lifespan variation, within these species. The jellyfish *Turritopsis dohrnii* can reverse its development and become larvae again, potentially achieving immortality [6, 13]. Matsumoto and colleagues compared the transcriptome of multiple jellyfish individuals at different developmental stages. Animals at the cyst stage (early in development) overexpressed genes related to DNA repair, telomere maintenance, and telomere organization, and underexpressed genes related to aging, compared to other stages. For example,

Table 1. Indicative characteristics of long- versus short-lived individuals, within extremely long-lived species.

Species	Sample size (# individuals in all experimental conditions of an assay)	Experimental approach	Characteristic features	Reference
<i>Arctica islandica</i>	18-180 depending on the assay	Molecular	No significant differences in the metabolic rate nor activity of antioxidant enzymes	[55]
<i>Arctica islandica</i>	at least 50 per assay	Metabolomics	Less accumulation of lipofuscin	[56]
<i>Arctica islandica</i>	62 or 25 depending on the assay	Molecular	No significant differences in the length of telomeres nor in the activity of telomerase	[57]
<i>Arctica islandica</i>	51 per assay	Molecular	No significant differences in the activity of the mitochondrial electron transfer chain, or in the susceptibility of mitochondria to oxidative damage	[58]
<i>Pinus longaeva</i>	7-9 depending on the assay	Molecular	Longer telomeres and increased activity of telomerase in the roots; no significant differences in the length of telomeres and lower activity of telomerase in needles	[24]
<i>Schmidtea mediterranea</i>	17 or 20 depending on the assay	Molecular	Better maintenance of telomeres	[12]
<i>Schmidtea mediterranea</i>	at least 24	Cytogenetics	Chromosomal translocations	[53, 54]

the cyst stage expressed 831 transcripts related to DNA repair whereas the other life stages expressed 574-742 DNA-repair-related transcripts [59]. In another extremely long-lived organism, hydra, the transcription factor FoxO has been implicated in maintaining the organism's ability to regenerate and reach potential immortality. Specifically, transcriptomic comparison of hydra individuals, with silenced versus non-silenced foxO gene, showed that the former had fewer terminally differentiated cells in the foot region of the hydra and more stem cell proliferation than the latter. FoxO regulates the expression of genes, such as *cnivi*, and *cnivi* is differentially expressed in somatic versus stem cells. Mature nematocytes of hydra do not express *cnivi*, whereas their interstitial stem cells do [60].

Metabolomics

Studies between individuals of *Arctica islandica* that varied in their lifespans showed that metabolic rate did not explain a significant amount of variation in the individuals' lifespans. By contrast, shorter-lived individuals had higher levels of an aging-related marker, compared to longer-lived individuals. Specifically, when studying different populations of *Arctica islandica*, Basova and colleagues found that while metabolic rate could not explain the variation in lifespan of the examined individuals, the levels of the autofluorescent age pigment lipofuscin that accumulates in lysosomes of old cells, are higher in the mantle and gills of individuals with shorter lifespan compared to those with longer lifespan [55, 56]. This points to the

potential ability of longer-lived *Arctica islandica* individuals to withstand aging-related damage.

Telomere maintenance

The length of telomeres and telomerase activity does not seem to consistently explain the variation in lifespan among individuals of extremely long-lived species. For example, the telomere length and telomerase activity among individuals of *Arctica islandica* did not correlate significantly with the different lifespans of these individuals [57]. Among different trees of the species *Pinus longaeva*, ranging from 20 to 3,500 years old, the length of telomeres and the activity of telomerase varied depending on the tissue of the tree. The roots of old trees had longer telomeres, compared to the roots of young trees, whereas the needles of old and younger trees did not show significant telomere length differences. The roots of old trees appeared to have an increased activity of telomerase, whereas the needles and the core samples of the old trees had lower activity of telomerase relative to younger trees [24]. When comparing the length of telomeres over time in asexual versus sexual strains of the freshwater planarian *Schmidtea mediterranea*, asexual strains which are considered immortal are better able to maintain the ends of their telomeres, compared to the sexually reproducing strains [12].

Stress resistance

Individuals of exceptionally long-lived species that vary in their lifespan do not show significant variation in the

way they handle oxidative stress. Out of all the types of stresses, only oxidative stress is mentioned here, given the absence of relevant literature on associations between intraspecific lifespan variation and other types of stresses, in extremely long-lived species. The activity of antioxidant enzymes was not significantly correlated with age, in a study where different individuals of the species *Arctica islandica* were examined [55]. The activity of the electron transport system in mitochondria, and the susceptibility of the mitochondrial membrane to damage caused by reactive oxygen species did not differ significantly among young versus old *Arctica islandica* individuals [58]. In addition, although hypoxia resistance has often been linked to longevity [61], varying oxygen concentrations did not increase the lifespan of *Arctica islandica* individuals, compared to normoxia [55].

Components of interspecific lifespan variation

Genomic analyses

Across diverse species, comparisons including extremely long-lived species have almost always shown that longer-lived species are characterized by a higher number of genes involved in DNA maintenance, and abiotic and biotic stress resistance, compared to shorter-lived species. In studies of animals, for example, it has been shown that the extremely long-lived *Lamellibrachia luymesii* has more copies of SOD genes, which are involved in protecting cellular damage from oxidative stress, than other shorter-lived species of Lophotrochozoa [62]. Three SOD2 proteins were detected in a proteomic analysis of *Lamellibrachia luymesii* [62], though it is unknown whether the extra SOD2 gene copies provide increased resistance to oxidative stress in this species. *Arctica islandica* has the highest percentage of repeats and a higher total number of genes, relatively to other shorter-lived bivalve species [63]. Additionally, by comparing the dN/dS ratio of short- versus long-lived bivalve species, Iannello and colleagues [64] found evolutionary convergence in many genes of long-lived species. These genes are involved in pathways regulating DNA damage repair, the resistance to viruses, and environmental insults, as well as pathways regulating apoptosis, and tumor suppression. The same study also identified 43 genes that were episodically positively selected in long-lived versus short-lived species of bivalves. These genes are involved in protein homeostasis, protein folding, the response to hypoxia, oxidative stress resistance, and apoptosis. *Turritopsis dohrnii* has more copies of replication regulating, DNA damage repair, oxidative stress resistance, and apoptosis-regulating genes, compared to the non-immortal *Turritopsis rubra*. Specifically, some of these

genes included the DNA polymerase delta and alpha, DNA topoisomerase III beta, X-ray repair cross complementing 5, RAD51 paralog C, gene endonuclease homolog 1, thioredoxin, glutathione reductase GSR, presenilin 1, and bone morphogenetic protein 7 [65]. The number of expressed gene copies is higher in *Turritopsis dohrnii* than *Turritopsis rubra* [65]. However, it is unknown whether in *Turritopsis dohrnii* the corresponding protein levels are higher than *Turritopsis rubra*, and whether this increased gene copy number leads to improved DNA replication and repair, resistance to oxidative stress, and tighter regulation of apoptosis, in *Turritopsis dohrnii*, compared to *Turritopsis rubra*.

In plants, the extreme lifespan of some species may be partly explained by increased copies of genes related to DNA damage response and stress resistance, but not to genome size or ploidy. Specifically, when comparing the genomes of 61 plant species, Aoyagi Blue and colleagues [66] found that certain extremely long-lived species have more copies of PARP genes, which are involved in responding to DNA damage. Specifically, *Pseudotsuga menziesii*, *Pinus sylvestris*, and *Malus domestica* had the highest copy number of PARPs [66]. A genomic comparison of three plant species, with *Sequoiadendron giganteum* being the longest-lived among the three, showed that *Sequoiadendron giganteum* has relatively more genes involved in the synthesis of flavonoids or tannins [67]. These tannins may help this species resist fires and thus help it live longer. Positive selection of genes, related to abiotic (e.g., osmosis, dehydration) and biotic (e.g., pathogens) stress resistance, in the extremely long-lived *Ficus benghalensis* and *Ficus religiosa* relative to other angiosperm species, may explain the extreme longevity of these two species [68]. The extremely long-lived *Ginkgo biloba* has almost five times more copies of genes related to the synthesis of metabolites, such as terpenes and flavonoids, involved in defenses against pathogens, than the shorter-lived *Arabidopsis thaliana* [69, 70]. However, genome size did not seem to explain the variation in the lifespans of plant species [71]. Polyploidy also does not appear to be associated with longevity in a study of hundreds of gymnosperm species [72].

Metabolomics

The ability to recycle proteins and the levels of oxidative stress resistance-related proteins do not appear to explain the variation in longevity among species. However, in some species the increased production of metabolites, such as monoterpenes, may help increase their lifespan. Longer-lived species are expected to be

better able to eliminate damaged proteins. To test whether the longer-lived species *Arctica islandica* are better at eliminating damaged proteins, compared to the shorter-lived *Mercenaria mercenaria*, Ungvari et al. compared the activity of three proteasome proteases in these two species. In the adductor muscle, the baseline activities of the three proteases did not differ significantly between the two species. In the gills, two out of the three proteases did not have significantly different activity between the two species, but surprisingly the activity of caspase was higher in the shorter-lived species than the longer-lived species. Thus, the extreme longevity of *Arctica islandica* relative to *Mercenaria mercenaria* cannot be clearly attributed to enhanced ability to recycle proteins [73]. The Greenland shark does not differ significantly in terms of the levels of glutathione peroxidase, an enzyme that protects cells from oxidative damage, in its red blood cells and muscles, compared with other, shorter-lived vertebrate species [74]. Moving to the plant kingdom, *Pinus longaeva* has higher maximum lifespan, compared to *Pinus balfouriana* and *Pinus flexilis*, and also displays a higher concentration of monoterpenes than the other two species [75]. Monoterpenes help plants defend themselves against pathogens [76, 77]. Thus, monoterpenes may be contributing to the increased longevity of *Pinus longaeva* relative to *Pinus balfouriana* and *Pinus flexilis*.

Stress resistance

Extremely long-lived species of some trees appear to have stronger stress responses, and, similarly, extremely long-lived clams appear to have properties that minimize the accumulation of oxidative damage in comparison to shorter-lived species. The extremely long-lived tree *Tilia platyphyllos* appears to have a much stronger response to the stress of urban life than other shorter-lived species. Specifically, comparing the stress responses of three plant species, *Celtis occidentalis*, *Platanus hispanica*, and the extremely long-lived *Tilia platyphyllos*, living in roads with lots of traffic versus less traffic, showed that, out of all the antioxidant enzymes (APX, CAT) and biomarkers of oxidative stress (TBARS, proline, GSH) measured, *Tilia platyphyllos* had a significantly higher concentration of all these compounds in the urban, versus the low traffic environment, whereas the other two species did not show a significant increase in all of these tested compounds [78]. Considering animals, *Arctica islandica* produce much lower amounts of the toxic H₂O₂ than the relatively shorter-lived clam *Mya arenaria* [79]. *Arctica islandica* has comparatively very low peroxidation index of its mitochondrial membrane. This indicates that this species may have a relatively better ability to prevent damage of its mitochondrial

membranes, and thus this may, in part, explain this species' relatively longer lifespan [80]. *Arctica islandica* are also relatively more resistant to death from stressors, such as DNA methylation and alkylation agents, in comparison to *Mercenaria mercenaria* [81].

Regeneration

The ability of some extremely long-lived species to regenerate and maintain a non-aging population of stem cells may contribute to their extreme longevity. In comparison to other species, sponges, corals, hydra, jellyfish, and planaria display remarkable ability to regenerate and maintain an active population of stem cells [6]. These properties likely enable them to live so long [6]. This is similar to the ability of germ cells to be transferred from one generation to the next, and to minimize aging-related damage, thus, almost achieving immortality. For example, hydra regularly (approximately every 20 days) replaces all of its cells, including aged cells [82]. *Turritopsis dohrnii* shows a unique capacity to reverse to a younger developmental stage [6]. The neoblasts of planaria are stem cells that circulate around the body of the flatworm and are able to generate any cell type [83], even after the worm has been cut into 279 pieces [84]. Furthermore, although the pluripotency of cells in the meristem region in the roots of the short-lived plant species *Arabidopsis thaliana* decays as they age [85], this decay does not appear in the longer-lived *Pinus sylvestris* and *Ginkgo biloba* [86, 87].

Tumorigenesis

Tumors are not very common in extremely long-lived species (Figure 1), relative to others, and cancer in these species has never been reported. Tumors have been found in several cacti [88], but no case of cancer has been reported in extremely long-lived plants [89, 90]. Tumors are also uncommon in *Arctica islandica* and *Margaritifera margaritifera* [91] relative to other bivalves [92]. Nevertheless, tumors that grew spontaneously have been found in lab strains of hydra [93]. In planaria, tumors have not been observed to develop spontaneously. The capacity of these planaria to not develop as many tumors as expected, based on their lifelong regenerative ability, has been attributed to enhanced DNA repair mechanisms [94], as well as, oncogenes and tumor suppressors, such as PTEN, AKT, p53, and Rb, that also regulate stem cell division [95–99]. However, tumors in planaria can be induced using mammalian carcinogens [100, 101]. The sparsity of reported tumors in these extremely long-lived species indicates that these species may have evolved anticancer mechanisms worthy of future investigations for human anticancer treatments. Other studies,

analyzing a range of vertebrate species, well below the 250-year maximum lifespan with ≥ 20 individual necropsy records per species, also support potential enhanced anticancer mechanisms in long-lived species, as indicated by the absence of a correlation between maximum lifespan and cancer prevalence [102–105] or cancer risk [106].

Caveats and next steps

As is evident from the studies described above, the molecular processes determining the variation in longevity within and across species have not been systematically investigated across a large number of species on the tree of life, or a large number of individuals within a given species. Therefore, our knowledge is fragmented, at best. For example, Iannello and colleagues [64] emphasized that they based their analyses on only a few genes. Apart from speculations about the remarkable ability of potentially immortal species to regenerate, it is not fully understood exactly why some species have extremely long lifespans or are potentially immortal. In other words, it is not known what particular molecular mechanisms position these species to have a variation in their life history traits that is shifted towards extreme longevity. Future studies, when more and higher quality genome sequences of extremely long-lived species become available, will allow more methodical genomic and transcriptomic analyses towards untangling the molecular mechanisms that enable organisms to live extremely long. Such a broader-scale investigation across species would increase statistical power as highlighted by Rechsteiner and colleagues [10], and would reveal whether the mechanisms that determine the variation in longevity, within and across species, are clade specific or apply to species across the tree of life. Unfortunately, there is very limited molecular mechanistic information for extremely long-lived plants and separately for extremely long-lived animals to allow conclusions about potential differences in the cellular and molecular mechanisms underlying longevity of extremely long-lived species between these two kingdoms. There are few studies per kingdom, and in some cases, there is no information at all about extremely long-lived plants. It remains to be found whether the processes that are thought to be essential for extreme longevity in some immortal species (e.g., regular regeneration in hydra, sponges, and planaria) are also essential for other extremely long-lived species for which relatively less research has been conducted (e.g., some extremely long-lived plants and the hexactinellid sponge).

While there is a lot of data on maximum longevity that allows examining the variation in maximum lifespan

across species (the AnAge database; [14]), there is no similar database of healthspan records within and across species. Future studies aiming to build such a database would be useful, as they could, perhaps, reveal new molecular pathways/components that contribute to increased healthspan. This is particularly important given the ongoing efforts to find ways of increasing healthspan in humans [107, 108].

Rechsteiner and colleagues concluded that “no current genetic model fully explains the variation observed across species” [10]. One explanation for this limitation could be that the variation in each study's approaches, scope, and number of species is preventing us from detecting a common genetic model of lifespan variation that could be applied across species. Previous studies aiming to discern the molecular mechanisms of lifespan variation, within and across species, have not all investigated the same species, nor used the same methods. The type of tissue sampled in each experiment may have had a large confounding effect on the interpretation of results from comparative studies. For example, a study comparing the markers of longevity across individuals and species, based on collected liver samples may show very different molecular markers, compared to a study of collected epithelial samples. Each tissue may likely be expressing different molecular markers at a given time point in different individuals and different species. Therefore, variation in the results of these studies could emerge due to methodological differences and examination of different species. Future studies ought to address this issue by using many more species and using consistent methodology to examine whether similarities or differences in the molecular and cellular mechanisms linked to longevity, truly contribute to longevity or have been associated to it due to other confounding variables. Unless we try to minimize this noise coming from attempting to compare dissimilar studies that employ different methodologies, it will be difficult to conclude with confidence whether or not a particular genetic/physiological model can fully capture the variation observed across species. Moreover, the molecular mechanisms explaining the variation in longevity within and across species may be slightly or largely different between laboratory-maintained, versus wild species. Various parameters in laboratory conditions may be different in the wild, and these parameters may affect which genes are expressed, and to what extent, in each environment, thus, affecting individual and species maximum lifespan. Therefore, future studies should carefully take into account the environmental conditions, where these organisms have been living, in order to more accurately pinpoint the components that affect lifespan.

The exact effective population size of every species and the evolutionary pressures that species experience is largely unknown. Therefore, the evolutionary forces that shape the lifespan of each species are unclear. It would be important to unravel these evolutionary forces in order to understand the life trajectory that each organism is likely to follow at the molecular level and, thus, improve strategies that prevent the development of age-related diseases in these organisms. Several environmental components may be affecting the lifespan of exceptionally long-lived species. For example, extreme cold temperatures may have led to unique temperature-dependent molecular pathways contributing to lifespan extension in some species; e.g., the bivalve species *Arctica islandica* found in arctic waters live up to 507 years and the Greenland shark lives up to 392 years. However, we must be cautious with generalizing the impact of environmental factors on physiology to all species, since, obviously, the same environmental conditions do not have the same effect on all species. The cold-sensitive *Hydra oligactis* ages at low temperatures (around 10° C), whereas it is considered immortal at higher temperatures (around 22° C) [109, 110]. By investigating the life-time records of extremely long-lived species, we could identify environmental conditions that extended their lifespans in specific periods and thus elucidate new interactions between the environment and molecular processes that increase the likelihood of living, in any given year. In the case of plants, environmental changes can be tracked by inspecting the rings of the trunks of long-lived trees [17]. The growth of the bivalve *Arctica islandica* each year for hundreds of years can be tracked by measuring the thickness of lines on its shell [111]. Understanding what environmental conditions extend the growth of this bivalve consistently in several years, and if a particular sequence of line thickness is linked to some individuals living longer than others, can reveal new environmental factors that contribute to lifespan extension. Overall, the interplay between environmental pressures and molecular biology leading to the evolution of extreme lifespans is largely unknown and worthy of future research.

Finally, research on extremely long-lived species may inform efforts to extend healthspan and lifespan of other species. Understanding the molecular and cell biology of extreme longevity, may facilitate translational approaches targeting human pathophysiology. Indeed, ginkgo tree root and leaves extracts are already used to prevent and treat aging-related diseases [112]. Leaf extracts from the extremely long-lived *Larrea tridentata* have anti-apoptotic effects on human neuroblastoma cells [113]. Additionally, olive oil from the extremely long-lived *Olea europaea* has shown many anti-aging benefits for humans and is integral to the Mediterranean

diet which has been associated with longevity and increased healthspan [114, 115]. Expression of the elephant *TP53* gene in human cancer cells increases apoptosis of these cells [116]. This indicates that expressing tumor-suppressor genes, found in multiple copies in extremely long-lived species relative to shorter-lived species, in human cells might have anti-aging effects in human cells.

CONCLUSIONS

Here, we reviewed the available literature on the molecular mechanisms associated with the variation in lifespan within and across extremely long-lived species. These studies converge to indicate that, at least for the species investigated, variations in the lifespan of individuals within a particular species is largely not due to variation in the length of telomeres, the activity of telomerase, the metabolic rates, the capacity to handle oxidative stress, or other large genetic changes. By contrast, the variation in lifespan within species seems to be mostly due to selection for specific stress-related genes and small chromosomal rearrangements, differences in how the DNA is maintained, and the expression of certain genes regulating stemness. When comparing extremely long-lived to shorter-lived species, we find that longer-lived species are often characterized by better DNA maintenance, improved ability to withstand abiotic and biotic stress, to minimize the effect of oxidative damage, to regenerate and maintain a minimally-aging population of stem cells, and to suppress cancer development (Table 2). Hence, the underpinnings of lifespan variation within and across extremely long-lived organisms show specific similarities. Both types of variation include mechanisms related to DNA maintenance, stemness, and stress resistance-related pathways. These pathways are all also characteristic of early developmental stages and germ cells. Living young appears to be a strategy of living long. These processes are ancient and have likely persisted over billions of years from LUCA (Last Universal Common Ancestor) to extant organisms [117, 118]. However, the vast majority of species with extremely long lifespans shown in Figure 1 are understudied in terms of the molecular biology sustaining their long lifespans.

In our article, we also strived to highlight areas that are only partially covered by available findings and require further investigation. First, collecting more molecular and healthspan data from many more individuals and species across the tree of life will contribute towards discovering potentially new anti-aging mechanisms. Second, we ought to be watchful of methodological differences, including effects of growing organisms in the lab versus species in the wild, when attempting to

Table 2. Indicative characteristics of extremely long-lived, compared to shorter-lived species.

Species	Sample size (# species)	Experimental approach	Characteristic features	Reference
<i>Lamellibrachia luymesii</i> vs. shorter-lived species of Lophotrochozoa	at least 40	Genomics analyses	Increased SOD gene copy number (protection from oxidative damage)	[62]
<i>Somniosus microcephalus</i> vs. other vertebrate species	24	Proteomics analyses	No significantly higher levels of glutathione peroxidase in red blood cells and muscles	[74]
<i>Arctica islandica</i> vs. shorter-lived bivalves	10	Genomics analyses	Higher percentage of DNA repeats and higher gene number	[63]
<i>Arctica islandica</i> vs. <i>Mercenaria mercenaria</i>	2	Proteomics analyses	No significant difference in the capacity to recycle proteins	[73]
<i>Arctica islandica</i> vs. <i>Mya arenaria</i>	2	Molecular genetics	Lower concentration of H ₂ O ₂	[79]
<i>Tilia platyphyllos</i> vs. <i>Celtis occidentalis</i> and <i>Platanus</i> × <i>hispanica</i>	3	Molecular genetics	Higher concentrations of all tested antioxidant enzymes and oxidative stress biomarkers in conditions of high traffic	[78]
<i>Arctica islandica</i> vs. <i>Mercenaria mercenaria</i>	2	Phenotypic characterization	Elevated resistance against death from alkylating (e.g., methyl methanesulfonate) and methylating (e.g., nitrogen mustard) agents	[81]
<i>Arctica islandica</i> and <i>Margaritifera margaritifera</i> vs. other bivalve species	at least 11	Observational characterization, Histology	Less tumor formation	[91, 92]
33 bivalve species, including the long-lived <i>Arctica islandica</i> , <i>Margaritifera margaritifera</i> , <i>Elliptio complanata</i> , and <i>Lampsilis siliquoidea</i>	33	Transcriptomics analyses	Convergent genetic pathways for DNA damage repair, tumor suppression, apoptosis, protein homeostasis, protein folding, hypoxia and oxidative stress resistance, defense against viruses and environmental stressors	[64]
61 plant species, including gymnosperms, angiosperms, perennial and annual herbs, and algae	61	Genomics analyses	Higher number of PARP genes (DNA damage repair)	[66]
Plant species, including <i>Ginkgo biloba</i> , <i>Sequoiadendron giganteum</i> , and <i>Pinus tabulaeformis</i>	1,224	Genomics analyses	Similar genome size	[71]
Gymnosperms	614	Cytogenetics	Similar ploidy	[72]
<i>Metasequoia glyptostroboides</i> , <i>Sequoia sempervirens</i> , and <i>Sequoiadendron giganteum</i>	3	Genomics analyses	Higher number of genes involved in flavonoid/tannin synthesis	[67]
<i>Ficus benghalensis</i> and <i>Ficus religiosa</i> vs. other angiosperm species	52	Genomics	Genes related to abiotic and biotic stress resistance are positively selected	[68]
<i>Ginkgo biloba</i> vs. <i>Arabidopsis thaliana</i>	2	Genomics analyses	Nearly five times more genes involved in terpene and flavonoid biosynthesis	[69, 70]
<i>Turritopsis dohrnii</i> vs. <i>Turritopsis rubra</i>	2	Genomics analyses	Higher number of genes involved in DNA replication and repair, oxidative stress responses, and apoptosis	[65]
<i>Pinus longaeva</i> vs. <i>Pinus balfouriana</i> and <i>Pinus flexilis</i>	3	Metabolomics analyses	Higher concentration of monoterpenes	[75]
<i>Pinus sylvestris</i> and <i>Ginkgo biloba</i> vs. shorter-lived plants	3	Transcriptomics analyses, Molecular genetics, Epigenomics analyses	Lower decay of cell pluripotency in the meristem during aging	[85–87]

draw conclusions from different studies. Third, we should investigate the role of the environment in more detail, including the effect of variation in species' population sizes, over evolutionary time scales, on age-related genes. Finally, we suggest that our options of increasing human healthspan and lifespan could be expanded by investing in research that translates findings from extremely long-lived species to human physiology.

AUTHOR CONTRIBUTIONS

S.E.K. and N.T. conceptualized the work, S.E.K. wrote the draft of the manuscript and prepared the display items. N.T. revised and edited the manuscript.

ACKNOWLEDGMENTS

We thank our colleagues at the Neurogenetics and Ageing Laboratory of the Institute of Molecular Biology and Biotechnology, for their valuable feedback and insightful comments, during the preparation of our manuscript. We acknowledge that not all relevant, published studies could be included in this review, and apologize to those colleagues, whose work we could not refer to, due to space limitations.

CONFLICTS OF INTEREST

The authors declare that they have no conflicts of interest.

FUNDING

Work in the authors' laboratory is funded by the Hellenic Foundation for Research and Innovation (H.F.R.I.) project NeuroMitophagy (grant agreement no.: HFRI—FM17C3- 0869), the H.F.R.I. project NeuroFlame (grant agreement no.:15546 80145), the General Secretariat for Research and Innovation of the Greek Ministry of Development and the European Union – NextGenerationEU project BrainPrecision (grant agreement no.: TAEDR-0535850), and the European Commission Research Executive Agency Excellence Hub “CHAgeing” (grant agreement no.: 101087071).

REFERENCES

- De Queiroz K. A unified concept of species and its consequences for the future of taxonomy. *Proc Calif Acad Sci.* 2005; 56:196–215. https://www.academia.edu/6796621/A_Unified_Concept_of_Species_and_Its_Consequences_for_the_Future_of_Taxonomy
- Rieseberg LH, Brouillet L. Are many plant species paraphyletic? *Taxon.* Wiley. 1994; 43:21–32. <https://doi.org/10.2307/1223457>
- Mallet J. A species definition for the modern synthesis. *Trends Ecol Evol.* 1995; 10:294–9. [https://doi.org/10.1016/0169-5347\(95\)90031-4](https://doi.org/10.1016/0169-5347(95)90031-4) PMID:21237047
- De Queiroz K. Species concepts and species delimitation. *Syst Biol.* 2007; 56:879–86. <https://doi.org/10.1080/10635150701701083> PMID:18027281
- Masfiah S, Kurnialandi A, Meij JJ, Maier AB. Definitions of healthspan: A systematic review. *Ageing Res Rev.* 2025; 111:102806. <https://doi.org/10.1016/j.arr.2025.102806> PMID:40523529
- Petralia RS, Mattson MP, Yao PJ. Aging and longevity in the simplest animals and the quest for immortality. *Ageing Res Rev.* 2014; 16:66–82. <https://doi.org/10.1016/j.arr.2014.05.003> PMID:24910306
- da Silva R, Conde DA, Baudisch A, Colchero F. Slow and negligible senescence among testudines challenges evolutionary theories of senescence. *Science.* 2022; 376:1466–70. <https://doi.org/10.1126/science.abl7811> PMID:35737795
- Cohen AA. Aging across the tree of life: The importance of a comparative perspective for the use of animal models in aging. *Biochim Biophys Acta Mol Basis Dis.* 2018; 1864:2680–9. <https://doi.org/10.1016/j.bbadis.2017.05.028> PMID:28690188
- Ma S, Gladyshev VN. Molecular signatures of longevity: Insights from cross-species comparative studies. *Semin Cell Dev Biol.* 2017; 70:190–203. <https://doi.org/10.1016/j.semcdb.2017.08.007> PMID:28800931
- Rechsteiner C, Morandini F, Kim SJ, Seluanov A, Gorbunova V. Unlocking longevity through the comparative biology of aging. *Nat Aging.* 2025; 5:1686–703. <https://doi.org/10.1038/s43587-025-00945-8> PMID:40877537
- Batalova AY, Krutovsky KV. Genetic and Epigenetic Mechanisms of Longevity in Forest Trees. *Int J Mol Sci.* 2023; 24:10403. <https://doi.org/10.3390/ijms241210403> PMID:37373550
- Tan TC, Rahman R, Jaber-Hijazi F, Felix DA, Chen C, Louis EJ, Aboobaker A. Telomere maintenance and telomerase activity are differentially regulated in

- asexual and sexual worms. *Proc Natl Acad Sci USA*. 2012; 109:4209–14.
<https://doi.org/10.1073/pnas.1118885109>
PMID:22371573
13. Mas-Bargues C. Translating lessons from immortal models: Hydra and the immortal jellyfish. *Rev Esp Geriatr Gerontol*. 2025; 60:101624.
<https://doi.org/10.1016/j.regg.2025.101624>
PMID:39889309
14. de Magalhães JP, Costa J. A database of vertebrate longevity records and their relation to other life-history traits. *J Evol Biol*. 2009; 22:1770–4.
<https://doi.org/10.1111/j.1420-9101.2009.01783.x>
PMID:19522730
15. Thomas H. Senescence, ageing and death of the whole plant. *New Phytol*. 2013; 197:696–711.
<https://doi.org/10.1111/nph.12047> PMID:23176101
16. Lara A, Villalba R. A 3620-Year Temperature Record from Fitzroya cupressoides Tree Rings in Southern South America. *Science*. 1993; 260:1104–6.
<https://doi.org/10.1126/science.260.5111.1104>
PMID:17806339
17. Yang B, Qin C, Wang J, He M, Melvin TM, Osborn TJ, Briffa KR. A 3,500-year tree-ring record of annual precipitation on the northeastern Tibetan Plateau. *Proc Natl Acad Sci USA*. 2014; 111:2903–8.
<https://doi.org/10.1073/pnas.1319238111>
PMID:24516152
18. Piovesan G, Biondi F. On tree longevity. *New Phytol*. 2021; 231:1318–37.
<https://doi.org/10.1111/nph.17148> PMID:33305422
19. Biondi F. Development of a tree-ring network for the Italian Peninsula. *Tree-Ring Bulletin*. 1992; 52:15–29.
<https://repository.arizona.edu/server/api/core/bitstreams/0de2abc3-c506-4fb0-81f2-63231d841e83/content>
20. Pederson N. External characteristics of old trees in the Eastern deciduous forest. *Natural Areas Journal*. 2010; 30:396–407.
<https://doi.org/10.3375/043.030.0405>
21. Stahle DW, Edmondson JR, Howard IM, Robbins CR, Griffin RD, Carl A, Hall CB, Stahle DK, Torbenson MCA. Longevity, climate sensitivity, and conservation status of wetland trees at Black River, North Carolina. *Environ Res Commun*. 2019; 1:041002.
<https://doi.org/10.1088/2515-7620/ab0c4a>
22. de Witte LC, Stöcklin J. Longevity of clonal plants: why it matters and how to measure it. *Ann Bot*. 2010; 106:859–70.
<https://doi.org/10.1093/aob/mcq191> PMID:20880935
23. Biondi F, Meko DM, Piovesan G. Maximum tree lifespans derived from public-domain dendro-chronological data. *iScience*. 2023; 26:106138.
<https://doi.org/10.1016/j.isci.2023.106138>
PMID:36926654
24. Flanary BE, Kletetschka G. Analysis of telomere length and telomerase activity in tree species of various life-spans, and with age in the bristlecone pine *Pinus longaeva*. *Biogerontology*. 2005; 6:101–11.
<https://doi.org/10.1007/s10522-005-3484-4>
PMID:16034678
25. Jones OR, Scheuerlein A, Salguero-Gómez R, Camarda CG, Schaible R, Casper BB, Dahlgren JP, Ehrlén J, García MB, Menges ES, Quintana-Ascencio PF, Caswell H, Baudisch A, Vaupel JW. Diversity of ageing across the tree of life. *Nature*. 2014; 505:169–73.
<https://doi.org/10.1038/nature12789>
PMID:24317695
26. Jochum KP, Wang X, Vennemann TW, Sinha B, Müller WEG. Siliceous deep-sea sponge *Monorhaphis chuni*: A potential paleoclimate archive in ancient animals. *Chem Geol*. 2012; 300–301:143–51.
<https://doi.org/10.1016/j.chemgeo.2012.01.009>
27. Overview of Ancient and Precious Trees in Beijing. Beijing Municipality.
https://yllhj.beijing.gov.cn/English/Resources/202107/t20210720_2445104.html
28. Notten A. *Welwitschia mirabilis* Hook.f. (= *W. bainesii* (Hook.f.) Carr.). South African National Biodiversity Institute. 2016. <https://pza.sanbi.org/welwitschia-mirabilis>
29. Bar-Ness YD. The world's largest trees? Cataloguing India's giant banyans. *Outreach Ecology*. 2010.
<https://www.outreachecology.com/landmark/media/pdfs/TheWorldsLargestTrees-CataloguingIndiasGiantBanyans-byYDBar-Ness-OutreachEcologyReport-Jun10.pdf>
30. Vasek FC. Creosote bush: Long-lived clones in the Mojave Desert. *Am J Bot*. Wiley. 1980; 67:246–55.
<https://doi.org/10.1002/j.1537-2197.1980.tb07648.x>
31. Brown PM. OLDLIST, a database of old trees. Rocky Mountain Tree-Ring Research.
<https://www.rmtrr.org/oldlist.htm>
32. Dalyell JG. Observations on some interesting phenomena in animal physiology, exhibited by several species of Planariae. Edinburgh: Archibald Constable. 1814.
<https://doi.org/10.5962/bhl.title.19795>
33. Dayton PK, Kim S, Jarrell SC, Oliver JS, Hammerstrom K, Fisher JL, O'Connor K, Barber JS, Robilliard G, Barry J, Thurber AR, Conlan K. Recruitment, growth and mortality of an Antarctic hexactinellid sponge, *Anoxycalyx joubini*. *PLoS One*. 2013; 8:e56939.

- <https://doi.org/10.1371/journal.pone.0056939>
PMID:23460822
34. Kutschera U. Tissue stresses in growing plant organs. *Physiol Plant*. Wiley. 1989; 77:157–63.
<https://doi.org/10.1111/j.1399-3054.1989.tb05992.x>
 35. Gusev EY, Zotova NV. Cellular Stress and General Pathological Processes. *Curr Pharm Des*. 2019; 25:251–97.
<https://doi.org/10.2174/138161282566619031911464>
1 PMID:31198111
 36. Sapolsky RM. Organismal stress and telomeric aging: an unexpected connection. *Proc Natl Acad Sci USA*. 2004; 101:17323–4.
<https://doi.org/10.1073/pnas.0408041101>
PMID:15579535
 37. Monaghan P. Organismal stress, telomeres and life histories. *J Exp Biol*. 2014; 217:57–66.
<https://doi.org/10.1242/jeb.090043> PMID:24353204
 38. Liu R, Bian Y, Liu L, Liu L, Liu X, Ma S. Molecular pathways associated with oxidative stress and their potential applications in radiotherapy. *Int J Mol Med*. 2022; 49:65.
<https://doi.org/10.3892/ijmm.2022.5121>
PMID:35293589
 39. West MD, Sternberg H, Labat I, Janus J, Chapman KB, Malik NN, de Grey AD, Larocca D. Toward a unified theory of aging and regeneration. *Regen Med*. 2019; 14:867–86.
<https://doi.org/10.2217/rme-2019-0062>
PMID:31455183
 40. Baines C, Meitern R, Kreitsberg R, Sepp T. Comparative study of the evolution of cancer gene duplications across fish. *Evol Appl*. 2022; 15:1834–45.
<https://doi.org/10.1111/eva.13481> PMID:36426117
 41. Cagan A, Baez-Ortega A, Brzozowska N, Abascal F, Coorens TH, Sanders MA, Lawson AR, Harvey LM, Bhosle S, Jones D, Alcantara RE, Butler TM, Hooks Y, et al. Somatic mutation rates scale with lifespan across mammals. *Nature*. 2022; 604:517–24.
<https://doi.org/10.1038/s41586-022-04618-z>
PMID:35418684
 42. Pepke ML, Eisenberg DTA. Accounting for phylogenetic relatedness in cross-species analyses of telomere shortening rates. *Exp Results*. 2020; 1:e11.
<https://doi.org/10.1017/exp.2020.18>
 43. Pepke ML, Eisenberg DT. On the comparative biology of mammalian telomeres: Telomere length co-evolves with body mass, lifespan and cancer risk. *Mol Ecol*. 2022; 31:6286–96.
<https://doi.org/10.1111/mec.15870>
PMID:33662151
 44. Gomes NM, Ryder OA, Houck ML, Charter SJ, Walker W, Forsyth NR, Austad SN, Venditti C, Pagel M, Shay JW, Wright WE. Comparative biology of mammalian telomeres: hypotheses on ancestral states and the roles of telomeres in longevity determination. *Aging Cell*. 2011; 10:761–8.
<https://doi.org/10.1111/j.1474-9726.2011.00718.x>
PMID:21518243
 45. Domínguez-de-Barros A, Sifaoui I, Borecka Z, Dorta-Guerra R, Lorenzo-Morales J, Castro-Fuentes R, Córdoba-Lanús E. An approach to the effects of longevity, sexual maturity, and reproduction on telomere length and oxidative stress in different Psittacidae species. *Front Genet*. 2023; 14:1156730.
<https://doi.org/10.3389/fgene.2023.1156730>
PMID:37021005
 46. Buddhachat K, Brown JL, Kaewkool M, Poommouang A, Kaewmong P, Kittiwattanawong K, Nganvongpanit K. Life Expectancy in Marine Mammals Is Unrelated to Telomere Length but Is Associated With Body Size. *Front Genet*. 2021; 12:737860.
<https://doi.org/10.3389/fgene.2021.737860>
PMID:34630527
 47. Xia C, Møller AP. Long-lived birds suffer less from oxidative stress. *Avian Res*. 2018; 9:41.
<https://doi.org/10.1186/s40657-018-0133-6>
 48. Harper JM, Wang M, Galecki AT, Ro J, Williams JB, Miller RA. Fibroblasts from long-lived bird species are resistant to multiple forms of stress. *J Exp Biol*. 2011; 214:1902–10.
<https://doi.org/10.1242/jeb.054643>
PMID:21562178
 49. Ma S, Yim SH, Lee SG, Kim EB, Lee SR, Chang KT, Buffenstein R, Lewis KN, Park TJ, Miller RA, Clish CB, Gladyshev VN. Organization of the Mammalian Metabolome according to Organ Function, Lineage Specialization, and Longevity. *Cell Metab*. 2015; 22:332–43.
<https://doi.org/10.1016/j.cmet.2015.07.005>
PMID:26244935
 50. Galván I, Naudí A, Erritzøe J, Møller AP, Barja G, Pamplona R. Long lifespans have evolved with long and monounsaturated fatty acids in birds. *Evolution*. 2015; 69:2776–84.
<https://doi.org/10.1111/evo.12754>
PMID:26294378
 51. Begum S, Brey T, Held C. Lack of association between genetical and morphological variations for bivalve *Arctica islandica* from six different sites of the NE Atlantic ocean. *Estuarine, Coastal and Shelf Science*. 2018; 212:34–9.
<https://doi.org/10.1016/j.ecss.2018.06.014>

52. Zhao YP, Fan G, Yin PP, Sun S, Li N, Hong X, Hu G, Zhang H, Zhang FM, Han JD, Hao YJ, Xu Q, Yang X, et al. Resequencing 545 ginkgo genomes across the world reveals the evolutionary history of the living fossil. *Nat Commun.* 2019; 10:4201.
<https://doi.org/10.1038/s41467-019-12133-5>
PMID:[31519986](https://pubmed.ncbi.nlm.nih.gov/31519986/)
53. Baguña J, Carranza S, Pala M, Ribera C, Giribet G, Arnedo MA, Ribas M, Riutort M. From morphology and karyology to molecules. New methods for taxonomical identification of asexual populations of freshwater planarians. A tribute to Professor Mario Benazzi. *Ital J Zool (Modena).* 1999; 66:207–14.
<https://doi.org/10.1080/11250009909356258>
54. De Vries EJ, Baguña J, Ball IR. Chromosomal polymorphism in planarians (Turbellaria, Tricladida) and the plate tectonics of the western Mediterranean. *Genetica.* 1984; 62:187–91.
<https://doi.org/10.1007/BF00056435>
55. Basova L, Begum S, Strahl J, Sukhotin A, Brey T, Philipp E, Abele D. Age-dependent patterns of antioxidants in *Arctica islandica* from six regionally separate populations with different life spans. *Aquatic Biology.* 2012; 14:141–52.
<https://doi.org/10.3354/ab00387>
56. Basova L, Strahl J, Philipp EER, Brey T, Sukhotin A, Abele D. Lipofuscin accumulation in tissues of *Arctica islandica* indicates faster ageing in populations from brackish environments. *Mar Biol.* 2017; 164.
<https://doi.org/10.1007/s00227-017-3110-4>
57. Gruber H, Schaible R, Ridgway ID, Chow TT, Held C, Philipp EE. Telomere-independent ageing in the longest-lived non-colonial animal, *Arctica islandica*. *Exp Gerontol.* 2014; 51:38–45.
<https://doi.org/10.1016/j.exger.2013.12.014>
PMID:[24394156](https://pubmed.ncbi.nlm.nih.gov/24394156/)
58. Rodríguez E, Déglétagne C, Hagen TM, Abele D, Blier PU. Mitochondrial Traits Previously Associated With Species Maximum Lifespan Do Not Correlate With Longevity Across Populations of the Bivalve *Arctica islandica*. *Front Physiol.* 2019; 10:946.
<https://doi.org/10.3389/fphys.2019.00946>
PMID:[31404340](https://pubmed.ncbi.nlm.nih.gov/31404340/)
59. Matsumoto Y, Piraino S, Miglietta MP. Transcriptome Characterization of Reverse Development in *Turritopsis dohrnii* (Hydrozoa, Cnidaria). *G3 (Bethesda).* 2019; 9:4127–38.
<https://doi.org/10.1534/g3.119.400487>
PMID:[31619459](https://pubmed.ncbi.nlm.nih.gov/31619459/)
60. Boehm AM, Khalturin K, Anton-Erxleben F, Hemmrich G, Klostermeier UC, Lopez-Quintero JA, Oberg HH, Puchert M, Rosenstiel P, Wittlieb J, Bosch TC. FoxO is a critical regulator of stem cell maintenance in immortal Hydra. *Proc Natl Acad Sci USA.* 2012; 109:19697–702.
<https://doi.org/10.1073/pnas.1209714109>
PMID:[23150562](https://pubmed.ncbi.nlm.nih.gov/23150562/)
61. Krivoruchko A, Storey KB. Forever young: mechanisms of natural anoxia tolerance and potential links to longevity. *Oxid Med Cell Longev.* 2010; 3:186–98.
<https://doi.org/10.4161/oxim.3.3.12356>
PMID:[20716943](https://pubmed.ncbi.nlm.nih.gov/20716943/)
62. Li Y, Tassia MG, Waits DS, Bogantes VE, David KT, Halanych KM. Genomic adaptations to chemosymbiosis in the deep-sea seep-dwelling tubeworm *Lamellibrachia luymesii*. *BMC Biol.* 2019; 17:91.
<https://doi.org/10.1186/s12915-019-0713-x>
PMID:[31739792](https://pubmed.ncbi.nlm.nih.gov/31739792/)
63. Gerhard GS, Allard JB, Kaniper S, Lynch D, Lee H, Kumar S. Genome Assembly of *Arctica islandica*, the Longest-Lived Non-Colonial Animal Species. *Animals (Basel).* 2025; 15:690.
<https://doi.org/10.3390/ani15050690>
PMID:[40075973](https://pubmed.ncbi.nlm.nih.gov/40075973/)
64. Iannello M, Forni G, Piccinini G, Xu R, Martellosi J, Ghiselli F, Milani L. Signatures of Extreme Longevity: A Perspective from Bivalve Molecular Evolution. *Genome Biol Evol.* 2023; 15:evad159.
<https://doi.org/10.1093/gbe/evad159>
PMID:[37647860](https://pubmed.ncbi.nlm.nih.gov/37647860/)
65. Pascual-Torner M, Carrero D, Pérez-Silva JG, Álvarez-Puente D, Roiz-Valle D, Bretones G, Rodríguez D, Maeso D, Mateo-González E, Español Y, Mariño G, Acuña JL, Quesada V, López-Otín C. Comparative genomics of mortal and immortal cnidarians unveils novel keys behind rejuvenation. *Proc Natl Acad Sci USA.* 2022; 119:e2118763119.
<https://doi.org/10.1073/pnas.2118763119>
PMID:[36037356](https://pubmed.ncbi.nlm.nih.gov/36037356/)
66. Aoyagi Blue Y, Kusumi J, Satake A. Copy number analyses of DNA repair genes reveal the role of poly(ADP-ribose) polymerase (PARP) in tree longevity. *iScience.* 2021; 24:102779.
<https://doi.org/10.1016/j.isci.2021.102779>
PMID:[34278274](https://pubmed.ncbi.nlm.nih.gov/34278274/)
67. Fu F, Song C, Wen C, Yang L, Guo Y, Yang X, Shu Z, Li X, Feng Y, Liu B, Sun M, Zhong Y, Chen L, et al. The Metasequoia genome and evolutionary relationships among redwoods. *Plant Commun.* 2023; 4:100643.
<https://doi.org/10.1016/j.xplc.2023.100643>
PMID:[37381601](https://pubmed.ncbi.nlm.nih.gov/37381601/)
68. Chakraborty A, Mahajan S, Bisht MS, Sharma VK. Genome sequencing and comparative analysis of *Ficus benghalensis* and *Ficus religiosa* species reveal evolutionary mechanisms of longevity. *iScience.* 2022;

- 25:105100.
<https://doi.org/10.1016/j.isci.2022.105100>
 PMID:[36164650](https://pubmed.ncbi.nlm.nih.gov/36164650/)
69. Cui J, Li X, Lu Z, Jin B. Plant secondary metabolites involved in the stress tolerance of long-lived trees. *Tree Physiol.* 2024; 44:tpae002.
<https://doi.org/10.1093/treephys/tpae002>
 PMID:[38196002](https://pubmed.ncbi.nlm.nih.gov/38196002/)
 70. Liu S, Meng Z, Zhang H, Chu Y, Qiu Y, Jin B, Wang L. Identification and characterization of thirteen gene families involved in flavonoid biosynthesis in *Ginkgo biloba*. *Ind Crops Prod.* 2022; 188:115576.
<https://doi.org/10.1016/j.indcrop.2022.115576>
 71. Liu S, Xu H, Wang G, Jin B, Cao F, Wang L. Tree Longevity: Multifaceted Genetic Strategies and Beyond. *Plant Cell Environ.* 2025; 48:244–59.
<https://doi.org/10.1111/pce.15146> PMID:[39254418](https://pubmed.ncbi.nlm.nih.gov/39254418/)
 72. Rastogi S, Ohri D. Chromosome numbers in gymnosperms - an update. *Silvae Genet.* 2020; 69:13–9.
<https://doi.org/10.2478/sg-2020-0003>
 73. Ungvari Z, Ridgway I, Philipp EE, Campbell CM, McQuary P, Chow T, Coelho M, Didier ES, Gelino S, Holmbeck MA, Kim I, Levy E, Sosnowska D, et al. Extreme longevity is associated with increased resistance to oxidative stress in *Arctica islandica*, the longest-living non-colonial animal. *J Gerontol A Biol Sci Med Sci.* 2011; 66:741–50.
<https://doi.org/10.1093/gerona/glr044>
 PMID:[21486920](https://pubmed.ncbi.nlm.nih.gov/21486920/)
 74. Costantini D, Smith S, Killen SS, Nielsen J, Steffensen JF. The Greenland shark: A new challenge for the oxidative stress theory of ageing? *Comp Biochem Physiol A Mol Integr Physiol.* 2017; 203:227–32.
<https://doi.org/10.1016/j.cbpa.2016.09.026>
 PMID:[27717642](https://pubmed.ncbi.nlm.nih.gov/27717642/)
 75. Bentz BJ, Hood SM, Hansen EM, Vandygriff JC, Mock KE. Defense traits in the long-lived Great Basin bristlecone pine and resistance to the native herbivore mountain pine beetle. *New Phytol.* 2017; 213:611–24.
<https://doi.org/10.1111/nph.14191>
 PMID:[27612209](https://pubmed.ncbi.nlm.nih.gov/27612209/)
 76. Marei GIK, Abdel Rasoul MA, Abdelgaleil SAM. Comparative antifungal activities and biochemical effects of monoterpenes on plant pathogenic fungi. *Pestic Biochem Physiol.* Elsevier BV. 2012; 103:56–61.
<https://doi.org/10.1016/j.pestbp.2012.03.004>
 77. Rasoul MAA, Marei GIK, Abdelgaleil SAM. Evaluation of antibacterial properties and biochemical effects of monoterpenes on plant pathogenic bacteria. *Afr J Microbiol Res [Internet]. Academic Journals;* 2012; 6:3667–72.
<https://doi.org/10.5897/AJMR12.118>
 78. Arsenov D, Borišev M, Nikolić N, Horak R, Pajević S. Physiological and Biochemical Indicators of Urban Environmental Stress in *Tilia*, *Celtis*, and *Platanus*: A Functional Trait-Based Approach. *Plants (Basel).* 2025; 14:3451.
<https://doi.org/10.3390/plants14223451>
 PMID:[41304602](https://pubmed.ncbi.nlm.nih.gov/41304602/)
 79. Buttemer WA, Abele D, Costantini D. From bivalves to birds: oxidative stress and longevity: Oxidative stress and longevity. *Funct Ecol.* 2010; 24:971–83.
<https://doi.org/10.1111/j.1365-2435.2010.01740.x>
 80. Munro D, Blier PU. The extreme longevity of *Arctica islandica* is associated with increased peroxidation resistance in mitochondrial membranes. *Aging Cell.* 2012; 11:845–55.
<https://doi.org/10.1111/j.1474-9726.2012.00847.x>
 PMID:[22708840](https://pubmed.ncbi.nlm.nih.gov/22708840/)
 81. Ungvari Z, Sosnowska D, Mason JB, Gruber H, Lee SW, Schwartz TS, Brown MK, Storm NJ, Fortney K, Sowa J, Byrne AB, Kurz T, Levy E, et al. Resistance to genotoxic stresses in *Arctica islandica*, the longest living noncolonial animal: is extreme longevity associated with a multistress resistance phenotype? *J Gerontol A Biol Sci Med Sci.* 2013; 68:521–9.
<https://doi.org/10.1093/gerona/gls193>
 PMID:[23051979](https://pubmed.ncbi.nlm.nih.gov/23051979/)
 82. Mueller WA, Hassel M, Greal M. Development and Reproduction in Humans and Animal Model Species. Berlin, Heidelberg: Springer Berlin Heidelberg. 2015. p. 585–93.
<https://doi.org/10.1007/978-3-662-43784-1>
 83. Baguñà J, Romero R. Quantitative analysis of cell types during growth, degrowth and regeneration in the planarians *Dugesia mediterranea* and *Dugesia tigrina*. *Hydrobiologia.* 1981; 84:181–94.
<https://doi.org/10.1007/BF00026179>
 84. Morgan TH. Experimental studies of the regeneration of *Planaria maculata*. *Dev Genes Evol.* 1898; 7:364–97.
<https://doi.org/10.1007/BF02161491>
 85. Wang Y, Kumaishi K, Suzuki T, Ichihashi Y, Yamaguchi N, Shirakawa M, Ito T. Morphological and Physiological Framework Underlying Plant Longevity in *Arabidopsis thaliana*. *Front Plant Sci.* 2020; 11:600726.
<https://doi.org/10.3389/fpls.2020.600726>
 PMID:[33224176](https://pubmed.ncbi.nlm.nih.gov/33224176/)
 86. Mencuccini M, Oñate M, Peñuelas J, Rico L, Munné-Bosch S. No signs of meristem senescence in old Scots pine. *J Ecol.* 2014; 102:555–65.
<https://doi.org/10.1111/1365-2745.12219>
 87. Wang L, Cui J, Jin B, Zhao J, Xu H, Lu Z, Li W, Li X, Li L,

- Liang E, Rao X, Wang S, Fu C, et al. Multifeature analyses of vascular cambial cells reveal longevity mechanisms in old Ginkgo biloba trees. *Proc Natl Acad Sci USA*. 2020; 117:2201–10.
<https://doi.org/10.1073/pnas.1916548117>
PMID:31932448
88. White PR. Neoplastic growth in plants. *Q Rev Biol*. 1951; 26:1–16.
<https://doi.org/10.1086/397879> PMID:14828053
 89. Doonan JH, Sablowski R. Walls around tumours - why plants do not develop cancer. *Nat Rev Cancer*. 2010; 10:794–802.
<https://doi.org/10.1038/nrc2942> PMID:20966923
 90. Doonan J, Hunt T. Cell cycle. Why don't plants get cancer? *Nature*. 1996; 380:481–2.
<https://doi.org/10.1038/380481a0> PMID:8606760
 91. Philipp EE, Abele D. Masters of longevity: lessons from long-lived bivalves--a mini-review. *Gerontology*. 2010; 56:55–65.
<https://doi.org/10.1159/000221004>
PMID:19468199
 92. Peters EC, Yevich PP, Harshbarger JC, Zaroogian GE. Comparative histopathology of gonadal neoplasms in marine bivalve mollusks. *Dis Aquat Organ*. 1994; 20:59–76.
<https://doi.org/10.3354/dao020059>
 93. Boutry J, Buysse M, Tissot S, Cazevielle C, Hamede R, Dujon AM, Ujvari B, Giraudeau M, Klimovich A, Thomas F, Tököllyi J. Spontaneously occurring tumors in different wild-derived strains of hydra. *Sci Rep*. 2023; 13:7449.
<https://doi.org/10.1038/s41598-023-34656-0>
PMID:37156860
 94. Oviedo NJ, Beane WS. Regeneration: The origin of cancer or a possible cure? *Semin Cell Dev Biol*. 2009; 20:557–64.
<https://doi.org/10.1016/j.semcdb.2009.04.005>
PMID:19427247
 95. Oviedo NJ, Pearson BJ, Levin M, Sánchez Alvarado A. Planarian PTEN homologs regulate stem cells and regeneration through TOR signaling. *Dis Model Mech*. 2008; 1:131–43.
<https://doi.org/10.1242/dmm.000117> PMID:19048075
 96. Pearson BJ, Sánchez Alvarado A. A planarian p53 homolog regulates proliferation and self-renewal in adult stem cell lineages. *Development*. 2010; 137:213–21.
<https://doi.org/10.1242/dev.044297>
PMID:20040488
 97. Peiris TH, Ramirez D, Barghouth PG, Oviedo NJ. The Akt signaling pathway is required for tissue maintenance and regeneration in planarians. *BMC Dev Biol*. 2016; 16:7.
<https://doi.org/10.1186/s12861-016-0107-z>
PMID:27068018
 98. Zhu SJ, Pearson BJ. The Retinoblastoma pathway regulates stem cell proliferation in freshwater planarians. *Dev Biol*. 2013; 373:442–52.
<https://doi.org/10.1016/j.ydbio.2012.10.025>
PMID:23123964
 99. Pearson BJ, Sánchez Alvarado A. Regeneration, stem cells, and the evolution of tumor suppression. *Cold Spring Harb Symp Quant Biol*. 2008; 73:565–72.
<https://doi.org/10.1101/sqb.2008.73.045>
PMID:19150962
 100. Schaeffer DJ. Planarians as a model system for *in vivo* tumorigenesis studies. *Ecotoxicol Environ Saf*. 1993; 25:1–18.
<https://doi.org/10.1006/eesa.1993.1001>
PMID:7682912
 101. Hall F, Morita M, Best JB. Neoplastic transformation in the planarian: I. Cocarcinogenesis and histopathology. *J Exp Zool*. 1986; 240:211–27.
<https://doi.org/10.1002/jez.1402400209>
PMID:3794621
 102. Compton ZT, Mellon W, Harris VK, Rupp S, Mallo D, Kapsetaki SE, Wilmot M, Kennington R, Noble K, Baciuc C, Ramirez LN, Peraza A, Martins B, et al. Cancer Prevalence across Vertebrates. *Cancer Discov*. 2025; 15:227–44.
<https://doi.org/10.1158/2159-8290.CD-24-0573>
PMID:39445720
 103. Kapsetaki SE, Compton ZT, Dolan J, Harris VK, Mellon W, Rupp SM, Duke EG, Harrison TM, Aksoy S, Giraudeau M, Vincze O, McGraw KJ, Aktipis A, et al. Life history traits and cancer prevalence in birds. *Evol Med Public Health*. 2024; 12:105–16.
<https://doi.org/10.1093/emph/eoae011>
PMID:39099847
 104. Butler G, Baker J, Amend SR, Pienta KJ, Venditti C. No evidence for Peto's paradox in terrestrial vertebrates. *Proc Natl Acad Sci USA*. 2025; 122:e2422861122.
<https://doi.org/10.1073/pnas.2422861122>
PMID:39993196
 105. Boddy AM, Abegglen LM, Pessier AP, Aktipis A, Schiffrman JD, Maley CC, Witte C. Lifetime cancer prevalence and life history traits in mammals. *Evol Med Public Health*. 2020; 2020:187–95.
<https://doi.org/10.1093/emph/eoaa015>
PMID:33209304
 106. Vincze O, Colchero F, Lemaître JF, Conde DA, Pavard S, Bieuville M, Urrutia AO, Ujvari B, Boddy AM, Maley CC, Thomas F, Giraudeau M. Cancer risk across

- mammals. *Nature*. 2022; 601:263–7.
<https://doi.org/10.1038/s41586-021-04224-5>
 PMID:[34937938](#)
107. Crimmins EM. Lifespan and Healthspan: Past, Present, and Promise. *Gerontologist*. 2015; 55:901–11.
<https://doi.org/10.1093/geront/gnv130>
 PMID:[26561272](#)
 108. Drake JC, Yan Z. Targeting healthspan to optimally combat non-communicable disease in an aging world. *Sports Med Health Sci*. 2019; 1:59–60.
<https://doi.org/10.1016/j.smhs.2019.08.005>
 PMID:[35782460](#)
 109. Yoshida K, Fujisawa T, Hwang JS, Ikeo K, Gojobori T. Degeneration after sexual differentiation in hydra and its relevance to the evolution of aging. *Gene*. 2006; 385:64–70.
<https://doi.org/10.1016/j.gene.2006.06.031>
 PMID:[17011141](#)
 110. Tomczyk S, Suknovic N, Schenkelaars Q, Wenger Y, Ekundayo K, Buzgariu W, Bauer C, Fischer K, Austad S, Galliot B. Deficient autophagy in epithelial stem cells drives aging in the freshwater cnidarian Hydra. *Development*. 2020; 147:dev177840.
<https://doi.org/10.1242/dev.177840>
 PMID:[31862842](#)
 111. Wanamaker AD Jr, Heinemeier J, Scourse JD, Richardson CA, Butler PG, Eiriksson J, Knudsen KL. Very long-lived mollusks confirm 17th century AD tephra-based radiocarbon reservoir ages for north Icelandic shelf waters. *Radiocarbon*. Cambridge University Press (CUP); 2008; 50:399–412.
<https://doi.org/10.1017/S0033822200053510>
 112. Barbalho SM, Direito R, Laurindo LF, Marton LT, Guiguer EL, Goulart RA, Tofano RJ, Carvalho AC, Flato UA, Capelluppi Tofano VA, Detregiachi CR, Bueno PC, Girio RS, Araújo AC. Ginkgo biloba in the Aging Process: A Narrative Review. *Antioxidants* (Basel). 2022; 11:525.
<https://doi.org/10.3390/antiox11030525>
 PMID:[35326176](#)
 113. Morán-Santibañez K, Vasquez AH, Varela-Ramirez A, Henderson V, Sweeney J, Odero-Marah V, Fenelon K, Skouta R. Larrea tridentata Extract Mitigates Oxidative Stress-Induced Cytotoxicity in Human Neuroblastoma SH-SY5Y Cells. *Antioxidants* (Basel). 2019; 8:427.
<https://doi.org/10.3390/antiox8100427>
 PMID:[31557847](#)
 114. Calabrò A, Aiello A, Silva P, Caruso C, Candore G, Accardi G. Geroprotective applications of oleuropein and hydroxytyrosol through the hallmarks of ageing. *Geroscience*. 2026; 48:4621–47.
<https://doi.org/10.1007/s11357-025-01697-4>
 PMID:[40425998](#)
 115. Nediani C, Ruzzolini J, Romani A, Calorini L. Oleuropein, a Bioactive Compound from Olea europaea L., as a Potential Preventive and Therapeutic Agent in Non-Communicable Diseases. *Antioxidants* (Basel). 2019; 8:578.
<https://doi.org/10.3390/antiox8120578>
 PMID:[31766676](#)
 116. Preston AJ, Rogers A, Sharp M, Mitchell G, Toruno C, Barney BB, Donovan LN, Bly J, Kennington R, Payne E, Iovino A, Furukawa G, Robinson R, et al. Elephant TP53-RETROGENE 9 induces transcription-independent apoptosis at the mitochondria. *Cell Death Discov*. 2023; 9:66.
<https://doi.org/10.1038/s41420-023-01348-7>
 PMID:[36797268](#)
 117. Moody ER, Álvarez-Carretero S, Mahendrarajah TA, Clark JW, Betts HC, Dombrowski N, Szánthó LL, Boyle RA, Daines S, Chen X, Lane N, Yang Z, Shields GA, et al. The nature of the last universal common ancestor and its impact on the early Earth system. *Nat Ecol Evol*. 2024; 8:1654–66.
<https://doi.org/10.1038/s41559-024-02461-1>
 PMID:[38997462](#)
 118. Metabolism, genome and age of the last universal common ancestor. *Nat Ecol Evol*. 2024; 8:1573–4.
<https://doi.org/10.1038/s41559-024-02474-w>
 PMID:[38997463](#)